



Conference on ‘Transforming the nutrition landscape in Africa’ Plenary Session 1

Feeding the immune system

Philip C. Calder

Human Development and Health Academic Unit, Faculty of Medicine, University of Southampton, Tremona Road,
Southampton SO16 6YD, UK and

National Institute for Health Research, Southampton Biomedical Research Centre, University Hospital Southampton NHS
Foundation Trust and University of Southampton, Tremona Road, Southampton SO16 6YD, UK

A well-functioning immune system is key to providing good defence against pathogenic organisms and to providing tolerance to non-threatening organisms, to food components and to self. The immune system works by providing an exclusion barrier, by identifying and eliminating pathogens and by identifying and tolerating non-threatening sources of antigens, and by maintaining a memory of immunological encounters. The immune system is complex involving many different cell types distributed throughout the body and many different chemical mediators some of which are involved directly in defence while others have a regulatory role. Babies are born with an immature immune system that fully develops in the first few years of life. Immune competence can decline with ageing. The sub-optimal immune competence that occurs early and late in life increases susceptibility to infection. Undernutrition decreases immune defences, making an individual more susceptible to infection. However, the immune response to an infection can itself impair nutritional status and alter body composition. Practically all forms of immunity are affected by protein–energy malnutrition, but non-specific defences and cell-mediated immunity are most severely affected. Micronutrient deficiencies impair immune function. Here, vitamins A, D and E, and Zn, Fe and Se are discussed. The gut-associated lymphoid tissue is especially important in health and well-being because of its close proximity to a large and diverse population of organisms in the gastrointestinal tract and its exposure to food constituents. Certain probiotic bacteria which modify the gut microbiota enhance immune function in laboratory animals and may do so in human subjects.

Lymphocyte: Infection: Cytokine: Nutrient: Gut-associated lymphoid tissue

The aim of this paper is to provide an overview of why good quality nutrition is important for the immune system to function properly and to summarise the evidence available, mainly, though not exclusively, from studies in human subjects, to support this idea. For a broader consideration of the topic the reader is referred to two multi-author books^(1,2), recent textbook chapters^(3,4), earlier comprehensive reviews of the topic^(5–7) and the topic- and nutrient-specific reviews cited within this paper.

The immune system

General overview

The immune system acts to protect the host from infectious agents, including bacteria, viruses, fungi and parasites that exist in the environment and from other noxious insults. It is a complex system involving various cells distributed in many locations throughout the body and moving between these locations in the lymph and the bloodstream. In some

Abbreviations: Th, T-helper.

Corresponding author: Professor P. C. Calder, fax +44 2380 795255, email pcc@soton.ac.uk

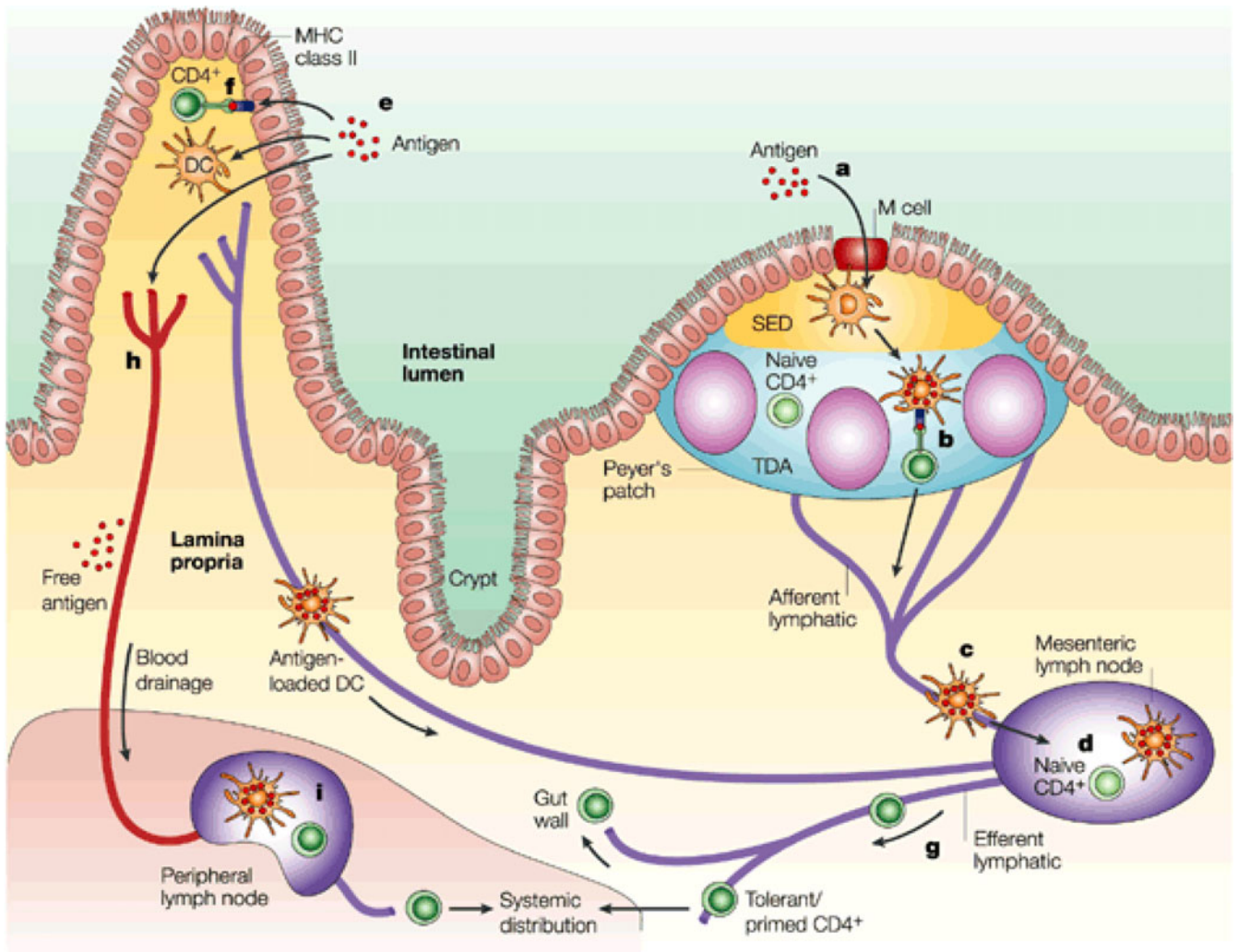


Fig. 1. (Colour online) Structure and organisation of the gut-associated lymphoid tissue. Reprinted by permission from Macmillan Publishers Ltd: *Nat Rev Immunol* 3, 331–341, copyright 2003. Antigen might enter through the microfold (M) cells (a), and after transfer to local dendritic cells (DC), might then be presented directly to T cells in the Peyer's patch (b). Alternatively, antigen or antigen-loaded DC from the Peyer's patch might gain access to draining lymph (c), with subsequent T-cell recognition in the mesenteric lymph nodes (d). A similar process of antigen or antigen-presenting cell dissemination to mesenteric lymph nodes might occur if antigen enters through the epithelium covering the lamina propria (e). In this case, there is also the possibility that enterocytes might act as local antigen presenting cells (f). In all cases, the antigen-responsive CD4⁺ T cells leave the mesenteric lymph nodes in the efferent lymph (g) and after entering the bloodstream through the thoracic duct, exit into the mucosa through vessels in the lamina propria. T cells which have recognised antigen first in the mesenteric lymph node might also disseminate from the bloodstream throughout the peripheral immune system. Antigen might also gain direct access to the bloodstream from the gut (h) and interact with T cells in peripheral lymphoid tissues (i). SED, subepithelial dome; TDA, thymus-dependent area.

locations, the cells are organised into discrete lymphoid organs, classified as primary lymphoid organs where immune cells arise and mature (bone marrow and thymus) and secondary lymphoid organs (lymph nodes, spleen and gut-associated lymphoid tissue) where mature immune cells interact and respond to antigens. The immune system has two general functional divisions: the innate (also called natural) immune system and the acquired (also termed specific or adaptive) immune system. A well functioning immune system is key to providing good defence against

pathogenic organisms and to providing tolerance to non-threatening organisms, to food components and to self. The immune system works by providing an exclusion barrier, by identifying and eliminating pathogens and by identifying and tolerating non-threatening sources of antigens and by maintaining a memory of immunological encounters. Full details of the components of the immune system, their roles and interactions and the chemical mediators involved can be found in any good quality immunology textbook^(8,9).



The gut-associated immune system

The immune system of the gut, often referred to as the gut-associated lymphoid tissue is extensive and includes the physical barrier of the intestinal wall and its mucosal coating as well as components of the innate and adaptive immune systems⁽¹⁰⁾. The physical barrier includes acid in the stomach, mucus and tightly connected epithelial cells, which all act to prevent the entry of pathogens. Within the intestinal wall, cells of the immune system are organised into specialised structures, termed Peyer's patches which are located directly beneath the epithelium in a region called the lamina propria (Fig. 1)⁽¹⁰⁾. This also contains M cells which sample small particles derived from food or from micro-organisms in the gut lumen. The gut-associated immune system not only plays a vital role in providing host defence against pathogens within the gastrointestinal lumen but also in generating tolerogenic responses to harmless micro-organisms and to food components⁽¹¹⁾.

The immune system changes over the life course

Newborn babies have an immature immune system. After birth, immunological competence is gained partly as a result of maturation factors present in breast milk and partly as a result of exposure to antigens (from food and from environmental micro-organisms, the latter starting during the birth process itself)^(12,13). Some of the early encounters with antigens play an important role in ensuring tolerance and a breakdown in this system of 'immune education' can lead to disease^(12,13). At the other end of the lifecycle, older people experience a progressive dysregulation of the immune system, leading to decreased acquired immunity and a greater susceptibility to infection^(14–17). This age-related decline in acquired immunity is termed immunosenescence. An additional consequence of immunosenescence is an impaired response to vaccination^(18,19). Innate immunity appears to be less affected by ageing than acquired immunity.

Why should nutrition affect immune function?

The immune system is functioning at all times, but specific immunity becomes increasingly active in the presence of pathogens. This results in a significant increase in the demand of the immune system for substrates and nutrients to provide a ready source of energy. This demand can be met from exogenous sources (i.e. from the diet) and/or from endogenous pools. Cells of the immune system are able to utilise glucose, amino acids and fatty acids as fuels for energy generation⁽²⁰⁾, which involves electron carriers and a range of coenzymes, which are usually derivatives of vitamins. The final component of the pathway for energy generation (the mitochondrial electron transfer chain) includes electron carriers that have Fe or Cu at their active site. Activation of the immune response induces the production of proteins (including Ig, cytokines, cytokine receptors, adhesion molecules and acute-phase proteins) and lipid-derived mediators (including prostaglandins and leucotrienes). To respond optimally to an immune

challenge there must be appropriate enzymic machinery in place for RNA and protein synthesis and their regulation and ample substrate available (including nucleotides for RNA synthesis, the correct mix of amino acids for protein synthesis and PUFA for eicosanoid synthesis). An important component of the immune response is oxidative burst, during which superoxide anion radicals are produced from oxygen in a reaction linked to the oxidation of glucose. The reactive oxygen species produced can be damaging to host tissues and thus antioxidant protective mechanisms are necessary. Among these are the classic antioxidant vitamins (vitamins E and C), glutathione, the antioxidant enzymes superoxide dismutase and catalase, and the glutathione recycling enzyme glutathione peroxidase. The antioxidant enzymes all have metal ions at their active site (Mn, Cu, Zn, Fe and Se). Cellular proliferation is a key component of the immune response, providing amplification and memory: before division there must be replication of DNA and then of all cellular components (proteins, membranes, intracellular organelles, etc.). In addition to energy, this clearly needs a supply of nucleotides (for DNA and RNA synthesis), amino acids (for protein synthesis), fatty acids, bases and phosphate (for phospholipid synthesis) and other lipids (e.g. cholesterol) and cellular components. Some of the cellular building blocks cannot be synthesised in mammalian cells and must come from the diet (e.g. essential fatty acids, essential amino acids and minerals). Amino acids (e.g. arginine) are precursors for synthesis of polyamines, which play roles in regulation of DNA replication and cell division. Various micronutrients (e.g. Fe, folic, Zn and Mg) are also involved in nucleotide and nucleic acid synthesis. Some nutrients, such as vitamins A and D, and their metabolites are direct regulators of gene expression in immune cells and play a key role in the maturation, differentiation and responsiveness of immune cells. Thus, the roles for nutrients in immune function are many and varied and it is easy to appreciate that an adequate and balanced supply of these is essential if an appropriate immune response is to be mounted. In essence, good nutrition creates an environment in which the immune system is able to respond appropriately to a challenge, irrespective of the nature of the challenge. The response may be an active destructive one, or a more passive tolerogenic one.

Protein–energy malnutrition and immune function

It is well known that undernutrition impairs the immune system, suppressing immune functions that are required for protection against pathogens and increasing susceptibility to infection^(5–7). Undernutrition leading to impairment of immune function can be due to insufficient intake of energy and macronutrients and/or due to deficiencies in specific micronutrients. These may occur in combination. There are a number of reviews of the effect of protein–energy malnutrition on aspects of immune function and on susceptibility to infection^(5–7,21–23). Practically all forms of immunity are affected by protein–energy malnutrition but non-specific defences and cell-mediated immunity are more severely affected than humoral (antibody) responses^(21–23). Barrier function can be impaired by

protein–energy malnutrition^(24,25), which may permit bacterial translocation into the circulation^(24,26). Protein–energy malnutrition causes atrophy of primary and secondary lymphoid organs and there is a decline in the number of circulating lymphocytes, in proportion to the extent of malnutrition^(27,28). The ability of T-lymphocytes to proliferate is decreased by protein–energy malnutrition as in the synthesis of cytokines central to cell-mediated immune response including IL-2 and interferon- γ ^(29,30), suggesting a decline in T-helper (Th)1-type responses. There is a lowered ratio of CD4⁺:CD8⁺ cells in the circulation⁽³¹⁾ and the activity of natural killer cells is diminished^(32–35). Phagocytic capacity of monocytes and macrophages appears to be unaffected^(36,37). The response to a controlled antigenic challenge is reduced by protein–energy malnutrition⁽³⁸⁾, reflecting the effects on individual cellular components. The numbers of B-cells in the circulation and serum Ig levels appear to be unaffected by malnutrition and may even be increased. The functional consequence of malnutrition-induced immune impairment was shown in a study in malnourished Bangladeshi children in which those with the fewest skin reactions to common bacterial antigens (i.e. the weakest cell-mediated immune response) had the greatest risk of developing diarrhoeal disease^(39,40).

The influence of individual micronutrients on immune function

The effects of individual micronutrients on immune function have been identified from studies of deficiency in animals and human subjects and from controlled animal studies in which the nutrient under investigation is included at known levels in the diet. These studies provide good evidence that a number of nutrients are required for an efficient immune response and that deficiency in one or more of them will impair immune function and provide a window of opportunity for pathogens. It seems likely that multiple nutrient deficiencies might have a more significant impact on immune function, and therefore resistance to infection, than a single nutrient deficiency. This section will describe the importance of six selected micronutrients on immune function and susceptibility to infection. These micronutrients have been chosen because each is widely studied and known to be of great importance for immune function and because they are each the focus of much current research activity with significant new discoveries being made.

Vitamin A

There are a number of reviews of the role of vitamin A and its metabolites in the immune system and in host susceptibility to infection^(5–7,41–45). Vitamin A deficiency impairs barrier function, alters immune responses and increases susceptibility to a range of infections^(5–7,41–45). Vitamin A-deficient mice show breakdown of the gut barrier and impaired mucus secretion (due to loss of mucus-producing goblet cells), both of which would facilitate entry of pathogens⁽⁴⁶⁾. Many aspects of innate immunity, in addition to barrier function, are affected by vitamin A^(5–7,41–45).

For example, vitamin A controls neutrophil maturation⁽⁴⁷⁾ and in vitamin A deficiency, blood neutrophil numbers are increased, although their phagocytic function is impaired⁽⁴⁸⁾ resulting in decreased ability to ingest and kill bacteria⁽⁴⁹⁾. Natural killer cell activity is diminished by vitamin A deficiency⁽⁵⁰⁾. The impact of vitamin A on acquired immunity is less clear, but there is some evidence that vitamin A deficiency alters the balance of Th1 and Th2 cells, decreasing Th2 response, without affecting or, in some studies enhancing, Th1 response^(41–45,51). This would suggest that vitamin A will enhance Th1-cell mediated immunity. However, in contrast to this, studies in several experimental models show that vitamin A metabolite retinoic acid decreases Th1-type responses (cytokines, cytokine receptors and the Th1-favouring transcription factor T-bet), while enhancing Th2-type responses (cytokines and the Th2-favouring transcription factor GATA-3)^(52–54). Vitamin A also appears to be important in differentiation of regulatory T-cells while suppressing Th17 differentiation^(55,56), effects which have implications for control of adverse immune reactions. Retinoic acid seems to promote movement of T-cells to the gut-associated lymphoid tissue⁽⁵⁷⁾, and, interestingly, some gut-associated immune cells are able to synthesise retinoic acid^(57,58). Vitamin A deficiency can impair response to vaccination, as discussed elsewhere⁽⁵⁰⁾. In support of this, vitamin A deficient Indonesian children provided with vitamin A showed a higher antibody response to tetanus vaccination than seen in vitamin A deficient children⁽⁵⁹⁾. Vitamin A deficiency is associated with increased morbidity and mortality in children, and appears to predispose to respiratory infections, diarrhoea and severe measles^(5–7,41–45). Replenishment of vitamin A in deficient children improves recovery from infectious diseases and decreases mortality^(5–7,41–45).

Vitamin D

There are a number of reviews of the role of vitamin D and its metabolites in the immune system, autoimmunity and host susceptibility to infection^(60–65). In this paper, vitamin D refers to the active form of vitamin D (1,25-dihydroxy vitamin D₃). Many immune cells express the cytosolic vitamin D receptor and some can synthesise the active form of vitamin D from its precursor^(66,67). These observations suggest that immune cells can both respond to and produce vitamin D indicating that it is likely to have immunoregulatory properties. Indeed, vitamin D can induce macrophages to synthesise anti-microbial peptides^(67,68), directly affecting host defence. Individuals with low vitamin D status have been reported to have a higher risk of respiratory tract viral infections⁽⁶⁹⁾, while supplementation of Japanese school children with vitamin D for 4 months during winter decreased the risk of influenza by about 40%⁽⁷⁰⁾. These studies suggest that vitamin D acts to reduce susceptibility to infection, which may result from improved immune function. However, in contrast, there is a large body of literature showing that vitamin D and its analogues have immunosuppressive effects^(71–73). It seems that under physiological conditions vitamin D probably aids immune responses, but that it may also play an active role in prevention of autoimmunity and that there may

even be a therapeutic role for vitamin D in some immune-mediated diseases. Vitamin D acts by binding to its receptor and regulating gene expression in target cells. Its effects include promotion of phagocytosis, superoxide synthesis and bacterial killing, but it is also reported to inhibit T-cell proliferation and production of Th1-type cytokines^(74–84) and of antibodies by B-cells⁽⁸⁵⁾, highlighting the paradoxical nature of its effects. Effects on Th2-type responses are not clear^(86–88) and there may be an increase in numbers of regulatory T-cells^(89,90). Overall, the current evidence suggests that vitamin D is a regulator of immune function but that its effects will depend upon the immunological situation (e.g. health, infectious disease and autoimmune disease).

Vitamin E

Vitamin E is the major lipid-soluble antioxidant in the body and is required for protection of membrane lipids from peroxidation. Free radicals and lipid peroxidation are immunosuppressive and hence vitamin E should act to maintain or even to enhance the immune response. There are a number of reviews of the role of vitamin E in the immune system and host susceptibility to infection^(91–94). In laboratory animals, vitamin E deficiency decreases lymphocyte proliferation, natural killer cell activity, specific antibody production following vaccination and phagocytosis by neutrophils^(91–94). Vitamin E deficiency also increases susceptibility of animals to infectious pathogens⁽⁹¹⁾. Vitamin E supplementation of the diet of laboratory animals enhances antibody production, lymphocyte proliferation, Th1-type cytokine production, natural killer cell activity and macrophage phagocytosis^(91–94). There is a positive association between plasma vitamin E and cell-mediated immune responses, and a negative association has been demonstrated between plasma vitamin E and the risk of infections in healthy older adults⁽⁹⁵⁾. Vitamin E appears to be of benefit in the elderly^(96–98), with studies demonstrating enhanced Th1 cell-mediated immunity (lymphocyte proliferation and IL-2 production) and improved vaccination responses at fairly high intakes^(96,97). Although some studies report that vitamin E decreases risk of upper respiratory tract infections in the elderly⁽⁹⁹⁾, other studies did not see an effect on the incidence, duration or severity of respiratory infections in elderly populations⁽¹⁰⁰⁾.

Zinc

Zn is important for DNA synthesis, in cellular growth and differentiation, and in antioxidant defence, all important to immune cell function. It is also a cofactor for many enzymes. There are a number of reviews of the role of Zn in the immune system and host susceptibility to infection^(5–7,101–105). Zn deficiency has a marked impact on bone marrow, decreasing the number of precursors to immune cells⁽¹⁰⁶⁾. Zn deficiency impairs many aspects of innate immunity, including phagocytosis, natural killer cell activity and respiratory burst^(107–111). There are also marked effects of Zn deficiency on acquired immunity, with decreases in the circulating number and function of T-cells and an imbalance to favour Th2 cells^(112,113).

Moderate or mild Zn deficiency or experimental Zn deficiency in human subjects decreases natural killer cell activity, lymphocyte proliferation, IL-2 production and cell-mediated immune responses which can all be corrected by Zn repletion^(111,113). In patients with Zn deficiency related to sickle-cell disease, natural killer cell activity is decreased, but Zn supplementation returns this to normal⁽¹¹⁴⁾. The wide ranging impact of Zn deficiency on immune components is an important contributor to increased susceptibility to infection, especially lower respiratory tract infection and diarrhoea, seen in Zn deficiency^(5–7,102–105). Correcting Zn deficiency lowers the likelihood of diarrhoea and of respiratory and skin infections, although some studies fail to show benefit of Zn supplementation in respiratory disease^(5–7,102–105).

Iron

There are a number of reviews of the role of Fe in the immune system and host susceptibility to infection^(115–122). Fe deficiency induces thymus atrophy and has multiple effects on immune function in human subjects^(115–118). The effects are wide ranging and include impairment of respiratory burst and bacterial killing, T-cell proliferation and production of Th1 cytokines^(115–118). However, the relationship between Fe deficiency and susceptibility to infection remains uncertain^(115–122). Indeed, there is evidence that infections caused by organisms that spend part of their life cycle intracellularly, such as plasmodia and mycobacteria, may actually be enhanced by Fe. In children in the tropics, Fe at doses above a particular threshold has been associated with increased risk of malaria and other infections, including pneumonia^(123–126). Thus, Fe intervention in malaria-endemic areas is not advised, particularly high doses in the young, those with compromised immunity (e.g. HIV infection) and during the peak malaria transmission season. Fe treatment for anaemia in a malarious area must be preceded by effective anti-malarial therapy and should be oral. There are different explanations for the detrimental effects of Fe administration on infections. First, Fe overload causes impairment of immune function^(115–118). Second, excess Fe favours damaging inflammation. Third, micro-organisms require Fe and providing it may favour the growth of the pathogen. Perhaps, for the latter reasons, several mechanisms have developed for withholding Fe from a pathogen⁽¹²⁷⁾. Oral Fe supplementation has not been shown to increase risk of infection in non-malarious countries⁽¹¹⁸⁾.

Selenium

Se is a cofactor for a number of enzymes including some involved in antioxidant defences such as glutathione peroxidase. Therefore, Se may protect against the immunosuppressive effects of oxidative stress, thus acting to enhance immune function. There are a number of reviews of the role of Se in the immune system and host susceptibility to infection^(128–132). Se deficiency in laboratory animals affects both innate and acquired immunity and increases susceptibility to infections. Lower Se concentrations in human subjects have also been linked with

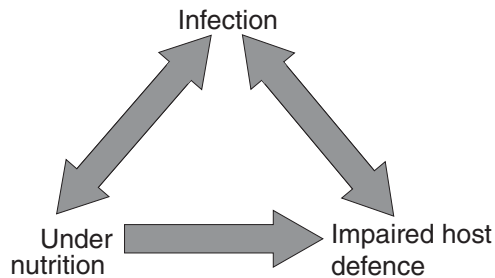


Fig. 2. Schematic depiction of the interrelationship between under-nutrition, impaired immunity and infection.

increased virulence^(131–133), diminished natural killer cell activity^(133,134) and increased mycobacterial disease⁽¹³⁵⁾. Se supplementation has been shown to improve various aspects of immune function in human subjects^(136–138), including in the elderly^(139,140). Se supplementation in Western adults with low Se status improved some aspects of their immune response to a poliovirus vaccine⁽¹⁴¹⁾.

Probiotics, prebiotics, immunity and infection

Indigenous commensal bacteria within the gastrointestinal tract are believed to play a role in host immune defence by creating a barrier against colonisation by pathogens. Disease and the use of antibiotics can disrupt this barrier, creating an environment that favours the growth of pathogenic organisms. There is now evidence that providing exogenous, live, 'desirable' bacteria, termed probiotics, can contribute to maintenance of the host's gastrointestinal barrier. Probiotic organisms are found in fermented foods including traditionally cultured dairy products and some fermented milks and the most commonly used commercial organisms are lactobacilli and bifidobacteria. These organisms are able to colonise the gut temporarily, making their regular consumption necessary. In addition to creating a physical barrier, some of the products of the metabolism of probiotic bacteria, including lactic acid and antibiotic proteins, can directly inhibit the growth of pathogens⁽¹⁴²⁾. Probiotic bacteria also compete with some pathogenic bacteria for available nutrients. In addition, to these direct interactions between commensal and probiotic organisms on the one hand and pathogens on the other, commensal and probiotic organisms can interact with the host's gut epithelium and gut-associated immune tissues⁽¹⁴²⁾. These communications with the host may occur through chemicals released from the bacteria or through direct cell–cell contact⁽¹⁴²⁾ and it is through these interactions that probiotics are thought to be able to influence immune function, even at sites distant from the gut⁽¹⁴³⁾. Nevertheless, the precise nature of these interactions is not very well understood, although there is significant research activity in this area⁽¹⁴⁴⁾. A large number of studies have examined the influence of various probiotic organisms, either alone or in combination, on immune function, infection and inflammatory conditions in human subjects⁽¹⁴⁵⁾. Certain probiotic organisms appear to enhance innate immunity (particularly phagocytosis and natural killer cell activity),

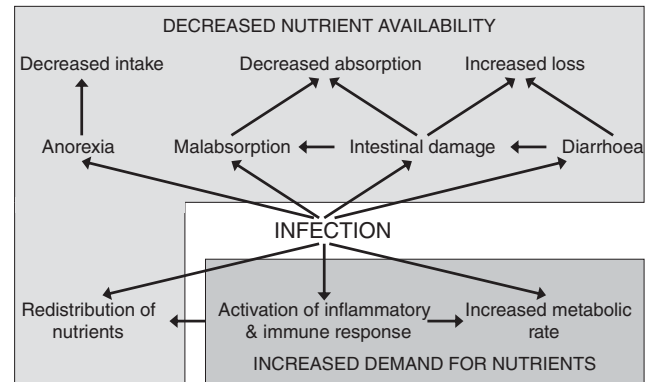


Fig. 3. Schematic depiction of the opposing effects of infection on nutrient availability and nutrient demand.

but they seem to have a less pronounced effect on acquired immunity. A small number of studies show improved vaccination responses in individuals taking probiotics^(146,147), as extensively reviewed recently⁽¹⁴⁸⁾. Some studies in children report lower incidence and duration of diarrhoea with certain probiotics⁽¹⁴⁵⁾. In adults, some studies demonstrate a reduction in the risk of traveller's diarrhoea in subjects taking probiotics⁽¹⁴⁵⁾, while there is now quite good evidence that probiotics protect against antibiotic-associated diarrhoea^(149–153). There are, however, considerable differences in the effects of different probiotic species and strains and effects observed with one type of probiotic cannot be extrapolated to another.

Prebiotics are typically, though not exclusively, carbohydrates which are not digestible by mammalian enzymes but which are selectively fermented by gut microbiota, leading to increased numbers of beneficial bacteria within the gut. Prebiotics include inulin-type fructooligosaccharides, galactooligosaccharides and xylooligosaccharides. The bacteria promoted by prebiotics are often lactobacilli and bifidobacteria. Consequently, prebiotics have the potential to induce the same sorts of immune effects as seen with probiotics, acting through similar mechanisms, although there may also be direct communications between the prebiotics themselves and the host immune cells⁽¹⁵⁴⁾. There is some evidence for immunomodulatory effects of prebiotics, but many experiments conducted in human subjects are difficult to interpret because prebiotics and probiotics are often used in combination⁽¹⁵⁴⁾.

Impact of infection on nutrient status

Although a poor nutritional state impairs immunity and predisposes to infections, the immune response to an infection can itself impair nutritional status and alter body composition^(5,6). Thus, there is a bidirectional interaction between nutrition, infection and immunity (Fig. 2). Infection impairs nutritional status and body composition in the following ways (Fig. 3):

- (1) Infection causes anorexia with reduced food intake ranging from as little as 5% to an almost complete loss of appetite. This can lead to nutrient deficiencies, even



if the host is not deficient before the infection, and may make apparent existing borderline deficiencies.

- (2) Infection can cause nutrient malabsorption and loss, especially infections that damage the intestinal wall or that cause diarrhoea or vomiting⁽¹⁵⁵⁾.
- (3) Infection increases resting energy expenditure, placing a demand on nutrient supply, particularly when coupled with anorexia, diarrhoea and other nutrient losses.
- (4) Infection causes altered metabolism and redistribution of nutrients, including both macronutrients (e.g. amino acids) and micronutrients (e.g. vitamin A, Zn and Fe). A catabolic response occurs with all infections and brings about a redistribution of energy substrates for energy and biosynthesis away from skeletal muscle and adipose tissue towards the host immune system and its supporting tissues including the liver. As a result plasma concentrations of vitamin A, Zn and Fe, among others, decrease with infection.

Anorexia, increased energy expenditure and redistribution of nutrients are brought about by host factors (mainly inflammatory cytokines), while malabsorption and maldigestion are brought about by the pathogen. The result is that an increased nutrient requirement coincides with reduced nutrient intake, reduced nutrient absorption and nutrient losses (Fig. 3).

Summary and conclusions

A well functioning immune system is key to providing good defence against pathogenic organisms and to providing tolerance to non-threatening organisms, to food components and to self. The immune system works by providing an exclusion barrier, by identifying and eliminating pathogens and by identifying and tolerating non-threatening sources of antigens, and by maintaining a memory of immunological encounters. The immune system is complex involving many different cell types distributed throughout the body and many different chemical mediators some of which are involved directly in defence while others have a regulatory role. Babies are born with an immature immune system that fully develops in the first few years of life. This immune maturation requires the presence of specific immune factors and exposure to antigens from food and from micro-organisms. Immune competence can decline with ageing. This process is termed immunosenescence. The sub-optimal immune competence that occurs early and late in life increases susceptibility to infection. Undernutrition impairs immune defences at all stages of the life cycle, although infants and the elderly may be more vulnerable, making an individual more susceptible to infection. However, the immune response to an infection can itself impair nutritional status and alter body composition. Practically all forms of immunity are affected by protein-energy malnutrition, but non-specific defences and cell-mediated immunity are most severely affected. Micronutrient deficiencies impair immune function. The gut-associated lymphoid tissue is especially important in health and well-being because of its close proximity to a large and diverse population of organisms in the

gastrointestinal tract and its exposure to food constituents. Probiotic bacteria which modify the gut microbiota may enhance immune function in human subjects lowering the risk of certain infections and improving the response to vaccination.

Acknowledgements

There is no funding associated with this paper. The author is partly supported by the National Institute for Health Research through the National Institute for Health Research Southampton Biomedical Research Centre.

The author serves on Scientific Advisory Boards of the Danone Research Centre in Specialised Nutrition and Terres-Syral; acts as a consultant to Mead Johnson Nutritionals; has received speaking honoraria from Abbott Nutrition, Nestle, Unilever, Danone and DSM; and currently receives research funding from Terres-Syral.

References

1. Suskind RM & Tontisirin K (2001) *Nutrition, Immunity, and Infection in Infants and Children*. Vevey/Philadelphia: Nestec/Lippincott Williams and Wilkins.
2. Calder PC, Field CJ & Gill HA (2002) *Nutrition and Immune Function*. Wallingford: CAB International.
3. Yaqoob P & Calder PC (2010) The immune and inflammatory systems. In *Nutrition and Metabolism*, 2nd ed., pp. 312–338 [SA Lanham-New, IA Macdonald and HM Roche, editors]. Oxford: Wiley-Blackwell.
4. Calder PC & Yaqoob P (2012) Nutrient regulation of the immune response. In *Present Knowledge in Nutrition*, 10th ed., pp. 688–708 [JW Erdman, IA Macdonald and SHH Zeisel, editors]. Ames: ILSI.
5. Chandra RK (1991) 1990 McCollum Award lecture. Nutrition and immunity: lessons from the past and new insights into the future. *Am J Clin Nutr* **53**, 1087–1101.
6. Scrimshaw NS & SanGiovanni JP (1997) Synergism of nutrition, infection, and immunity: an overview. *Am J Clin Nutr* **66**, 464S–477S.
7. Calder PC & Jackson AA (2000) Undernutrition, infection and immune function. *Nutr Res Rev* **13**, 3–29.
8. Abbas AK, Lichtman AH & Pillai S (2011) *Cellular and Molecular Immunology*, 7th ed., Philadelphia, PA: Elsevier Saunders.
9. Male D, Brostoff J, Roth DB *et al.* (2012) *Immunology*, 8th ed., Philadelphia, PA: Elsevier Saunders.
10. Mowat AM (2003) Anatomical basis of tolerance and immunity to intestinal antigens. *Nat Rev Immunol* **3**, 331–341.
11. Suzuki K, Kawamoto S, Maruya M *et al.* (2010) GALT: organization and dynamics leading to IgA synthesis. *Adv Immunol* **107**, 153–185.
12. Bernt KM & Walker WA (1999) Human milk as a carrier of biochemical messages. *Acta Paed Suppl* **88**, 27–41.
13. Calder PC, Krauss-Etschmann S, de Jong EC *et al.* (2006) Early nutrition and immunity – progress and perspectives. *Br J Nutr* **96**, 774–790.
14. Castle SC (2000) Clinical relevance of age-related immune dysfunction. *Clin Infect Dis* **31**, 578–585.
15. Burns EA & Goodwin JS (2004) Effect of aging on immune function. *J Nutr Health Aging* **8**, 9–18.
16. Agarwal S & Busse PJ (2010) Innate and adaptive immunosenescence. *Ann Allergy Asthma Immunol* **104**, 183–190.

17. Pawelec G, Larbi A & Derhovanessian E (2010) Senescence of the human immune system. *J Comp Pathol* **142**, Suppl. 1, S39–S44.
18. Fulop T, Pawelec G, Castle S *et al.* (2009) Immunosenescence and vaccination in nursing home residents. *Clin Infect Dis* **48**, 443–448.
19. Goodwin K, Viboud C & Simonsen L (2006) Antibody response to influenza vaccination in the elderly: a quantitative review. *Vaccine* **24**, 1159–1169.
20. Calder PC (1995) Fuel utilisation by cells of the immune system. *Proc Nutr Soc* **54**, 65–82.
21. Kuvibidila S, Yu L, Ode D *et al.* (1993) The immune response in protein–energy malnutrition and single nutrient deficiency. In *Nutrition and Immunology*, pp. 121–155 [DM Klurfield, editor]. New York and London: Plenum Press.
22. Woodward B (1998) Protein, calories and immune defences. *Nutr Rev* **56**, S84–S92.
23. Woodward B (2001) The effect of protein–energy malnutrition on immune competence. In *Nutrition, Immunity and Infection in Infants and Children*, pp. 89–120 [RM Suskind and K Tontisirin, editors]. Vevey/Philadelphia: Nestec/Lippincott Williams and Wilkins.
24. Deitch EA, Ma WJ, Ma L *et al.* (1990) Protein malnutrition predisposes to inflammatory-induced gut-origin septic states. *Ann Surg* **211**, 560–567.
25. Sherman P, Forstner J, Roomi N *et al.* (1985) Mucin depletion in the intestine of malnourished rats. *Am J Physiol* **248**, G418–G423.
26. Katayama M, Xu D, Specian RD *et al.* (1997) Role of bacterial adherence and the mucus barrier on bacterial translocation: effects of protein malnutrition and endotoxin in rats. *Ann Surg* **225**, 317–326.
27. Woodward BD & Miller RG (1991) Depression of thymus-dependent immunity in wasting protein–energy malnutrition does not depend on an altered ratio of helper (CD4+) to suppressor (CD8+) T cells or on a disproportionately large atrophy of the T-cell relative to the B-cell pool. *Am J Clin Nutr* **53**, 1329–1335.
28. Lee WH & Woodward BD (1996) The CD4/CD8 ratio in the blood does not reflect the response of this index in secondary lymphoid organs of weanling mice in models of protein–energy malnutrition known to depress thymus-dependent immunity. *J Nutr* **126**, 849–859.
29. McMurray DN, Mintzer CL, Bartow RA *et al.* (1989) Dietary protein deficiency and *Mycobacterium bovis* BCG affect interleukin-2 activity in experimental pulmonary tuberculosis. *Infect Immun* **57**, 2606–2611.
30. Mengheri E, Nobili F, Crocchioni G *et al.* (1992) Protein starvation impairs the ability of activated lymphocytes to produce interferon-gamma. *J Interferon Res* **12**, 17–21.
31. Parent G, Chevalier P, Zalles L *et al.* (1994) *In vitro* lymphocyte-differentiating effects of thymulin (Zn-FTS) on lymphocyte subpopulations of severely malnourished children. *Am J Clin Nutr* **60**, 274–278.
32. Scott P & Trinchieri G (1995) The role of natural killer cells in host-parasite interactions. *Curr Opin Immunol* **7**, 34–40.
33. Ingram KG, Crouch DA, Douez DL *et al.* (1995) Effects of triiodothyronine supplements on splenic natural killer cells in malnourished weanling mice. *Int J Immunopharmacol* **17**, 21–32.
34. Salimonu LS, Ojo-Amaize E, Williams AI *et al.* (1982) Depressed natural killer cell activity in children with protein-calorie malnutrition. *Clin Immunol Immunopathol* **24**, 1–7.
35. Weindruch R, Devens BH, Raff HV *et al.* (1983) Influence of dietary restriction and aging on natural killer cell activity in mice. *J Immunol* **130**, 993–996.
36. Skerrett SJ, Henderson WR & Martin TR (1990) Alveolar macrophage function in rats with severe protein calorie malnutrition. Arachidonic acid metabolism, cytokine release, and antimicrobial activity. *J Immunol* **144**, 1052–1061.
37. Salimonu LS, Johnson AOK, Willians AIO *et al.* (1982) Phagocyte function in protein–calorie malnutrition. *Nutr Res* **2**, 445–454.
38. Rivera J, Habicht J-P, Torres N *et al.* (1986) Decreased cellular immune response in wasted but not in stunted children. *Nutr Res* **6**, 1161–1170.
39. Koster FT, Palmer DL, Chakraborty J *et al.* (1987) Cellular immune competence and diarrheal morbidity in malnourished Bangladeshi children: a prospective field study. *Am J Clin Nutr* **46**, 115–120.
40. Baqui AH, Sack RB, Black RE *et al.* (1993) Cell-mediated immune deficiency and malnutrition are independent risk factors for persistent diarrhea in Bangladeshi children. *Am J Clin Nutr* **58**, 543–548.
41. Semba RD (1998) The role of vitamin A and related retinoids in immune function. *Nutr Rev* **56**, S38–S48.
42. Semba RD (1999) Vitamin A and immunity to viral, bacterial and protozoan infections. *Proc Nutr Soc* **58**, 719–727.
43. Semba RD (2002) Vitamin A, infection and immune function. In *Nutrition and Immune Function*, pp. 151–169 [PC Calder, CJ Field and HS Gill, editors]. Wallingford: CAB International.
44. Stephensen CB (2001) Vitamin A, infection, and immune function. *Annu Rev Nutr* **21**, 167–192.
45. Villamor E & Fawzi WW (2005) Effects of vitamin A supplementation on immune responses and correlation with clinical outcomes. *Clin Microbiol Rev* **18**, 446–464.
46. Ahmed F, Jones DB & Jackson AA (1990) The interaction of vitamin A deficiency and rotavirus infection in the mouse. *Br J Nutr* **63**, 363–373.
47. Lawson ND & Berliner N (1999) Neutrophil maturation and the role of retinoic acid. *Exp Hematol* **27**, 1355–1367.
48. Twining SS, Schulte DP, Wilson PM *et al.* (1997) Vitamin A deficiency alters rat neutrophil function. *J Nutr* **127**, 558–565.
49. Ongsakul M, Sirisinha S & Lamb AJ (1985) Impaired blood clearance of bacteria and phagocytic activity in vitamin A-deficient rats. *Proc Soc Exp Biol Med* **178**, 204–208.
50. Ross AC (1996) Vitamin A deficiency and retinoid depletion regulate the antibody response to bacterial antigens and the maintenance of natural killer cells. *Clin Immunol Immunopathol* **80**, S36–S72.
51. Cantorna MT, Nashold FE & Hayes CE (1994) In vitamin A deficiency multiple mechanisms establish a regulatory T helper cell imbalance with excess Th1 and insufficient Th2 function. *J Immunol* **152**, 1515–1522.
52. Iwata M, Eshima Y & Kagechika H (2003) Retinoic acids exert direct effects on T cells to suppress Th1 development and enhance Th2 development via retinoic acid receptors. *Int Immunol* **15**, 1017–1025.
53. Ma Y, Chen Q & Ross AC (2005) Retinoic acid and polyribonucleosinic acid stimulate robust anti-tetanus antibody production while differentially regulating type 1/type 2 cytokines and lymphocyte populations. *J Immunol* **174**, 7961–7969.
54. Hoag KA, Nashold FE, Goverman J *et al.* (2002) Retinoic acid enhances the T helper 2 cell development that is

- essential for robust antibody responses through its action on antigen-presenting cells. *J Nutr* **132**, 3736–3739.
55. Ivanov II, Zhou L & Littman DR (2007) Transcriptional regulation of Th17 cell differentiation. *Semin Immunol* **19**, 409–417.
 56. Takaki H, Ichiyama K, Koga K *et al.* (2008) STAT6 inhibits TGF-beta1-mediated Foxp3 induction through direct binding to the Foxp3 promoter, which is reverted by retinoic acid receptor. *J Biol Chem* **283**, 14955–14962.
 57. Iwata M, Hirakiyama A, Eshima Y *et al.* (2004) Retinoic acid imprints gut-homing specificity on T cells. *Immunity* **21**, 527–538.
 58. Mucida D, Park Y & Cheroutre H (2009) From the diet to the nucleus: vitamin A and TGF-beta join efforts at the mucosal interface of the intestine. *Semin Immunol* **21**, 14–21.
 59. Semba RD, Muhilal, Scott AL *et al.* (1992) Depressed immune response to tetanus in children with vitamin A deficiency. *J Nutr* **122**, 101–107.
 60. Hewison M (2012) Vitamin D and immune function: auto-crine, paracrine or endocrine? *Scand J Clin Lab Invest Suppl* **243**, 92–102.
 61. Ooi JH, Chen J & Cantorna MT (2012) Vitamin D regulation of immune function in the gut: why do T cells have vitamin D receptors? *Mol Aspects Med* **33**, 77–82.
 62. Hewison M (2012) An update on vitamin D and human immunity. *Clin Endocrinol* **76**, 315–325.
 63. Di Rosa M, Malaguarnera M, Nicoletti F *et al.* (2011) Vitamin D3: a helpful immuno-modulator. *Immunology* **134**, 123–139.
 64. Van Belle TL, Gysemans C & Mathieu C (2011) Vitamin D in autoimmune, infectious and allergic diseases: a vital player? *Best Pract Res Clin Endocrinol Metab* **25**, 617–632.
 65. Hewison M (2012) Vitamin D and immune function: an overview. *Proc Nutr Soc* **71**, 50–61.
 66. Hewison M, Freeman L, Hughes SV *et al.* (2003) Differential regulation of vitamin D receptor and its ligand in human monocyte-derived dendritic cells. *J Immunol* **170**, 5382–5390.
 67. Liu PT & Modlin RL (2008) Human macrophage host defense against *Mycobacterium tuberculosis*. *Curr Opin Immunol* **20**, 371–376.
 68. Liu PT, Stenger S, Li H *et al.* (2006) Toll-like receptor triggering of a vitamin D-mediated human antimicrobial response. *Science* **311**, 1770–1773.
 69. Sabetta JR, DePetrillo P, Cipriani RJ *et al.* (2010) Serum 25-hydroxyvitamin D and the incidence of acute viral respiratory tract infections in healthy adults. *PLoS ONE* **5**, e11088.
 70. Urashima M, Segawa T, Okazaki M *et al.* (2010) Randomized trial of vitamin D supplementation to prevent seasonal influenza A in schoolchildren. *Am J Clin Nutr* **91**, 1255–1260.
 71. Hayes CE, Nashold FE, Spach KM *et al.* (2003) The immunological functions of the vitamin D endocrine system. *Cell Mol Biol* **49**, 277–300.
 72. Griffin MD, Xing N & Kumar R (2003) Vitamin D and its analogs as regulators of immune activation and antigen presentation. *Annu Rev Nutr* **23**, 117–145.
 73. van Etten E & Mathieu C (2005) Immunoregulation by 1,25-dihydroxyvitamin D3: basic concepts. *J Steroid Biochem Mol Biol* **97**, 93–101.
 74. Lemire JM, Archer DC, Beck L *et al.* (1995) Immunosuppressive actions of 1,25-dihydroxyvitamin D3: preferential inhibition of Th1 functions. *J Nutr* **125**, 1704S–1708S.
 75. Cantorna MT & Mahon BD (2004) Mounting evidence for vitamin D as an environmental factor affecting autoimmune diseases prevalence. *Exp Biol Med* **229**, 1136–1142.
 76. Deluca HF & Cantorna MT (2001) Vitamin D: its role and uses in immunology. *FASEB J* **15**, 2579–2585.
 77. Mathieu C & Adorini L (2002) The coming of age of 1,25-dihydroxyvitamin D(3) analogs as immunomodulatory agents. *Trends Mol Med* **8**, 174–179.
 78. Mathieu C, van Etten E, Decallonne B *et al.* (2004) Vitamin D and 1,25-dihydroxyvitamin D₃ as modulators in the immune system. *J Steroid Biochem Mol Biol* **89/90**, 449–452.
 79. Muller K, Odum N & Bendtzen K (1993) 1,25-dihydroxyvitamin D₃ selectively reduces interleukin-2 levels and proliferation of human t cell lines *in vitro*. *Immunol Lett* **35**, 177–182.
 80. Rigby WF, Stacy T & Fanger MW (1984) Inhibition of T lymphocyte mitogenesis by 1,25-dihydroxyvitamin D₃ (calcitriol). *J Clin Invest* **74**, 1451–1455.
 81. Tsoukas CD, Watry D, Escobar SS *et al.* (1989) Inhibition of interleukin-1 production by 1,25-dihydroxyvitamin D₃. *J Clin Endocrinol Metab* **69**, 127–133.
 82. Cipitelli M & Santoni A (1998) Vitamin D₃: a transcriptional modulator of the interferon-gamma gene. *Eur J Immunol* **28**, 3017–3030.
 83. Reichel H, Koeffler HP, Tobler A *et al.* (1987) 1alpha,25-dihydroxyvitamin D₃ inhibits gamma-interferon synthesis by normal human peripheral blood lymphocytes. *Proc Natl Acad Sci USA* **84**, 3385–3389.
 84. Tang J, Zhou R, Luger D *et al.* (2009) Calcitriol suppresses antiretinal autoimmunity through inhibitory effects on Th17 effector response. *J Immunol* **182**, 4624–4632.
 85. Lemire JM, Adams JS, Sakai R *et al.* (1984) 1 alpha,25-dihydroxyvitamin D₃ suppresses proliferation and immunoglobulin production by normal human peripheral blood mononuclear cells. *J Clin Invest* **74**, 657–661.
 86. Boonstra A, Barrat FJ, Crain C *et al.* (2001) 1alpha,25-dihydroxyvitamin D₃ has a direct effect on naïve CD4⁺ T cells to enhance the development of Th2 cells. *J Immunol* **167**, 4974–4980.
 87. Imazeki I, Matsuzaki J, Tsuji K *et al.* (2006) Immunomodulating effect of vitamin D₃ derivatives on type-1 cellular immunity. *Biomed Res* **27**, 1–9.
 88. Pichler J, Gerstmayr M, Szépfalusi Z *et al.* (2002) 1 alpha,25(OH)2D3 inhibits not only Th1 but also Th2 differentiation in human cord blood T cells. *Pediatr Res* **52**, 12–18.
 89. Barrat FJ, Cua DJ, Boonstra A *et al.* (2002) *In vitro* generation of interleukin 10-producing regulatory CD4⁺ T cells is induced by immunosuppressive drugs and inhibited by T helper type 1 (Th1)- and Th2-inducing cytokines. *J Exp Med* **195**, 603–616.
 90. Gregori S, Giarratana N, Smiroldo S *et al.* (2002) A 1alpha,25-dihydroxyvitamin D₃ analog enhances regulatory T-cells and arrests autoimmune diabetes in NOD mice. *Diabetes* **51**, 1367–1374.
 91. Meydani SN & Beharka AA (1998) Recent developments in vitamin E and immune response. *Nutr Rev* **56**, S49–S58.
 92. Serafini M (2000) Dietary vitamin E and T cell-mediated function in the elderly: effectiveness and mechanism of action. *Int J Dev Neurosci* **18**, 401–410.
 93. Meydani SN, Han SN & Wu D (2005) Vitamin E and immune response in the aged: mechanisms and clinical implications. *Immunol Rev* **205**, 269–284.
 94. Wu D & Meydani SN (2008) Age-associated changes in immune and inflammatory responses: impact of vitamin E intervention. *J Leukoc Biol* **84**, 900–914.
 95. Chavance M, Herbeth B, Fournier C *et al.* (1989) Vitamin status, immunity and infections in an elderly population. *Eur J Clin Nutr* **43**, 827–835.

96. Meydani SN, Barklund MP, Liu S *et al.* (1990) Vitamin E supplementation enhances cell-mediated immunity in healthy elderly subjects. *Am J Clin Nutr* **52**, 557–563.
97. Meydani SN, Meydani M, Blumberg JB *et al.* (1997) Vitamin E supplementation and *in vivo* immune response in healthy subjects. *JAMA* **277**, 1380–1386.
98. Pallast EG, Schouten EG, de Waart FG *et al.* (1999) Effect of 50- and 100-mg vitamin E supplements on cellular immune function in noninstitutionalized elderly persons. *Am J Clin Nutr* **69**, 1273–1281.
99. Meydani SN, Leka LS, Fine BC *et al.* (2004) Vitamin E and respiratory tract infections in elderly nursing home residents: a randomized controlled trial. *JAMA, J Am Med Assoc* **292**, 828–836.
100. Graat JM, Schouten EG & Kok FJ (2002) Effect of daily vitamin E and multivitamin-mineral supplementation on acute respiratory tract infections in elderly persons: a randomized controlled trial. *JAMA, J Am Med Assoc* **288**, 715–721.
101. Fraker PJ, King LE, Garvy BA *et al.* (1993) The immunopathology of zinc deficiency in humans and rodents: a possible role for programmed cell death. In *Nutrition and Immunology*, pp. 267–283 [DM Klurfeld, editor]. New York: Plenum Press.
102. Shankar AH & Prasad AS (1998) Zinc and immune function: the biological basis of altered resistance to infection. *Am J Clin Nutr* **68**, 447S–463S.
103. Prasad AS (2002) Zinc, infection and immune function. In *Nutrition and Immune Function*, pp. 193–207 [PC Calder, CJ Field and HS Gill, editors]. Wallingford: CAB International.
104. Prasad AS (2008) Zinc in human health: effect of zinc on immune cells. *Mol Med* **14**, 353–357.
105. Fischer Walker C & Black RE (2004) Zinc and the risk for infectious disease. *Annu Rev Nutr* **24**, 255–275.
106. Fraker PJ & King LE (2004) Reprogramming of the immune system during zinc deficiency. *Annu Rev Nutr* **24**, 277–298.
107. Allen JL, Perri RT, McClain CJ *et al.* (1983) Alterations in human natural killer cell activity and monocyte cytotoxicity induced by zinc deficiency. *J Lab Clin Med* **102**, 577–589.
108. Keen CL & Gershwin ME (1990) Zinc deficiency and immune function. *Annu Rev Nutr* **10**, 415–430.
109. Rink L & Kirchner H (2000) Zinc-altered immune function and cytokine production. *J Nutr* **130**, 1407S–1411S.
110. Rink L, Cakman I & Kirchner H (1998) Altered cytokine production in the elderly. *Mech Ageing Dev* **102**, 199–210.
111. Kahmann L, Uciechowski P, Warmuth S *et al.* (2008) Zinc supplementation in the elderly reduces spontaneous inflammatory cytokine release and restores T cell functions. *Rejuvenation Res* **11**, 227–237.
112. Prasad AS (2000) Effects of zinc deficiency on Th1 and Th2 cytokine shifts. *J Infect Dis* **182**, 62–68.
113. Beck FW, Prasad AS, Kaplan J *et al.* (1997) Changes in cytokine production and T cell subpopulations in experimentally induced zinc-deficient humans. *Am J Physiol* **272**, E1002–E1007.
114. Tapazoglou E, Prasad AS, Hill G *et al.* (1985) Decreased natural killer cell activity in patients with zinc deficiency with sickle cell disease. *J Lab Clin Med* **105**, 19–22.
115. Sherman AR, Spear AT (1993) Iron and immunity. In *Nutrition and Immunology*, pp. 285–307 [DM Klurfeld, editor]. New York and London: Plenum Press.
116. Kuvibidila S, Baliga BS (2002) Role of iron in immunity and infection. In *Nutrition and Immune Function*, pp. 208–228 [PC Calder, CJ Field and HS Gill, editors]. Wallingford: CAB International.
117. Weiss G (2002) Iron and immunity: a double-edged sword. *Eur J Clin Invest* **32**, Suppl. 1, 70–78.
118. Oppenheimer SJ (2001) Iron and its relation to immunity and infectious disease. *J Nutr* **131**, 616S–635S.
119. Schaible UE & Kaufmann SH (2004) Iron and microbial infection. *Nat Rev Microbiol* **2**, 946–953.
120. Markel TA, Crisostomo PR, Wang M *et al.* (2007) The struggle for iron: gastrointestinal microbes modulate the host immune response during infection. *J Leukoc Biol* **81**, 393–400.
121. Cherayil BJ (2010) Iron and immunity: immunological consequences of iron deficiency and overload. *Arch Immunol Ther Exp* **58**, 407–415.
122. Kumar V & Choudhry VP (2010) Iron deficiency and infection. *Indian J Pediatr* **77**, 789–793.
123. Barry DMJ & Reeve AW (1977) Increased incidence of gram negative neonatal sepsis with intramuscular iron administration. *Pediatrics* **60**, 908–912.
124. Murray MJ, Murray AB, Murray MB *et al.* (1978) Diet and cerebral malaria: the effect of famine and re-feeding. *Am J Clin Nutr* **31**, 57–61.
125. Murray MJ, Murray AB, Murray MB *et al.* (1978) The adverse effect of iron repletion on the course of certain infections. *Br Med J* **2**, 1113–1115.
126. Smith AW, Hendrickse RG, Harrison C *et al.* (1989) The effects on malaria of treatment of iron-deficiency anaemia with oral iron in Gambian children. *Ann Trop Paediatr* **9**, 17–23.
127. Johnson EE, Sandgren A, Cherayil BJ *et al.* (2010) Role of ferroportin in macrophage-mediated immunity. *Infect Immunology* **78**, 5099–5106.
128. McKenzie RC, Rafferty TS & Beckett GJ (1998) Selenium: an essential element for immune function. *Immunol Today* **19**, 342–345.
129. Huang Z, Rose AH & Hoffmann PR (2012) The role of selenium in inflammation and immunity: from molecular mechanisms to therapeutic opportunities. *Antioxid Redox Signal* **16**, 705–743.
130. Arthur JR, McKenzie RC & Beckett GJ (2003) Selenium in the immune system. *J Nutr* **133**, Suppl. 1, 1457S–1459S.
131. Beck MA & Levander OA (2000) Host nutritional status and its effect on a viral pathogen. *J Infect Dis* **182**, Suppl. 1, S93–S96.
132. Beck MA, Handy J & Levander OA (2004) Host nutritional status: the neglected virulence factor. *Trends Microbiol* **12**, 417–423.
133. Wang C, Wang H, Luo J *et al.* (2009) Selenium deficiency impairs host innate immune response and induces susceptibility to *Listeria monocytogenes* infection. *BMC Immunol* **10**, 55.
134. Ravaglia G, Forti P, Maioli F *et al.* (2000) Effect of micronutrient status on natural killer cell immune function in healthy free-living subjects aged ≥ 90 y. *Am J Clin Nutr* **71**, 590–598.
135. Shor-Posner G, Miguez MJ, Pineda LM *et al.* (2002) Impact of selenium status on the pathogenesis of mycobacterial disease in HIV-1-infected drug users during the era of highly active antiretroviral therapy. *J Acquir Immune Defic Syndr* **29**, 169–173.
136. Roy M, Kiremidjian-Schumacher L, Wishe HI *et al.* (1994) Supplementation with selenium and human immune cell functions. I. Effect on lymphocyte proliferation and interleukin 2 receptor expression. *Biol Trace Elem Res* **41**, 103–114.
137. Hawkes WC, Kelley DS & Taylor PC (2001) The effects of dietary selenium on the immune system in healthy men. *Biol Trace Elem Res* **81**, 189–213.



138. Kiremidjian-Schumacher L, Roy M, Wishe HI *et al.* (1994) Supplementation with selenium and human immune cell functions. II. Effect on cytotoxic lymphocytes and natural killer cells. *Biol Trace Elem Res* **41**, 115–127.
139. Peretz A, Nève J, Desmedt J *et al.* (1991) Lymphocyte response is enhanced by supplementation of elderly subjects with selenium-enriched yeast. *Am J Clin Nutr* **53**, 1323–1328.
140. Roy M, Kiremidjian-Schumacher L, Wishe HI *et al.* (1995) Supplementation with selenium restores age-related decline in immune cell function. *Proc Soc Exp Biol Med* **209**, 369–375.
141. Broome CS, McArdle F, Kyle JA *et al.* (2004) An increase in selenium intake improves immune function and poliovirus handling in adults with marginal selenium status. *Am J Clin Nutr* **80**, 154–162.
142. Thomas CM & Versalovic J (2010) Probiotics-host communication: modulation of signaling pathways in the intestine. *Gut Microbes* **1**, 148–163.
143. Hemarajata P & Versalovic J (2013) Effects of probiotics on gut microbiota: mechanisms of intestinal immunomodulation and neuromodulation. *Therap Adv Gastroenterol* **6**, 39–51.
144. Dong H, Rowland I & Yaqoob P (2012) Comparative effects of six probiotic strains on immune function *in vitro*. *Brit J Nutr* **108**, 459–470.
145. Lomax AR & Calder PC (2009) Probiotics, immune function, infection and inflammation: a review of the evidence from studies conducted in humans. *Curr Pharma Design* **15**, 1428–1518.
146. Boge T, Rémy M, Vaudaine S *et al.* (2009) A probiotic fermented dairy drink improves antibody response to influenza vaccination in the elderly in two randomised controlled trials. *Vaccine* **27**, 5677–5684.
147. Rizzardini G, Eskesen D, Calder PC *et al.* (2012) Evaluation of the immune benefits of two probiotic strains *Bifidobacterium animalis ssp. lactis*, BB-12[®] and *Lactobacillus paracasei ssp. paracasei*, L. casei 431[®] in an influenza vaccination model: a randomised, double-blind, placebo-controlled study. *Br J Nutr* **107**, 876–884.
148. Maidens C, Childs C, Przemaska A *et al.* (2013) Modulation of vaccine response by concomitant probiotic administration. *Br J Clin Pharmacol* **75**, 663–670.
149. Hickson M, D'Souza AL, Muthu N *et al.* (2007) Use of probiotic *Lactobacillus* preparation to prevent diarrhoea associated with antibiotics: randomised double blind placebo controlled trial. *Br Med J* **335**, 80–83.
150. McFarland LV (2006) Meta-analysis of probiotics for the prevention of antibiotic associated diarrhea and the treatment of *Clostridium difficile* disease. *Am J Gastroenterol* **101**, 812–822.
151. Allen SJ, Martinez EG, Gregorio GV *et al.* (2010) Probiotics for treating acute infectious diarrhoea. *Cochrane Database Syst Rev* **11**, CD003048.
152. Hempel S, Newberry SJ, Maher AR *et al.* (2012) Probiotics for the prevention and treatment of antibiotic-associated diarrhea. *JAMA, J Am Med Assoc* **307**, 1959–1969.
153. Calder P & Hall V (2012) Understanding gut-immune interactions in management of acute infectious diarrhoea. *Nurs Older People* **24**, 29–37.
154. Lomax AR & Calder PC (2009) Probiotics, immune function, infection and inflammation: a review of the evidence. *Br J Nutr* **101**, 633–658.
155. Mitra AK, Akramuzzaman SM, Fuchs GJ *et al.* (1997) Long-term oral supplementation with iron is not harmful for young children in a poor community of Bangladesh. *J Nutr* **127**, 1451–1455.