

Standard Paper

A revision of the *Rostania occultata* (Collemaataceae) complex in Fennoscandia

Alica Košuthová¹ , Martin Westberg² and Mats Wedin¹ 

¹Department of Botany, Swedish Museum of Natural History, P.O. Box 50007, SE-104 05, Stockholm, Sweden and ²Museum of Evolution, Uppsala University, Norbyvägen 16, SE-752 36, Uppsala, Sweden

Abstract

The *Rostania occultata* species complex ('*Collema occultatum* s. lat.') is revised in Fennoscandia and found to consist of four species, all epiphytes on deciduous trees: *Rostania effusa* A. Košuth., M. Westb. & Wedin sp. nov., *R. occultata* (Bagl.) Otálora *et al.*, *R. pallida* A. Košuth., M. Westb. & Wedin sp. nov. and *R. populina* (Th. Fr.) A. Košuth., M. Westb. & Wedin comb. nov. *Rostania effusa* and *R. pallida* are newly described from humid habitats in old-growth boreal coniferous forests, usually with a mixture of deciduous trees, and from similar areas in the subalpine birch-dominated forests of Fennoscandia. *Rostania effusa* is characterized by apothecia with red-brown apothecium discs and an excipulum thallinum with a simple pseudocortex and cubic to oblong, muriform spores. *Rostania pallida* has apothecia with whitish to pale yellowish discs and an excipulum thallinum with a distinct cellular pseudocortex, and ellipsoid, muriform mature spores that are often constricted at the centre. A lectotype is designated for *Collema quadratum* J. Lahm ex Körb. The new combination *Rostania populina* is introduced for the species recognized until now as the variety *Rostania occultata* var. *populina* (Th. Fr.) Perlmutter & Rivas Plata. A key to the six species in *Rostania* s. str. is included.

Key words: cyanolichens, morphology, new species, *Peltigerales*, Scandinavia, systematics, taxonomy

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Introduction

Rostania Trevis. (*Collemaataceae*, *Peltigerales*) is a genus of cyanolichens which corresponds to the informal 'Occultatum' group in *Collema* as circumscribed by Degelius (1954, 1974). *Rostania* was resurrected by Otálora *et al.* (2013a, b) following major phylogenetic studies of the whole family, and studied further by Košuthová and co-workers, who refined the generic delimitation (Košuthová *et al.* 2019). *Rostania* is, within *Collemaataceae*, characterized by a comparatively small crustose to subfruticulose thallus lacking rhizines. The apothecia are initially immersed, later becoming sessile, with a pale whitish or yellowish to red-brown disc that is concave when young, plane when older but never convex. The spores are muriform with at least five cells, cuboid to oblong or ellipsoid (Košuthová *et al.* 2019; Cannon *et al.* 2020). Košuthová *et al.* (2019) excluded three species from *Rostania* and transferred them to other genera as *Collema coccophyllum* Nyl., *Leptogium paramense* (P. M. Jørg. & Palice) A. Košuth. & Wedin, and *Scytinium quadrifidum* (D. F. Stone & McCune) A. Košuth. & Wedin. The position of *Rostania callibotrys* (Tuck.) Otálora *et al.* and *R. laevispora* (Swinscow & Krog) Otálora *et al.* in *Rostania* was uncertain. In the restricted sense, Košuthová *et al.* (2019) accepted three species (*R. ceranisca* (Nyl.) Otálora *et al.*, *R. multipunctata* (Degel.)

Otálora *et al.* and *R. occultata* (Bagl.) Otálora *et al.*). *Rostania ceranisca* and *R. multipunctata* have been well delimited morphologically since Degelius (1954), where the arctic-alpine *R. ceranisca* is characterized by its terricolous habit, a lobate thallus with accessory teretiform lobules, and large oblong spores. *Rostania multipunctata*, a mainly Mediterranean epiphytic species, is distinguished primarily by its large, more or less pulvinate and distinctly lobate thallus. *Rostania occultata*, a small, morphologically quite variable epiphytic taxon with more or less cuboid spores, is currently divided into two, somewhat vaguely delimited varieties (Degelius 1954; Perlmutter & Rivas Plata 2018). The genetic and morphological variation in *R. occultata* and the previously unnoticed variation in photobiont morphology spurred Košuthová *et al.* (2020) to conduct a species delimitation study of *Rostania* s. str. using two coalescent-based methods. They concluded that neither of the two varieties of *R. occultata* corresponded to monophyletic species, but that *R. occultata* consisted of at least four species in Europe, bringing the number of species in *Rostania* s. str. to at least six in this region. Here, we will clarify the taxonomy and nomenclature of the *R. occultata* complex, including the formal description of two new species, and present a key to the currently accepted species of *Rostania* s. str.

Materials and Methods

The study is based mainly on material collected by the authors in Sweden and complemented by additional herbarium material

Corresponding author: Alica Košuthová. E-mail: kosuthova.alica@gmail.com

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from the herbaria S and UPS (abbreviations according to Thiers (2021)), primarily Fennoscandian material. Our own collections are also housed in S and UPS. Specimens were initially studied using a dissecting microscope. Anatomical features were studied using a light microscope on thin sections cut with a razor blade or squash preparations mounted in water. Measurements of mature spores outside of the asci were made in water, under $\times 100$ magnification using oil immersion with a precision of $0.5\ \mu\text{m}$, or from calibrated digital photographs using NIS-Elements (Nikon, Japan) with a precision of $0.1\ \mu\text{m}$. All microscopic images were taken on sections mounted in water unless otherwise stated. Spore measurements are presented in the format: (minimum value observed–) range including 80% of the observed values (–maximum value observed), with the mean of all observed values in the centre and italicized, and n being the number of measurements. Chemical reactions were observed using KOH (10% KOH) and Lugol's iodine (1% of I) without pretreatment of KOH. A list of selected representative specimens is cited under each species. Full lists of specimens examined in this study with DNA voucher codes and GenBank Accession numbers for newly generated sequences are given in the Supplementary Material (available online).

Results and Discussion

In our previous study (Košuthová et al. 2020), we concluded through species delimitation analyses that the *Rostania occultata* complex consists of a minimum of four species, which we informally called *R. occultata* 1, *R. occultata* 2, *R. occultata* 3 and *R. occultata* 4,5,6. We tested and evaluated further potential candidate species within these informal species but found no distinct morphology, substratum ecology or geographical distribution to suggest that these genetically comparatively distinct but poorly sampled candidates should be treated as species. The four accepted species are treated here as *Rostania effusa* ('*R. occultata* 2'), *R. occultata* ('*R. occultata* 4,5,6'), *R. pallida* ('*R. occultata* 1') and *R. populina* ('*R. occultata* 3').

Degelius' concept of dividing *Rostania occultata* into two varieties (Degelius 1954), which has often been followed in the literature, does not correspond in a clear way to the species presented here. There were apparently few specimens of *R. pallida* and *R. effusa* available to Degelius at that time and he appears to have used the name var. *juniperinum* in the herbarium for some of the material, but later abandoned that taxon and included them in var. *occultatum* in his monograph. Specimens of *R. populina* were consistently identified as var. *populinum* while specimens that we recognize as *R. occultata* were identified by Degelius either as var. *occultatum* or var. *populinum*.

In Sweden, *Rostania occultata* s. lat. is currently red-listed in the near-threatened (NT) category (SLU Artdatabanken 2020). Our study has not focused on conducting re-inventories of old localities, and we are not able to comment on trends in the abundance or distribution of any of the accepted species. We note, however, that several depend on old-growth, humid forest ecosystems and that recently these have been much affected by forestry activities throughout Fennoscandia and certainly elsewhere. Future red-listing assessments need to consider these four species separately, and appreciate that old reports of the two varieties of *Rostania occultata* cannot be assigned to these segregated species without studying voucher material. All *Rostania* species, particularly the two newly described here, are under-collected and should be actively searched for.

Observations regarding some characters

Thallus structure. The morphological thallus structure varies from granular in *Rostania effusa* (Fig. 1A–D), *R. occultata* (Fig. 2A & B) and *R. pallida* (Fig. 3A & B) to small foliose to subfruticulose in *R. populina* (Fig. 4A & B). The granules in *R. effusa* and *R. pallida* are more or less rounded, sometimes coalescing into larger aggregates in *R. effusa*, and small (up to $100\ \mu\text{m}$ wide) compared to those in *R. occultata*, which can become up to $200\ \mu\text{m}$ wide and sometimes form minute, lobate squamules often conglomerated in rosette-like structures. *Rostania populina* is the only species where the thallus is usually formed by lobes up to $0.5\ \text{mm}$ long and to $250\ \mu\text{m}$ thick with swollen ends, often forming small rosettes, with the lobes frequently lobulated (Fig. 4A & B).

Colour of the thallus and apothecium disc. The thallus colour in most *Rostania* species has the typical greenish to brownish tones found in *Collema* s. lat. Specimens of *R. occultata* and *R. populina* are sometimes almost black. In *Rostania pallida*, however, the thallus is usually pale grey, which separates it from the other species. Sometimes it becomes darker with brownish and greenish tones. The apothecium disc in *R. pallida* is almost always pale, whitish, greyish or yellowish, rarely pale brown (Fig. 3A & B). The remaining species have the red-brown apothecium disc (Figs 1C, 2A & 4A) that is commonly associated with *Collema* s. str., for example in *C. nigrescens*.

Iodine reactions in the thallus. In the majority of the specimens of *Rostania populina* (8, including the type, of 11 specimens examined) we have observed a slow but distinct I+ magenta to purple reaction to Lugol's solution in the thallus (Fig. 4D). This reaction was discussed by Degelius (1954: p 247) and is apparently due to the presence of glycogen in the gelatinous matrix derived from the photobiont (see Degelius (1954) and references therein). The reaction is common throughout *Collemataceae* and it is also present in *R. multipunctata* and *R. ceranisca*, as well as in *Scytinium fragrans*, a species similar to *R. populina*. The reaction is not constant in *R. populina* but we have never observed it in the other species of the *R. occultata* complex. However, in *R. occultata* s. str., as well as in the species described here as *R. effusa* and *R. pallida*, one can often find single *Nostoc* cells (these are probably heterocysts) surrounded by an I+ purple envelope, c. $1\ \mu\text{m}$ thick. These cells occur scattered in the thallus and excipulum thallinum of the apothecia but the number of such cells varies and they are not always found.

Photobionts. Košuthová et al. (2020) found a correlation between thallus morphology and the *Nostoc* morphotypes; species with granular thalli have *Nostoc* that produce either single cells or cells in small clusters (Fig. 1E), whereas species with lobate thalli have *Nostoc* with cells in short to long chains (Figs 2D & 4C). We have not investigated whether these *Nostoc*-morphotypes constitute distinct species, or if they are the result of the different medullary structure that the granular and lobate thalli produce, but we plan to return to this issue in a future study.

Cortical structures of the excipulum thallinum. Degelius (1954: pp. 46–47) recognized three different types of cortical structure in *Collemataceae*: a true cortex (or eucortex), which is not present in any *Rostania* species, a 'typical' (to follow the terminology of Degelius (1954)) pseudocortex, and what Degelius called a 'primitive' pseudocortex. The eucortex is characterized by one (rarely

more) continuous layer of uniform cells, sharply delimited towards the inner part of the thallus. The ‘typical’ pseudocortex is formed by two or more continuous layers of isodiametric (globose or polygonal) or somewhat oblong cells indistinctly separated from the inner part. A ‘primitive’ pseudocortex is a simple structure that does not have a continuous layer of cells. We refer to this here as a ‘simple’ pseudocortex, to avoid implying that this is something primitive from an evolutionary viewpoint. These distinctions are primarily relevant in the excipulum thallinum because cortical structures are usually poorly developed in *Rostania* thalli. *Rostania effusa* (Fig. 1E), *R. occultata* (Fig. 2C) and *R. populina* all have a very simple pseudocortex. *Rostania pallida* (Fig. 3C) has, at least in the apothecium margin, a more distinct and well-developed cellular pseudocortex with a continuous layer of isodiametric cells that fits Degelius’ description of a typical pseudocortex. In *R. effusa*, a more well-developed structure can sometimes occur on the lower side of the apothecium to form a supporting tissue with a single layer of rounded to ellipsoid cells (Fig. 1F).

Excipulum proprium. Degelius (1954; see also Swinscow & Krog (1986)) described three types of excipulum proprium in *Collema* s. lat.: euthyplechtenchymatous (Fig. 2D), subparaplechtenchymatous and euparaplechtenchymatous (Fig. 4C).

Key to *Rostania* s. str.

For an updated key to the genera in the *Collemataceae*, see Cannon *et al.* (2020).

- 1 Terricolous in arctic-alpine areas; thallus lobate with accessory teretiform lobules, forming dense, intricate cushions; spores 20–40 µm long **R. ceranisca**
Corticolous; thallus granular, squamulose or foliose with flattened lobes; spores < 20 µm long 2
- 2(1) Thallus distinctly foliose, up to c. 3 cm wide **R. multipunctata**
Thallus an effuse, granular to subsquamulose crust, or with small foliose to subfruticulose lobes and then thallus up to c. 2 mm wide 3
- 3(2) Apothecium disc whitish, yellowish or rarely with a pale brown tinge (Fig. 3A & B); mature spores ellipsoid, often constricted at the centre (Fig. 5A & B); excipulum thallinum with a distinctly cellular pseudocortex (Fig. 3C) **R. pallida**
Apothecium disc red-brown; mature spores cuboid to oblong with truncated ends (Fig. 5C & D); excipulum thallinum with a simple pseudocortex without clear cellular structure (Figs 1E & 2C) 4
- 4(3) Thallus with distinct, up to 0.5 mm long lobes raised from the substratum; excipulum proprium below the hymenium euparaplechtenchymatous with rounded cells up to 15 µm wide (Fig. 4C); *Nostoc* mainly in distinct, long chains (up to 20 cells) **R. populina**
Thallus granular or rarely with minute squamules up to 0.2 mm long; excipulum proprium below the hymenium euthyplechtenchymatous with mostly narrow, elongated or sometimes rounded cells up to 8 µm wide (Fig. 2D); *Nostoc* single or in clusters, or in shorter, often indistinct chains (up to 10 cells) 5
- 5(4) Thallus an effuse to crowded crust of small granules, c. 30–75(–100) µm diam.; apothecia mostly sparse; *Nostoc* in single cells or 2–4 cells in small clusters (Fig. 1) **R. effusa**
Thallus granular to subsquamulose, of scattered to crowded granules or forming small cushions, rarely forming small, lobate squamules c. 120–170(–200) µm diam. (Fig. 2B); apothecia often abundant and dominating the thallus; *Nostoc* cells in short chains or when coiled, up to 10 cells long (Fig. 2C & D) **R. occultata**

Taxonomy

Rostania effusa A. Košuth., M. Westb. & Wedin sp. nov.

Mycobank No.: MB 842249

Similar to *Rostania pallida* but differs in the dark green to dark brown, granular thallus, apothecia with reddish brown discs and

The first is characterized by elongated, narrow cells, the second has comparatively short but still elongated cells and the third has isodiametric (globose to polygonal) cells. The differences are not clear-cut and variation occurs to some degree throughout the excipulum in an individual apothecium as the cells change shape and size depending on the position relative to the hymenium. Nevertheless, the structure of the excipulum is useful to distinguish between species; in particular, the euparaplechtenchymatous structure with large cells in *Rostania populina* (Fig. 4C) is a distinctive feature compared to the other three species in the *R. occultata* complex (e.g. Fig. 2D).

Ascospores. The spores in the *Rostania occultata* species complex are often stated to be more or less cuboid (Degelius 1954; Jørgensen 2007; Košuthová *et al.* 2019), but in reality they vary from ellipsoid-oblong to oblong with truncated ends, or almost cuboid (Košuthová *et al.* 2020). They are often not easy to classify since immature spores of all species are first globose and then become ellipsoid to more edged. Mature spores in *R. pallida* (Fig. 5A & B) are ellipsoid, often somewhat constricted at the centre. In the other species, mature spores are usually oblong with truncated ends or cuboid (Fig. 5C & D). When we observed spores in KOH, immature or otherwise untypical spores often appeared more typical.

distinctly cuboid mature spores. Differs from *R. occultata* in the much smaller, up to 100 µm wide thallus granules, and in having a *Nostoc* photobiont that is single-celled or with 2–4 cells in small clusters.

Type: Sweden, Härjedalen, Lillhärda par., Bäreggen, N of Mt Galberget, c. 5.4 km SW of Lillhärda church, on *Populus tremula*, elev. 529 m, 61.82408°N, 13.98514°E, 11 September 2018,

Westberg HH032 (UPS L-929096—holotype; US, PRA—isotypes). DNA voucher (holotype) AL507, GenBank Accession nos: MT705311 (mtSSU), MT906284 (*Mcm7*), MT906220 (*RPB2* part 5–7), MT906257 (*RPB2* part 7–11).

(Fig. 1)

Thallus effuse, of more or less globose granules scattered to contiguous or coalescing and forming a granular crust (Fig. 1A–D); granules c. 30–75(–100) µm wide, dark green to dark brown, ecori-cate or with a very indistinct pseudocortex; *medulla* densely packed with reticulately branched hyphae forming a net around the photobiont. *Photobiont* *Nostoc* with cells (up to 8 µm diam.) singular or in small clusters (2–4 cells usually visible) (Fig. 1E).

Apothecia often sparse or absent, perithecial when young, rarely expanding when mature, up to 0.3 mm wide; disc flat, usually distinctly red-brown, up to 0.15 mm diam. with excipulum proprium normally not visible in surface view; *excipulum thallinum* thin, persistent, up to c. 75 µm wide with hyphal arrangement similar to that of the thallus and with a simple pseudocortex (Fig. 1E); a basal supporting tissue (Henssen 1981) sometimes present, 6.5–8 µm thick, consisting of one layer of continuous isodiametric polygonal cells (Fig. 1F); *excipulum proprium* thin (up to 10 µm wide), euty-plectenchymatous, composed of hyphae with elongated, rectangular cells (up to 15.5 × 3 µm), cells below the hymenium and in the parathecial crown shorter (up to 10 × 3 µm); *hypothecium* colourless, I+ blue, but reaction is often weak, up to 20 µm thick; *hymenium* I+ blue, c. 85–125 µm tall, in the upper part with a reddish to brownish colour; *paraphyses* simple, sometimes anastomosing, up to 1.5 µm wide in mid-hymenium, tips usually 3(–5) µm wide, cylindrical to clavate; *asci* ($n = 6$) sub-clavate to sub-cylindrical, c. 70–110 × 12–30 µm; *ascospores* cubic to oblong with truncated ends when mature (immature spores globose to ellipsoid), muriform with up to 12 visible cells, (8.7–)10.3–13.2–15(–16.3) × (8–)10–10.9–12.5(–14.5) µm ($n = 95$).

Pycnidia not seen.

Etymology. *Effusa* ('effuse') refers to the structure of the thallus, composed of granules spreading, without a distinct margin, into an effuse crust (Fig. 1).

Ecology and distribution. This species occupies a wide range of substrata but in Fennoscandia is most common on *Populus tremula*, *Salix caprea* and *Sorbus aucuparia*. It is also known from *Acer platanoides*, *Alnus incana*, *Betula* spp., *Fraxinus excelsior*, *Picea abies* and *Salix fragilis*. It has been found in many different habitats, mostly in humid and rather open situations such as along forest margins near streams, lakes or bogs, but also on solitary trees in gardens and avenues, in open coniferous forests and in subalpine birch forests. *Rostania effusa* is probably the most common and widespread of the four species in the *R. occultata* complex in the Fennoscandian boreal zone but scattered finds have also been made further south. Outside of Fennoscandia it is so far known only from single collections from Russia, Scotland and Slovenia.

Notes. *Rostania effusa* is the species called '*R. occultata* 2' in Košuthová et al. (2020). It often has a distinct appearance with very small granules, which often appear blastidiate or are similar to the gonocysts that can be found in the *Micarea prasina* group.

It can be mistaken for *Rostania pallida*, which also has a minutely granular thallus. However, *R. effusa* is often sterile and the apothecia, when produced, are very small and often quite indistinct, with a brownish to reddish disc which distinguishes it from the pale, whitish to yellowish, rarely pale brown disc in *R. pallida*. The spores are cubic when mature compared to the oblong to ellipsoid mature spores in *R. pallida*. When the thal-line granules are sparsely produced it can be very similar to *R. occultata*, but the granules are always smaller in *R. effusa* than in *R. occultata*. Furthermore, the *Nostoc* forms singular cells or small clusters in *R. effusa* and not long, coiled chains as in *R. occultata*.

Exsicccates. **Finland:** *Regio kuusamoënsis*: Salla, 1 July 1937, Laurila (Räsänen, *Lich. Fenn. Exs.* 626; UPS L-534969, S F64160). *Tavastia borealis*: Saarijärvi, 23 Aug. 1943, Koskinen (Räsänen, *Lich. Fenn. Exs.* 948; UPS L-535423, S F64158).—**Sweden:** *Södermanland*: Stora Malm par. 15 Sept. 1914, Malme (Malme, *Lich. Suec. Exs.* 468; UPS L-109060, in mixture with *R. occultata*, S F147920). *Värmland*: Södra Finnskoga par. 27 July 1979, Sundell (Vězda, *Lich. Sel. Exs.* 2005; UPS L-004538, S F147910).

Selected specimens examined. **Finland:** *Regio kuusamoënsis*: Kuusamo, 1963, Ahti 18020 (UPS L-609250). *Savonia borealis*: Nilsä, 1959, Henssen 4549 (UPS L-003498). *Tavastia borealis*: Mahlu [Saarijärvi], 1943, Koskinen (S F64158); Saarijärvi, 1944, Koskinen 56 (UPS L-079700).—**Norway:** *Finnmark*: Sør-Varanger, 1957, Th. M. Fries (UPS L-003494). *Nordland*: Vega, 1976, Degelius V-1520 (UPS L-079696).—**Russia:** *Komi Republic*: 2004, Hermansson & Pystina 13548a (UPS L-162655).—**Slovenia:** *Notranjsko-kraska*: Snežnik-Javorniki, 1997, Prügger & Surina (GZU 56-2003, DNA voucher AL485).—**Sweden:** *Dalarna*: Idre par., 2002, Hermansson 11912 (UPS L-656297); Svärdsjö par., 2012, Westberg (S F304739, DNA voucher AL256); Söderbärke par., 1997, Hermansson 8055 (UPS L-088894). *Gästrikland*: Järbo par., 2006, Delin (UPS L-163158). *Hälsingland*: Järvsö par., 2017, Košuthová et al. 103 (S F388772, DNA voucher AL308). *Härjedalen*: Lillhärda par., 2018, Westberg HH053 (UPS L-929106, DNA voucher AL506). *Jämtland*: Krokomb par., 2017, Jonsson FU6545 (S F388734, DNA vouchers AL270 and AL269, GenBank Accession number MW724307); Åre par., 1913, Du Rietz (UPS L-611182, L-616784). *Lule Lappmark*: Jokkmokk par., 1993, Nordin 3335 (UPS L-058678); Liden par., 2017, Westberg URL339 (UPS L-880300). *Närke*: Kvistbro par., Berglund (S F403779). *Pite Lappmark*: Arjeplog par., Westberg PL021, PL035 (S L876859, DNA voucher AL510; S F388749, DNA voucher AL316). *Småland*: Hult par., 2020, Westberg (UPS L-963723, S F403831, DNA voucher AL618, GenBank Accession number MW724312). *Södermanland*: Stora Malm par., 1913, Malme (S L18353). *Uppland*: Åland par., 2017, Westberg, Ekman, Zamora (UPS L-834451, DNA voucher AL244). *Värmland*: Bjurtjärn par., 2017, Berglund 37 (S F388782, DNA voucher AL480). *Västerbotten*: Skellefteå par., 1984, Muhr (UPS L-596495). *Åsele Lappmark*: Dorotea par., 1939, Du Rietz (UPS L-947502, L-947504).—**Great Britain:** *Scotland*: Highland, 2017, Coppins 25250 (S F321109, DNA voucher AL481).

***Rostania occultata* (Bagl.) Otálora, P. M. Jørg. & Wedin**

Fungal Diversity 64, 289 (2013).—*Collema occultatum* Bagl. *Comm. Soc. Crittog. Ital.* 1 (Fasc. 1), 23 (1861); type: [Italy,

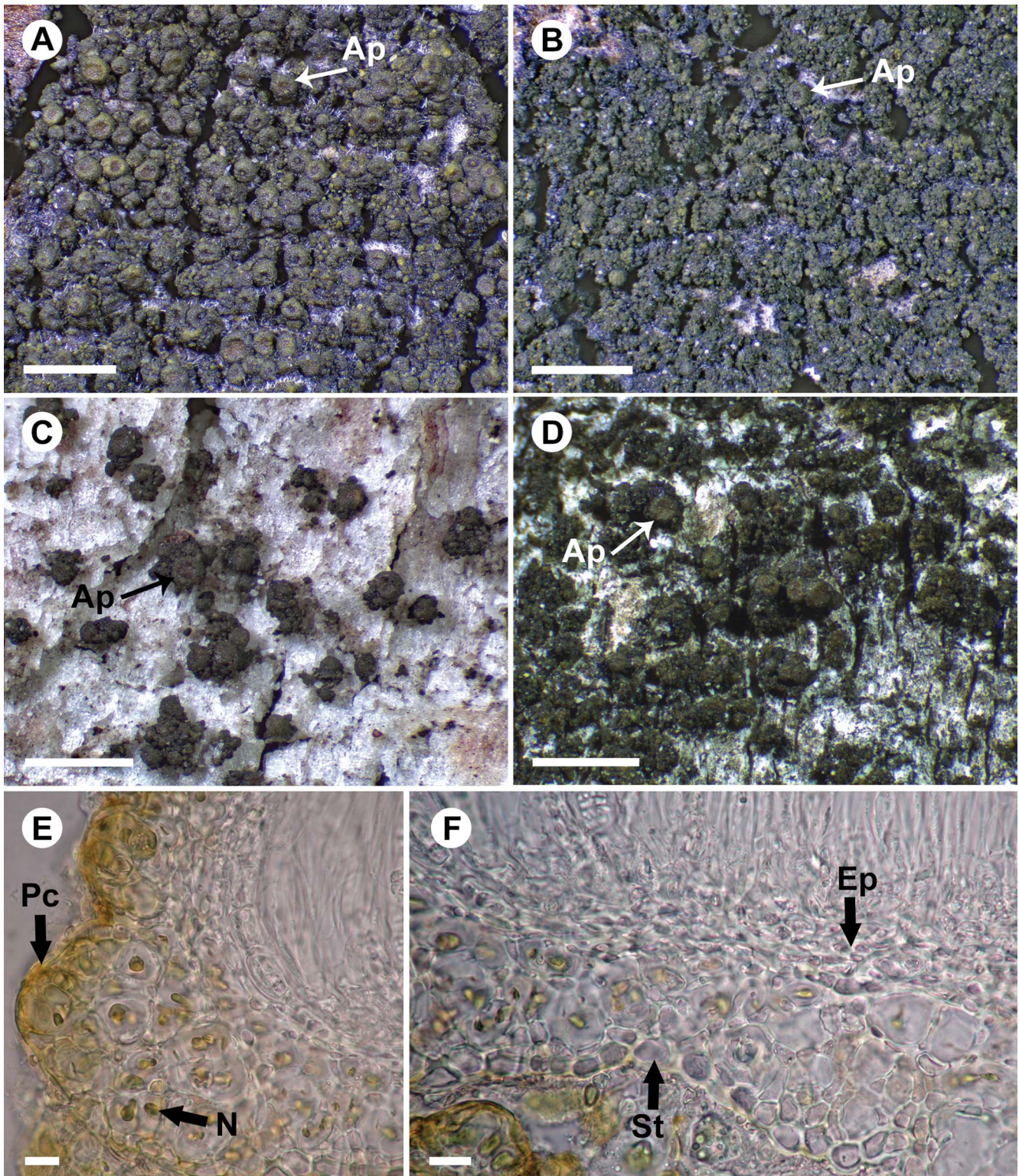


Fig. 1. *Rostania effusa* (A, UPS L-611182; B, UPS L-088894; C, UPS L-656297; D, UPS L-929096—holotype; E & F, S F388734). A, habitus – granular thallus with an unusually large number of apothecia (**Ap**); note the small globose thallus granules. B, habitus – granular thallus dominated by small blastidia-like granules with apothecia (**Ap**). C, habitus – granular thallus forming small, scattered patches with apothecia (**Ap**). D, habitus – granular thallus forming scattered patches with apothecia (**Ap**). E, section of apothecium; note the excipulum thallinum with a simple pseudocortex (**Pc**) and *Nostoc* as single cells or 2–4 cells in clusters (**N**). F, section of apothecium; note the supporting tissue (**St**) formed by a single layer of rounded to ellipsoid cells and the euthyplechtenchymatous excipulum proprium (**Ep**). Scales: A–D = 1 mm; E & F = 10 μ m. In colour online.

Liguria:] ‘Ad castanearum truncos in Alpen. Ligustiae prope Serravalle’. *Pietro Ferrari* (UPS L-074630—lectotype (Degelius 1954: p. 248, 250), S F148057—isolectotype).

Collema coccophylloides Hepp ex. Müll. Arg. *Mém. Soc. Phys. Hist. Nat. Genève* 16(2), 426 (1862); type: Switzerland, Genève, ‘tres-rare, sur un *Populus pyramidalis*, avant d’arriver au Bois de

la Batie, en venant du vieux pont sur l'Arve', Hepp 1860, in hb. Müll. Arg. (G00278941—syntype).

Rostania quadrata (J. Lahm ex Körb.) Trevis., *Rc. Ist. Lomb., Milano*, ser. 2 13, 75 (1880).—*Collema quadratum* J. Lahm ex Körb., *Parerga Lichenol.* (Breslau) 5, 411 (1865); type: Germany, Am Fuße des Burgberges (Ob Crailsheim an Maßholder), 1859, Kemmler (S F148064—lectotype, designated here, MBT 10004691; UPS L-986294, L-986289—isolectotypes).

(Figs 2 & 5C)

Thallus granular to sometimes indistinctly subsquamulose, of widely scattered granules or conglomerates of granules forming small cushions (Fig. 2A & B); granules globose, dark green to almost black, 120–170(–200) µm diam., granular cushions up to c. 2 mm wide, sometimes of minute lobes that can form minute rosette-like thalli, lobes to c. 0.2 mm long, rosettes to c. 1 mm wide; *medulla* composed of a network of reticulately branched hyphae with thin walls, at the surface forming a simple pseudo-cortex as the end cells swell to a rounded, up to 5 µm wide cell or with the hyphae looping around the outermost photobiont cells, interior densely filled with the photobiont. *Photobiont* *Nostoc* in distinct but mostly coiled chains of up to 10 cells (Fig. 2C & D), individual cells 4–7 µm diam.

Apothecia common, scattered to aggregated, sometimes covering the small thallus cushions, perithecial when young; disc expanding to up to 0.14 mm diam., dark red-brown, occasionally poorly pigmented and then pale yellow-brown (Fig. 2E) with excipulum proprium indistinct or visible as a pale, diffusely delimited, often incomplete ring surrounding the disc and with a thin, persistent excipulum thallinum; *excipulum thallinum* similar to the thallus, a simple pseudocortex (Fig. 2C); *excipulum proprium* I–, euthyplechtenchymatous, usually thin below the hymenium, up to c. 10 µm thick, rather compact with small ellipsoid cells, up to c. 8 × 3 µm (Fig. 2D) but mostly smaller, next to the hymenium thin, to c. 10 µm thick but often less, with more narrow and elongated, often more or less rectangular cells, the uppermost cells becoming shorter and rounded, not or slightly expanding at the top to form an up to 15 µm wide parathecial crown; *hypothecium* colourless, I+ blue but the reaction is often weak and sometimes not apparent, variable in thickness, often thin, to c. 15 µm thick but can be up to 50 µm thick; *hymenium* I+ blue, 80–140 µm tall, upper part with a red-brown colour; *paraphyses* to 2 µm wide, tips clavate, to 5 µm wide; *asci* ($n = 3$) subclavate to subcylindrical, c. 60–110 × 8–15 µm; *ascospores* cubic-oblong with truncated ends when mature (immature spores globose to ellipsoid), muriform with (4–)8–12 cells in optical view, (11–)13–14.6–16(–19) × (9.5–)10–11.0–12(–14.5) µm ($n = 54$).

Pycnidia not seen.

Ecology and distribution. In the northern part of its range in Sweden, *Rostania occultata* often grows on *Populus tremula*, but otherwise it is known from a wide range of deciduous trees, such as *Ulmus*, *Fraxinus*, *Acer* and *Malus*. It occurs in a variety of habitats and has been collected in moist deciduous forests, mixed forests and in spruce-dominated forests. It is often found in open situations, for example along gravel roads or on solitary trees around farms. In dense forests it can grow on *Populus tremula* high up on the trunk near the canopy. *Rostania occultata* is distributed throughout southern Sweden, extending to the province of Uppland in the north. It is widely distributed throughout Europe and we have

seen specimens from Austria, Bosnia and Herzegovina, Germany, Greece, Italy, Scotland, Spain and Switzerland.

Nomenclatural notes on *Collema coccophylloides* Hepp ex Müll. Arg. We have studied the original specimen that Degelius (1954: p. 246 and p. 250) mentions, but we likewise refrain from lectotypifying on this rather poor sample.

Nomenclatural notes on *Collema quadratum* J. Lahm ex Körb. In the original description, a number of syntypes (see Supplementary Material, available online) were included, collected by Lahm, Nitschke and Kemmler. Körber distributed material collected by Kemmler in 1860, in the exsiccate *Lich. Sel. Germ.* 269, but the duplicate we have studied in S (F148065) is rather poorly developed. As lectotype we designate the sample in S, F148064, which is a rich and well-developed sample from the same locality as distributed in the exsiccate, but collected by Kemmler a year earlier (in 1859). Kemmler clearly collected this species on several occasions at Crailsheim.

Notes. *Rostania occultata* was informally called 'R. occultata 4,5,6' in our previous species delimitation study (Košuthová et al. 2020), which suggested that it may be composed of three lineages that could represent species. We have, however, not been able to find any distinguishing characters or geographical distribution patterns that could support a further separation and we currently view this clade as a variable and widely distributed species. *Rostania occultata* can be confused with both *R. effusa* and *R. populina*. Compared to *R. effusa* it usually has larger and fewer granules and often has a distinctly different aspect to its habitus, that is somewhat *Collema* s. str.-like with a green thallus and red-brown apothecia, compared to the small and often brownish and/or blastidiate appearance in *R. effusa*. Well-developed specimens with a more squamulose habit can be difficult to separate from small and poorly developed specimens of *R. populina*. *Rostania occultata* often has a less distinct excipulum proprium in external view, but anatomically there are several differences: *R. populina* has a distinct excipulum proprium which is euparaplechtenchymatous in section (Fig. 4C) and a comparatively loose medulla with scattered, long *Nostoc* chains (Fig. 4C). A small number of specimens of *R. occultata*, mainly from continental Europe, are somewhat intermediate between *R. occultata* and *R. populina* (e.g. the type material of *Collema quadratum*) having a subparaplechtenchymatous excipulum proprium below the hymenium and more distinct *Nostoc* chains. The medulla, however, appears less loose and the photobiont densely fills the thallus interior and, on the whole, the appearance is rather different from *R. populina*. In addition, we have never observed a purple iodine reaction of the gelatinous matrix of the thallus interior similar to that present in most specimens of *R. populina* (Fig. 4D).

Exsiccates. **France:** *Grand-Est:* Vosges, Harmand (Claudel, Claudel & Harmand, *Lich. Gall.* 304) (UPS L-987571). *Occitanie:* Aveyron. Marc (Krypt. Exs. (Vindob.) 1542) (UPS L-677687, S F148059); Aveyron. Marc (Zahlbruckner, *Lich. Rar. Exs.* 77) (UPS L-987569).—**Germany:** *Baden-Württemberg:* Crailsheim, Kemmler (Zwack-Holzhausen, *Lich. Exs.* 412) (UPS L-987570, S F148061—syntypes of *Collema quadratum*); Crailsheim, 1861, Kemmler (Körber, *Lich. Sel. Germ.* 269) (S F148065—syntype of *Collema quadratum*).—**Sweden:** *Södermanland:* Stora Malm par., 15 Sept. 1914, Malme (Malme,

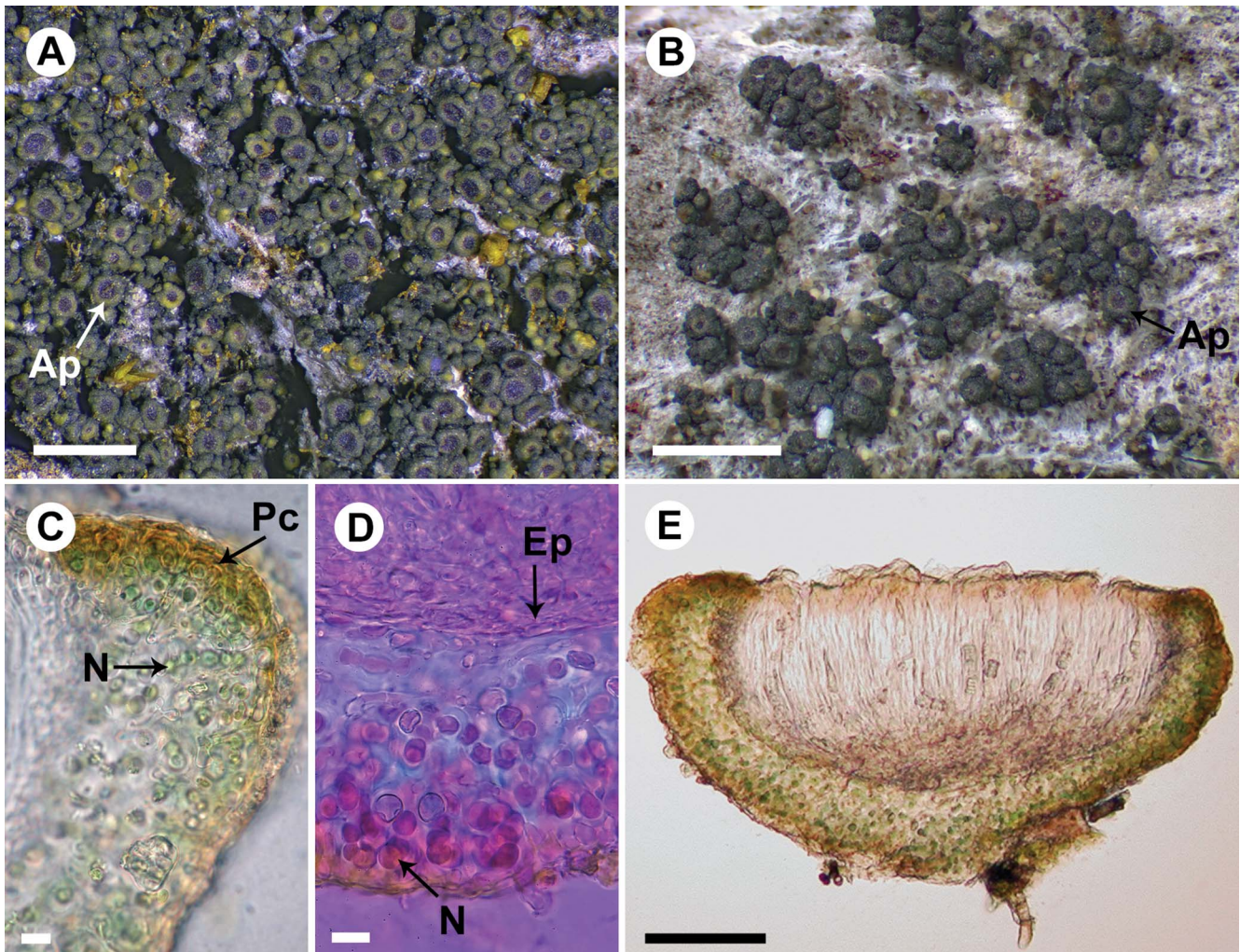


Fig. 2. *Rostania occultata* (A, UPS L-003461; B, S F388790; C & E, UPS L-872297; D, UPS L-123018). A, habitus – granular thallus composed of globose granules and apothecia dominating the thallus (**Ap**). B, habitus – thallus composed of granules and small squamules and apothecia (**Ap**). C, section of apothecium; note the excipulum thallinum with a simple pseudocortex (**Pc**) and *Nostoc* cells in short chains (**N**). D, section of apothecium mounted in Phloxin; note the eutheplechtenchymatous excipulum proprium (**Ep**) below the hymenium and *Nostoc* cells (**N**) with short coiled chains. E, section of apothecium. Scales: A & B = 1 mm; C & D = 10 μ m; E = 50 μ m. In colour online.

Lich. Suec. Exs. 468) (UPS L-109060, in mixture with *Rostania effusa*).

Selected specimens examined. **Austria:** Kärnten: Karawanken, 1985, Hafellner 13607 (GZU P-85). **Niederösterreich:** 1917, Suza (UPS L-973902). **Steiermark:** Hochschwab-Gruppe, 1993, Poelt (GZU 1-93, DNA voucher AL488).—**Bosnia and Herzegovina:** *Republika Srpska:* 2008, Bilovitz 3819 (GZU 58-2012, DNA voucher AL528).—**France:** *Alpes Cote d’Azur:* 2012, Jonsson (S F403821, DNA voucher AL661, GenBank Accession number MW724317). *Pays de la Loire:* 1907, (S F148063).—**Germany:** *Baden-Württemberg:* Kemmler (S F148052—syntype of *Collema quadratum*). *Niedersachsen:* 1930, Erichsen (UPS L-973901). *Nordrhein-Westfalen:* Lahm (UPS L-973903, S F148068—syntypes of *Collema quadratum*).—**Greece:** *Crete:* nomos Chania, 2005, Llop 506060303 (S F233720, DNA voucher AL264).—**Italy:** *Toscana:* passo del’Abetone, 2002, Tretiach (TSB 36983).—**Spain:** *Asturias:* 2018, Westberg (UPS L-933974, DNA voucher AL502).—**Sweden:** *Bohuslän:* Ljung par., 1940, Degelius

(UPS L-079759); Marstrand par., 1920, Magnusson 5186 (UPS L-003461). *Dalsland:* Dalskog par., 1944, Degelius (UPS L-079762). *Halland:* Släp par., 1932, Magnusson 13782 (UPS L-003470, S F18349). *Närke:* Svennevad par., 1872, Björkman (UPS L-003475). *Skåne:* Torekov par., 2008, Hultengren 2365 (UPS L-704977, DNA voucher AL508). *Småland:* Sävsjö par., 2017, Isaksson (S F388790, DNA voucher AL294). *Södermanland:* Tumbo par., 2002, Nordin 5407 (UPS L-120396, DNA voucher MWE87). *Uppland:* Funbo par., 1955, Santesson 11127 (UPS L-123018); Vänge par., Ekman & Westberg 352 (UPS L-872297, DNA voucher AL367). *Västergötland:* Västerlövsta par., 2020, Westberg (UPS L-963072). *Värmland:* Färnebo par., 2010, Odelvik 10528 (S F178903, DNA voucher AL262).—**Switzerland:** *Bern:* Berner Alps, 2006, Hafellner 77470 (GZU 39-2011, DNA voucher AL487). *Jura:* 1996, Groner 13988 (hb. SwissLichens WSL 6793, DNA voucher AL607, GenBank Accession number MW724311). *Zürich:* 1883, Hegetschweiler (S F148060).—**Great Britain:** *Scotland:* Argyll and Bute, 1949, Degelius (UPS L-081088).

***Rostania pallida* A. Košuth., M. Westb. & Wedin sp. nov.**

Mycobank No.: MB 842248

Similar to *Rostania effusa* but differs in the ellipsoid (not cuboid when mature) spores, the grey, granular thallus, and abundant apothecia with pale, white to yellowish discs and a cellular pseudocortex with cells up to 15 µm wide.

Type: Sweden, Hälsingland, Färila par., 3.5 km SW of Kårböle church, just outside Grötvallsskogen Nature Reserve, E of the forest road, wet spruce forest with *Equisetum*, on *Salix caprea*, alt. 250 m., 61.966389°N, 15.257778°E, 15 July 2017, Alica Košuthová, Martin Dinga, Maros Dinga, Mats Wedin 117 (S F388771—holotype; UPS L-987704, US—iso-types). DNA voucher (holotype) AL309, GenBank Accession nos.: MT705303 (mtSSU), MT906250 (RPB2 partial).

Collema byssinum β *juniperinum* Sommerf., *Suppl. Fl. lapp.* (Oslo), 118 (1826); type: Norway, 'Norvegia septentr.: Saltdalen, Nordlandiae, in juniperis, 1822, Sommerfelt' (UPS L-003492—syntype; O L-014065, not seen—syntype).

(Figs 3, 5A & B)

Thallus effuse, granular; granules scattered to contiguous or coalescing and forming small, up to 1.5 mm wide cushion-like structures, spreading over the substratum to form widespread patches, pale grey (usually) or brownish grey or sometimes greenish grey; individual granules c. 35–100 µm wide (mostly below 70 µm) (Fig. 3A), ecorticate or with a very indistinct pseudocortex; *medulla* of densely packed, reticulately branched hyphae forming a net around the photobiont. *Photobiont* *Nostoc* forming singular cells or in small clusters (2–3(–4) cells usually visible).

Apothecia common and usually abundant, globose, perithecial when young; *disc* when mature smooth, flat, whitish, yellowish or rarely with a pale brown tinge, up to 0.25(–0.35) mm diam. (mostly between c. 0.12–0.17 mm) in fully developed apothecia (Fig. 3A & B) with excipulum proprium visible as a distinctly paler (whitish) ring around the disc in surface view (Fig. 3B); *excipulum thallinum* usually thin, up to c. 25–30 µm wide, with a distinct up to 20 µm thick cellular pseudocortex best visible towards the lower surface (Fig. 3C & D), composed of 1–2(–3) cell layers, individual cells rounded to ellipsoid, thin-walled, cell lumina up to 15 µm wide; *excipulum proprium* I–, below the hymenium euthyplectenchymatous forming a dense, up to 10 µm thick layer with narrow hyphae with elongated, rectangular cells similar to those beside the hymenium that become thicker and rounded towards the surface, finally forming an up to 30 µm thick layer in the parathecial crown; *hypothecium* colourless, I+ blue but the reaction is often weak, c. 15–50 µm thick; *hymenium* I+ blue, c. 85–100 µm tall, colourless throughout or upper 5–20 µm yellowish; *paraphyses* simple, sometimes anastomosing, mostly c. 1–1.5 µm wide in mid-hymenium, with cylindrical to clavate, up to 3 µm wide tips, some paraphyses becoming considerably thicker towards the surface and having tips up to 6 µm wide; *asci* subclavate to subcylindrical, c. 60–90 × 15–28 µm (*n* = 9); *ascospores* ellipsoid, often slightly constricted at the centre (Fig. 5B), muriform with 8–12 visible cells (11–)13–14.3–15.5(–20) × (7–)8–9.1–10(–11) µm (*n* = 80).

Pycnidia not seen.

Etymology. *Pallida* ('pale') refers to the pale colour of the thallus and especially the apothecial discs.

Ecology and distribution. The vast majority of collections in Fennoscandia have been made on *Salix caprea* and *Sorbus aucuparia* with only a small number of collections made on other substrata (i.e. *Juniperus communis*, *Salix pentandra* and on a *Betula* stump). It appears to be most common in humid localities, for example along streams and in swampy spruce forests, but also grows in subalpine birch forests up to the tree limit. In very humid localities, it may also occur on thin twigs of *Picea abies*. It has the most pronounced northern distribution of the species in the *Rostania occultata* complex and occurs throughout the boreal zone and the subalpine birch forests in Fennoscandia. The most southern locality known in Sweden lies in the province of Värmland. Otherwise, it is known from Russia (Komi Republic), where it also grows on *Populus tremula*, and from a single find in the USA (Alaska) on *Alnus* sp. Other material belonging to the *Rostania occultata* complex reported from Alaska by McCune et al. (2020) was not studied.

Notes. This species was called '*Rostania occultata* 1' by Košuthová et al. (2020) and is usually easily recognized by its small and granular, grey thallus and abundant apothecia with pale discs. However, the colour varies somewhat and the thallus can be very similar to *R. effusa* which sometimes lacks the otherwise typical distinctly red-brown disc. The ellipsoid spores being slightly constricted at the centre is a unique character for *R. pallida* (Fig. 5A & B). The presence of distinct cuboid spores, or oblong spores with truncated ends, should rule out *R. pallida* (Fig. 5A–D). However, spore shape can unfortunately be difficult to observe since mature spores are often lacking. In apothecial sections, the well-developed cellular pseudocortex with large, isodiametric cells (Fig. 3C & D) is another character distinguishing *R. pallida*.

Exsiccates. **Finland:** *Tavastia australis*: Hollola, 1874, Lang (Norrlin, *Herb. Lich. Fenn.* 153) (UPS L-984886); Jämsä, 13 Apr. 1938, Koskinen (Räsänen, *Lich. Fenn. Exs.* 627) (UPS L-534970, S F64162); Saarijärvi, 21 Aug. 1947, Koskinen (Räsänen, *Lichenoth. Fenn.* 332) (UPS L-175740, S F64156).

Selected specimens examined. **Finland:** *Karelia borealis*: Juuka, 1954, Degelius (UPS L-079701). *Savonia borealis*: Kuopio, 1954, Degelius (UPS L-079702). *Tavastia australis*: Evo [Hämeenlinna], 1873, Norrlin (UPS L-003499, S F64155).—**Norway:** *Finnmark*: Sør-Varanger, 1864, Fries (UPS L-003495). *Nordland*: Grane, 2013, Tønsberg 44463 (BG L100120, DNA voucher AL491); Rana, 2013, Tønsberg 43237 (BG L97889, DNA voucher AL490).—**Russia:** *Komi Republic*: 2007, Hermansson 15468 (UPS L-933975, DNA voucher AL503).—**Sweden:** *Dalarna*: Särna par., 2015, Hermansson 19593 (UPS L-767676, DNA voucher AL377). *Hälsingland*: Ramsjö par., 2004, Berglund (S F317571). *Härjedalen*: Ljungdalen par., 2004, Odelvik 04811 (S F57382, DNA voucher AL263). *Jämtland*: Krokom par., 2017, Jonsson FU6541 (S F388719, DNA vouchers AL268 and AL267, GenBank Accession number MW724306); Lit par., 2017, Košuthová 173 (S F388769, DNA voucher AL302); Kall par., 2020, Westberg JMT 127, 158 (UPS L-980331, L-980332). *Lule Lappmark*: Gällivare par., 2010, Nordin 7094 (UPS L-520806, DNA voucher AL266). *Lycksele Lappmark*: Tärna par., 1967, Sundell 5897 (UPS L-012109). *Medelpad*: Sättna par., Jonsson (S F403820). *Pite Lappmark*: Arjeplog par., 2017, Westberg & Hedenäs PL460 (UPS L-974037). *Torne Lappmark*: Jukkasjärvi par., 1827, Sylvén (S F148023). *Värmland*: Dalby par., 2020, Ekman 5755 (UPS L-989198). *Ångermanland*: Björna par., 1991,

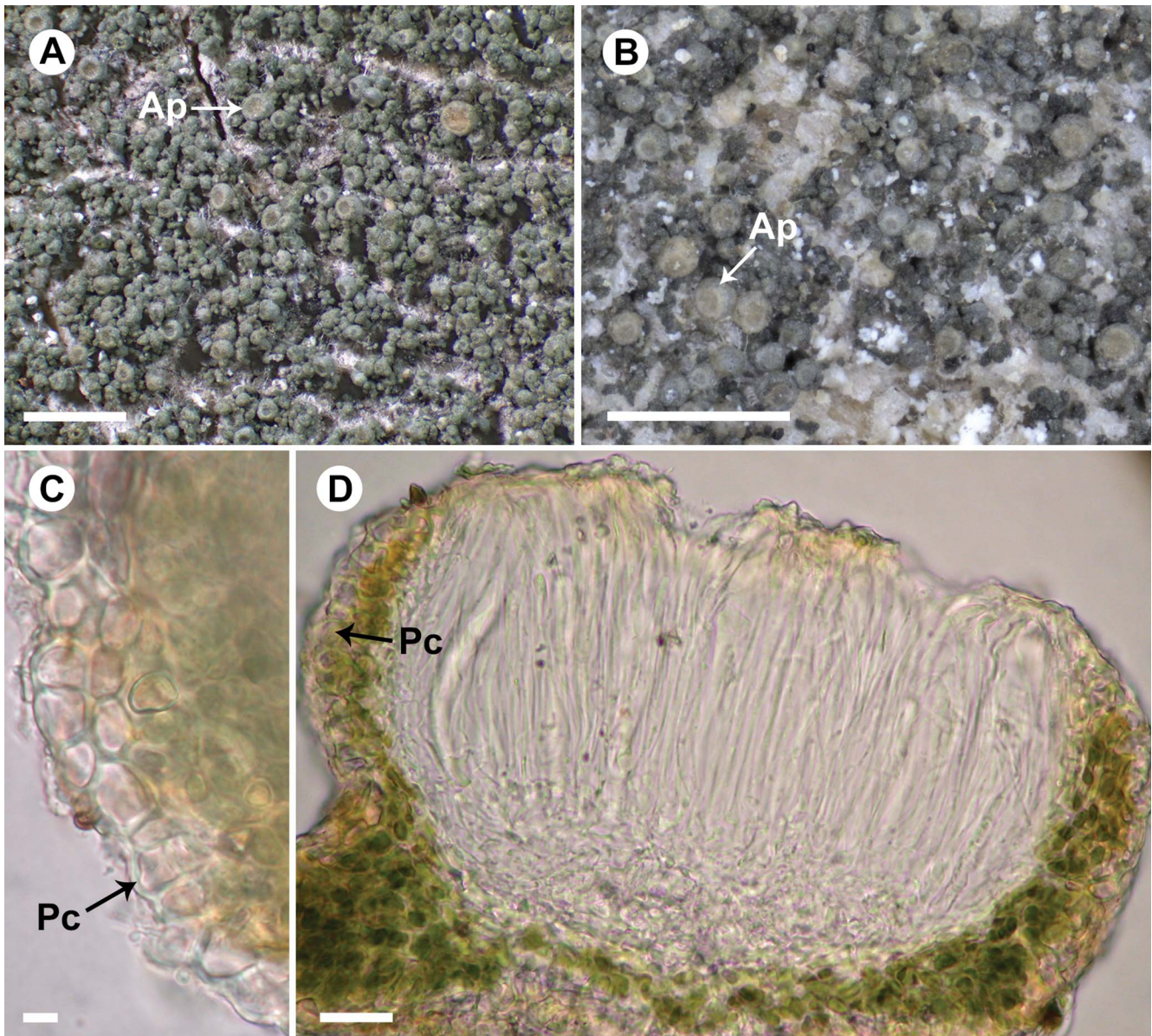


Fig. 3. *Rostania pallida* (A, UPS L-984886; B, S F388771—holotype; C, S F388769; D, UPS L-880093). A, habitus – granular thallus with apothecia (Ap) with the typical pale whitish to brownish disc. B, habitus – granular thallus dominated by the apothecia (Ap) with a pale disc; note the excipulum proprium which is visible at the surface as a whitish ring around the disc. C, section of apothecium margin with an excipulum thallinum which has a pseudocortex (Pc) with rounded cells. D, section of apothecium; note the excipulum thallinum with a pseudocortex (Pc) with rounded cells. Scales: A & B = 1 mm; C = 10 μ m; D = 20 μ m. In colour online.

Hermansson 2767, 2766 (UPS L-125630, L-093928); Tåsjö par., 2017, *Westberg* ULF073 (UPS L-880093, DNA voucher AL242). *Åsele Lappmark*: Vilhelmina par., 1995, *Löfgren* 95-334 (UPS L-147303).—USA: Alaska: Kenai Peninsula Borough, 2015, *Tønberg* 45685 (BG L101159, DNA voucher AL409).

***Rostania populina* (Th. Fr.) A. Košuth., M. Westb. & Wedin comb. nov.**

MycoBank No.: MB 842250

Collema verruciforme b. *populinum* Th. Fr., *Lich. Arct.*, 279 (1860).—*Collema occultatum* var. *populinum* (Th. Fr.) Degel., *Symb. Bot. Upsal.* 13(2), 245 (1954).—*Rostania occultata* var. *populina* (Th. Fr.) Perlmutter & Rivas Plata, *Opuscula Philolichenum*

17, 320 (2018); type: Norway, Nordland, Saltdalen, *Sommerfelt* (UPS L-003493—holotype; O L-14066—isotype).

(Figs 4, 5D)

Thallus (Fig. 4A & B) foliose to subfruticulose, dark green to nearly black, fertile thalli 0.5 to c. 2 mm wide, consisting of small ascending to erect lobes, loosely attached to the substratum, lobes often distinct, up to 0.5 mm long and to 250 μ m thick; lobes entire or somewhat lobulated, often with swollen ends, usually a few arranged in irregular rosettes, sometimes indistinct and crowded and forming pulvinate structures; *medulla* composed of a loose network of reticulately branched hyphae, at the surface forming a thin simple pseudocortex. *Photobiont* *Nostoc* in distinct, long chains (up to 20 cells) (Fig. 4C), cells globose or

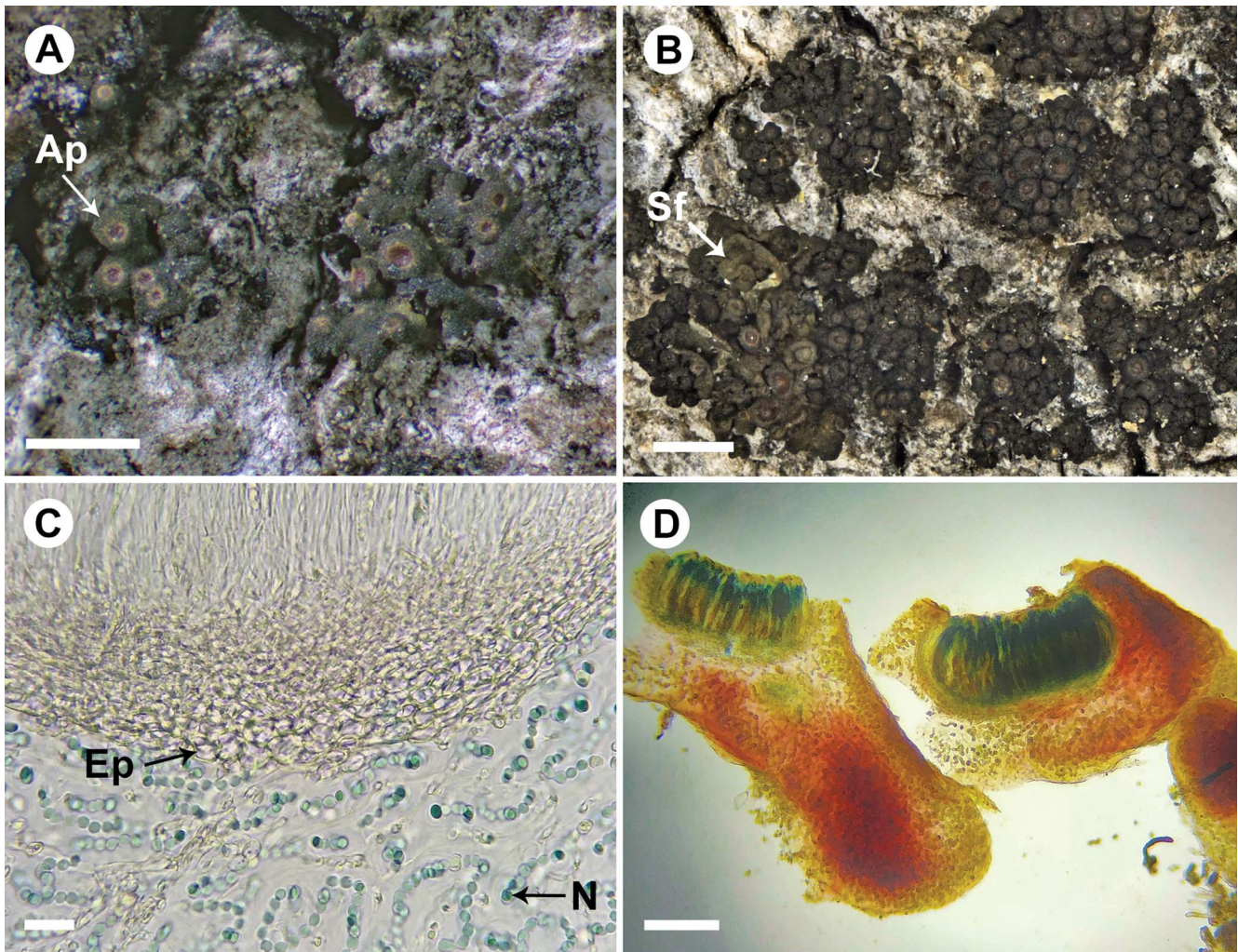


Fig. 4. *Rostania populina* (A, UPS L-125813; B, S F322805; C & D, UPS L-530093). A, habitus – distinctly lobate, subfruticlose thallus with apothecia (**Ap**); note the distinct excipulum proprium visible as a whitish ring around the disc. B, habitus – indistinct lobes covered by apothecia, growing together here with the similar species *Scytinium fragrans* (**Sf**); note the colour difference and the larger, rounded lobes in the latter species. C, section of apothecium; note the euparaplechtenchymatous excipulum proprium (**Ep**) formed by large, ellipsoid to round cells below the hymenium and *Nostoc* cells in long chains (**N**). D, apothecium section treated with Lugol's solution; note the nearly innate habitus of the apothecia and the I+ magenta to purple reaction of the thallus matrix. Scales: A & B = 1 mm; C = 20 μ m; D = 100 μ m. In colour online.

somewhat ellipsoid, 5 \times 5(–6) μ m wide, chains rather widely scattered in the central parts of the thallus and becoming more dense towards the surface, embedded in a gelatinous matrix which is colourless in the central parts and yellow to brownish yellow near the surface. In most specimens this matrix reacts distinctly I+ magenta to purple (Fig. 4D).

Apothecia common, stipitate to nearly innate (Fig. 4D); *disc* flat, dark red-brown, up to 0.22 mm diam. with the excipulum proprium usually visible as a distinct, yellowish white and rather sharply delimited ring surrounding the disc and the excipulum thallinum persistent, smooth to uneven or sometimes minutely lobate in young apothecia still semi-immersed in the lobes; *excipulum thallinum* similar to that of the thallus; *excipulum proprium* I–, below the hymenium to 30 μ m thick, euparaplechtenchymatous, with large, ellipsoid to round cells, variable in size, 3–10 \times 7–15 μ m (Fig. 4C), towards the hymenium the cells often become more elongate to almost rectangular, up to 10 μ m thick next to the hymenium, and towards the surface the cells become smaller and rounded, near the surface usually expanding to form an up to 50 μ m wide parathecial crown; *hypothecium*

colourless, I+ blue but the reaction is often weak, up to 20 μ m thick; *hymenium* I+ blue, 105–180 μ m tall, upper part with a red-brown colour; *paraphyses* 1.5–2 μ m wide, tips clavate, to 4 μ m wide; *asci* ($n = 8$) 70–120 \times 15–25 μ m; *ascospores* cubic-oblong with truncated ends when mature (immature spores globose to ellipsoid), muriform with (4–)8–12 cells in optical view, (12–) 15–16.7–19(–20) \times 10–11.8–13(–13.5) μ m ($n = 33$).

Pycnidia rare, immersed in the thallus; conidia oblong, 3.5–4 \times 1.5 μ m.

Ecology and distribution. This species is so far known only to grow on trunks of *Populus tremula*. It appears to prefer somewhat open habitats and has been recorded in small openings or in slopes in mixed forests, along edge-zones in spruce-dominated forests, for example at the margin of bogs, and in *Populus* successions after forest fires. We currently know this species with certainty only from northern Europe. In Sweden it seems to have a limited distribution and is so far known from 15–20 localities in the northern boreo-nemoral to southern boreal zone in the provinces of Gästrikland, Jämtland, Närke, Södermanland,

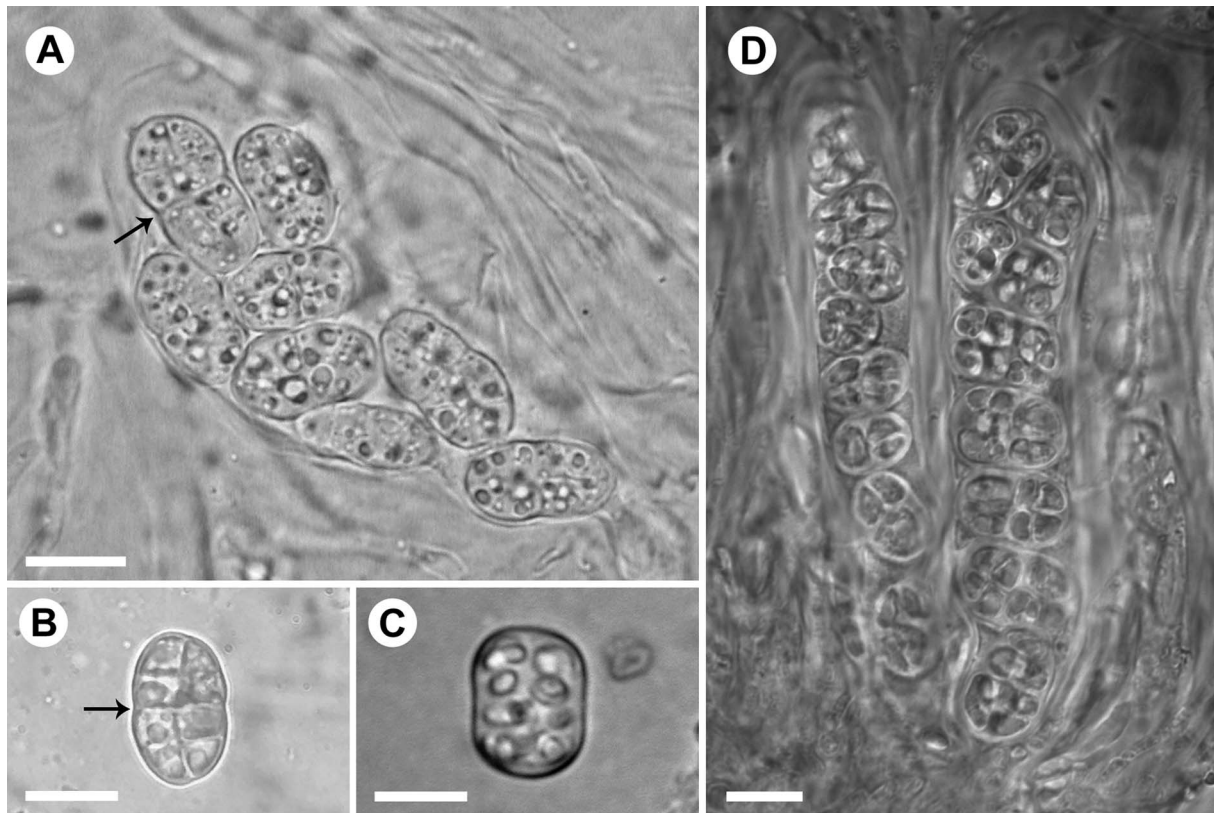


Fig. 5. Asci and spores in the *Rostania occultata* species complex. A, *R. pallida* (S F388719) – ascus with ellipsoid immature spores; note constriction at the centre (arrow). B, *R. pallida* (UPS L-980332) – ellipsoid mature spore; note constriction at the centre (arrow). C, *R. occultata* (UPS L-872297) – oblong mature spore with truncated ends. D, *R. populina* (S F332481) – ascus with oblong spores with truncated ends (right) and ascus with oblong immature spores (left). Scales: A–D = 10 μ m.

Uppland and Västmanland. In Norway, so far we know it only from the type locality in Nordland and in Russia from one locality near Kurkijoki, north of Lake Ladoga. Material reported from North America as *R. occultata* var. *populina* by Perlmutter & Rivas Plata (2018) and McCune *et al.* (2020) was not studied.


Notes. *Rostania populina* was called '*R. occultata* 3' in Košuthová *et al.* (2020). The large size and the lobate thallus make this a distinct species in the *R. occultata* complex but poorly developed thalli can be difficult to separate from *R. occultata*. Typically, the apothecia in *R. populina* have a more distinct excipulum proprium in surface view and in section the loose medulla with scattered, long *Nostoc* chains in combination with a euparaplechtenchymatous excipulum proprium makes it easy to identify. Well-developed specimens can possibly be confused with *Scytinium fragrans*, which is normally considerably larger, with larger, ellipsoid spores.

Exsiccata examined. **Russia:** Karelia: Kurkijoki, 1935, Räsänen (Räsänen, *Lich. Fenn. Exs.* 267; UPS L-533271, S F64161, mixed with *Scytinium fragrans*).

Selected specimens examined. **Sweden:** Gästrikland: Hedesunda par., 2001, Odelvik 01269 (S L42490, DNA voucher AL261). Jämtland: Brunflo par., 2002, Jonsson 4523 (S F322805); Lit par., 2017, Košuthová *et al.* 174 (S F332481, DNA voucher AL300); Mattmar par., 2017, Košuthová 137 (S F388773, DNA voucher AL304). Närke: Götunda par., 1872, Blomberg (UPS L-200588). Södermanland: Julita par., 1888, Malme (S F147914). Uppland:

Vänge par., 1947, Degelius, Hedlund & Ahlner (UPS L-530093); Åland par., 2014, Knutsson (UPS L-689811, DNA voucher AL257). Västmanland: Enåker par., 1991, Hermansson 2614 (UPS L-125813).

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Author ORCIDs.  Alica Košuthová, 0000-0001-5991-7444; Mats Wedin, 0000-0002-8295-5198; Martin Westberg, 0000-0002-8346-0322.

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