

## Workshop Report

# Risk of iron overload in carriers of genetic mutations associated with hereditary haemochromatosis: UK Food Standards Agency workshop

Mamta Singh<sup>1\*</sup>, Margaret Ashwell<sup>2</sup>, Peter Sanderson<sup>1</sup>, Janet Cade<sup>3</sup>, Jennifer Moreton<sup>3</sup>, Susan Fairweather-Tait<sup>4</sup>, Mark Roe<sup>4</sup>, Joannes J. M. Marx<sup>5</sup>, Mark Worwood<sup>6</sup> and James D. Cook<sup>7</sup>

<sup>1</sup>Food Standards Agency, Aviation House, 125 Kingsway, London WC2B 6NH, UK

<sup>2</sup>Ashwell Associates, Ashwell Street, Ashwell, Hertfordshire SG7 5PZ, UK

<sup>3</sup>Leeds University, Centre for Epidemiology and Biostatistics, 30–32 Hyde Terrace, Leeds LS2 9JT, UK

<sup>4</sup>Institute of Food Research, Norwich Research Park, Colney, Norwich NR4 7UA, UK

<sup>5</sup>Eijkman-Winkler Institute for Microbiology, Infections Diseases and Inflammation, University Medical Centre, Utrecht, the Netherlands

<sup>6</sup>University of Wales College of Medicine, Cardiff CF14 4XN, UK

<sup>7</sup>University of Kansas Medical Centre, Kansas City, KS 66160, USA

(Received 6 May 2006 – Revised 23 May 2006 – Accepted 23 May 2006)

The UK Food Standards Agency convened a group of expert scientists to review current research investigating diet and carriers of genetic mutations associated with hereditary haemochromatosis. The workshop concluded that individuals who are heterozygous for the C282Y mutation of the *HFE* gene do not appear to respond abnormally to dietary Fe and therefore do not need to change their diet to prevent accumulation of body Fe.

### Iron overload: Hereditary haemochromatosis: Food Standards Agency workshops

#### Background

The UK Food Standards Agency (FSA) convened a workshop on 12 November 2003 on the nutritional implications of mutations of the gene linked to haemochromatosis. The results from recently completed studies were presented, both FSA- and non-FSA funded, and the workshop was chaired by Professor Jim Cook.

#### Introduction

Hereditary haemochromatosis is a common, autosomally recessive, genetic disorder of Fe metabolism. It is characterised by excessive Fe absorption, resulting in the accumulation of high levels of Fe body stores. If left untreated it can have serious consequences, including diabetes, arthritis and cirrhosis of the liver (Bothwell & MacPhail, 1998).

Mutation of the *HFE* gene, which is involved in Fe absorption and storage, has been identified as the major cause of haemochromatosis (Feder *et al.* 1996). Two mutations of the *HFE* gene, C282Y and H63D, have been linked with the disease. In the UK, most patients with haemochromatosis are homozygous for

the C282Y mutation of the *HFE* gene (The UK Haemochromatosis Consortium, 1997), although the majority of homozygotes in the population do not develop clinical symptoms leading to a diagnosis of haemochromatosis (McCune *et al.* 2002). In a minority of cases, there may be mutations in the genes encoding transferrin receptor 2, ferroportin-1, haemojuvelin and hepcidin (Roetto *et al.* 2003; Swinkels *et al.* 2006).

Mutations in the *HFE* gene are common in populations of European origin (Merryweather-Clarke *et al.* 2000). In a study of blood donors in the UK, 15% were heterozygous, and 0.7% homozygous, for C282Y, while 25% were heterozygous, and 2% homozygous, for the H63D mutation (Jackson *et al.* 2001). Fe stores (as measured by transferrin saturation (TS), serum ferritin (sFn) concentration and serum Fe binding capacity) were observed to increase in the order: wild types, H63D heterozygotes, C282Y heterozygotes, H63D homozygotes, compound heterozygotes and C282Y homozygotes; although sFn concentrations were no higher in H63D heterozygotes and C282Y heterozygous women than in wild types. If these increases in TS and sFn truly reflect increases in tissue Fe levels, subjects carrying *HFE* mutations may be at increased risk of disease.

### Assessment of body tissue iron

Professor Jim Cook described a technique for measuring body tissue Fe, either the surplus of storage Fe or the deficit of tissue Fe, which uses the serum transferrin receptor (TfR):sFn ratio (Cook *et al.* 2003). A low TfR:sFn ratio indicates high Fe stores. The method was demonstrated to estimate the body Fe content of healthy subjects (Cook *et al.* 2003). The method provides individual values for storage Fe and also permits a substantial reduction in the number of subjects needed to assess response to an intervention study. The use of this method to compare different studies will not be possible, however, until the assay has been standardised.

The hepatic protein, hepcidin, found in urine and blood, is thought to regulate dietary Fe uptake and its distribution in the body by controlling the concentration of ferroportin, an Fe exporter present on the surface of absorptive enterocytes, macrophages and hepatocytes (Nemeth *et al.* 2004). Hepcidin might serve as a useful indicator of body tissue Fe in future studies (Kemna *et al.* 2005).

### Genetic influence on iron status: morbidity and mortality

Professor Mark Worwood presented epidemiological evidence of morbidity and mortality outcomes associated with hereditary haemochromatosis. A study in the USA observed no increase in the frequency of the most common symptoms of haemochromatosis in subjects homozygous for C282Y, or heterozygotes for both H63D and C282Y, relative to wild-type subjects (Beutler *et al.* 2002). The frequency was only higher for C282Y homozygotes reporting liver problems or with an increased aspartate transaminase activity. A study in Wales of haemochromatosis patients observed that only 1.2% of adults homozygous for C282Y had been diagnosed with Fe overload (McCune *et al.* 2002); furthermore, the frequency of homozygosity for C282Y is not under-represented in old age (Willis *et al.* 1999; Coppin *et al.* 2003).

Several prospective studies have suggested that heterozygotes for C282Y may be at increased risk of CVD (Roest *et al.* 1999; Tuomainen *et al.* 1999; Rasmussen *et al.* 2001); however, other prospective studies have observed no association between *HFE* mutations and CVD risk (Gunn *et al.* 2004; Ellervik *et al.* 2005).

### The iron hypothesis of atherosclerosis

Professor Joe Marx presented evidence on the hypothesis that body Fe stores are associated with CVD risk (Sullivan, 1981). Several prospective studies have observed an association between increased sFn concentrations and increased carotid atherosclerosis (Kiechl *et al.* 1997; Wolff *et al.* 2004) and CVD (Salonen *et al.* 1992; van der A *et al.* 2005). Ferritin is an acute-phase protein, which is increased in inflammation. Chronic inflammation (for example, as assessed by plasma C-reactive protein concentrations) is associated with an increased risk of CVD (Danesh *et al.* 1998). One study, which adjusted for C-reactive protein concentrations, found an association between sFn concentrations and ischaemic stroke risk in postmenopausal women (van der A *et al.* 2005). Other prospective studies, however, have found little

or no evidence that sFn concentrations are a risk factor for CVD (Danesh & Appleby, 1999; Knuiman *et al.* 2003).

TS has not been associated with CVD risk (Sempos *et al.* 1994; Reunanen *et al.* 1995; Danesh & Appleby, 1999; van der A *et al.* 2005), although the TfR:sFn ratio was associated with increased CVD risk in one study (Tuomainen *et al.* 1998). The results from epidemiological studies, therefore, remain equivocal.

In plasma, Fe is normally bound to transferrin, making it less reactive. If Fe is in a labile and reactive form in plasma, unbound to transferrin, it can react with cells and molecules. Non-transferrin-bound Fe has been found in the plasma of patients with Fe-overload conditions (Grootveld *et al.* 1989), homozygotes and heterozygotes for C282Y (de Valk *et al.* 2000) and in patients with end-stage renal disease (Kooistra *et al.* 2002). Non-transferrin-bound Fe has also been shown to increase the expression of adhesion molecules in cultured endothelial cells and monocytes, representing an early phase of atherosclerosis (Kartikasari *et al.* 2004). These studies provide some support for the Fe hypothesis; however, a recent prospective study found no association between CVD risk and plasma non-transferrin-bound Fe concentrations in postmenopausal women (van der A *et al.* 2006).

### The effects of diet on iron absorption and iron stores in *HFE* genotypes

Regulation of Fe absorption is affected by mutations in the *HFE* gene. The consequences for C282Y heterozygotes, however, are not clear. One study suggested that absorption from diets of high bioavailability may be increased in heterozygotes (Lynch *et al.* 1989), but these findings were not replicated in another study (Hunt & Zeng, 2004).

Professor Sue Fairweather-Tait presented results from an FSA-funded project that measured Fe absorption from a diet containing highly bioavailable Fe (high meat, fish and poultry, high vitamin C, low phytate) extrinsically labelled with <sup>57</sup>Fe-enriched ferrous sulfate and a diet of fortified cereal products extrinsically labelled with <sup>54</sup>Fe-enriched ferrous sulfate, over a 5 d period in C282Y heterozygotes (*n* 15), C282Y/H63D compound heterozygotes (*n* 5), and wild-type men (*n* 15). An erythrocyte incorporation technique was used to evaluate the effects of high dietary Fe on Fe homeostasis. Overall, the differences in Fe absorption between the various genotypes were small and genotype did not affect Fe absorption (Roe *et al.* 2005). There was no association between genotype and sFn concentrations.

The relationship between habitual diet and body Fe stores (Hb, sFn, TS, TfR) in C282Y heterozygote (*n* 46) and wild-type (*n* 92) men aged 40 years and over was also assessed, to determine the extent to which diet modulates Fe accumulation. A positive association was found between haem-Fe intake and sFn concentration. The C282Y heterozygotes had slightly higher TS, but genotype only accounted for 5% of the variation in TS and there was no difference in other measures of Fe stores.

Professor Janet Cade presented results from another FSA-funded project where 2531 women, from the UK Women's Cohort Study (aged 35–69 years at baseline), were genotyped for C282Y and H63D mutations and assessed for Fe intake and stores. Although C282Y homozygotes (*n* 31) had higher

sFn concentrations, there was no difference between heterozygotes for C282Y or H63D ( $n$  726) and wild types ( $n$  1774). Heterozygotes for C282Y did have slightly higher TS concentrations than wild-type subjects (Greenwood *et al.* 2005).

Dietary assessment showed that sFn concentration was positively associated with haem-Fe intake. Women heterozygous for the C282Y mutation responded to haem Fe similarly to normal women with regard to their sFn concentrations. A strong interaction between genotype, haem-Fe intake and raised sFn concentrations was observed, however, primarily in postmenopausal women who were homozygous for C282Y (Cade *et al.* 2005).

Overall, these studies confirm previously published reports that C282Y heterozygotes have a slightly higher TS compared with wild-type controls, but do not have a higher sFn concentration (Jackson *et al.* 2001; Beutler *et al.* 2002). Absorption of Fe, and Fe stores, do not appear to be significantly higher in C282Y heterozygotes than in wild types.

## Discussion

Individuals who are heterozygous for the C282Y mutation of the *HFE* gene appear to respond normally to dietary Fe and do not need to alter their diet to avoid increased levels of stored Fe.

The following research recommendations were identified:

- (1) Standardisation of the serum TfR assay to encourage its use as a marker of body Fe content, in conjunction with sFn, in clinical and population studies;
- (2) Adjustment for inflammation and infection in studies associating chronic disease with sFn concentrations;
- (3) Prospective studies to explore the possible link between penetrance of *HFE* mutations in relation to dietary patterns and other factors associated with Fe metabolism and risk of chronic disease.

## Participants

Professor James Cook, University of Kansas Medical Center, USA; Professor Joe J. M. Marx, University Medical Centre, Utrecht, The Netherlands; Professor Mark Worwood, University of Wales College of Medicine; Professor Sue Fairweather-Tait, Institute of Food Research; Dr Janet Cade, Leeds University; Dr Mark Roe, Institute of Food Research; Dr Birgit Teucher, Institute of Food Research; Dr Tim Key, Scientific Advisory Committee on Nutrition (SACN); Professor Peter Aggett, SACN; Dr Kathryn Robson, John Radcliffe Hospital, Oxford; Dr Alison Merryweather Clark, John Radcliffe Hospital, Oxford; Dr H el ene Coppin, INSERM U563, CHU Purpan, Toulouse; Dr Marie Paule Roth, INSERM U563, CHU Purpan, Toulouse; Dr Gavin Willis, Norfolk & Norwich University; Dr Jennifer Moreton, Leeds University; Dr Anne McCune, Gloucestershire Royal Hospital; Dr Margaret Ashwell, FSA Programme Adviser; Dr Alison Tedstone, FSA; Rachel Elsom, FSA; Mamta Singh, FSA; Cheryl White, FSA.

## References

Beutler E, Felitti VJ, Koziol JA, Ho NJ & Gelbart T (2002) Penetrance of 845G  $\rightarrow$  A (C282Y) *HFE* hereditary haemochromatosis mutation in the USA. *Lancet* **359**, 211–218.

Bothwell TH & MacPhail AP (1998) Hereditary hemochromatosis: etiologic, pathologic, and clinical aspects. *Semin Hematol* **35**, 55–71.

Cade JE, Moreton JA, O'Hara B, Greenwood DC, Moor J, Burley VJ, Kukulizch K, Bishop DT & Worwood M (2005) Diet and genetic factors associated with iron status in middle-aged women. *Am J Clin Nutr* **82**, 813–820.

Cook JD, Flowers CH & Skikne BS (2003) The quantitative assessment of body iron. *Blood* **101**, 3359–3364.

Coppin H, Bensaid M, Fruchon S, Borot N, Blanche H & Roth MP (2003) Longevity and carrying the C282Y mutation for haemochromatosis on the *HFE* gene: case control study of 492 French centenarians. *BMJ* **327**, 132–133.

Danesh J & Appleby P (1999) Coronary heart disease and iron status: meta-analyses of prospective studies. *Circulation* **99**, 852–854.

Danesh J, Collins R, Appleby P & Peto R (1998) Association of fibrinogen, C-reactive protein, albumin, or leukocyte count with coronary heart disease: meta-analyses of prospective studies. *JAMA* **279**, 1477–1482.

de Valk B, Addicks MA, Gosriwatana I, Lu S, Hider RC & Marx JJ (2000) Non-transferrin-bound iron is present in serum of hereditary haemochromatosis heterozygotes. *Eur J Clin Invest* **30**, 248–251.

Ellervik C, Tybjaerg-Hansen A, Grande P, Appleyard M & Nordestgaard BG (2005) Hereditary hemochromatosis and risk of ischemic heart disease: a prospective study and a case-control study. *Circulation* **112**, 185–193.

Feder JN, Gnirke A, Thomas W, *et al.* (1996) A novel MHC class I-like gene is mutated in patients with hereditary haemochromatosis. *Nat Genet* **13**, 399–408.

Greenwood DC, Cade JE, Moreton JA, O'Hara B, Burley VJ, Randsen-Moor JA, Kukulizch K, Thompson D, Worwood M & Bishop DT (2005) *HFE* genotype modifies the influence of heme iron intake on iron status. *Epidemiology* **16**, 802–805.

Grootveld M, Bell JD, Halliwell B, Aruoma OI, Bomford A & Sadler PJ (1989) Non-transferrin-bound iron in plasma or serum from patients with idiopathic hemochromatosis. Characterization by high performance liquid chromatography and nuclear magnetic resonance spectroscopy. *J Biol Chem* **264**, 4417–4422.

Gunn IR, Maxwell FK, Gaffney D, McMahon AD & Packard CJ (2004) Haemochromatosis gene mutations and risk of coronary heart disease: a west of Scotland coronary prevention study (WOSCOPS) substudy. *Heart* **90**, 304–306.

Hunt JR & Zeng H (2004) Iron absorption by heterozygous carriers of the *HFE* C282Y mutation associated with hemochromatosis. *Am J Clin Nutr* **80**, 924–931.

Jackson HA, Carter K, Darke C, Guttridge MG, Ravine D, Hutton RD, Napier JA & Worwood M (2001) *HFE* mutations, iron deficiency and overload in 10,500 blood donors. *Br J Haematol* **114**, 474–484.

Kartikasari AE, Georgiua NA, Visseren FL, Kats-Renaud H, van Asbeck BS & Marx JJM (2004) Intracellular labile iron modulates adhesion of human monocytes to human endothelial cells. *Arterioscler Thromb Vasc Biol* **24**, 2257–2262.

Kemna E, Tjalsma H, Laarakkers C, Nemeth E, Willems H & Swinkels D (2005) Novel urine hepcidin assay by mass spectrometry. *Blood* **106**, 3268–3270.

Kiechl S, Willeit J, Egger G, Poewe W & Oberhollenzer F (1997) Body iron stores and the risk of carotid atherosclerosis: prospective results from the Bruneck Study. *Circulation* **96**, 3300–3307.

Knuiman MW, Divitini ML, Olynyk JK, Cullen DJ & Bartholomew HC (2003) Serum ferritin and cardiovascular disease: a 17-year follow-up study in Busselton, Western Australia. *Am J Epidemiol* **158**, 144–149.

Kooistra MP, Kersting S, Gosriwatana I, Lu S, Nijhoff-Schutte J, Hider RC & Marx JJ (2002) Nontransferrin-bound iron in the plasma of haemodialysis patients after intravenous iron saccharate infusion. *Eur J Clin Invest* **32**, Suppl. 1, 36–41.

Lynch SR, Skikne BS & Cook JD (1989) Food iron absorption in idiopathic hemochromatosis. *Blood* **74**, 2187–2193.

- McCune CA, Al Jader LN, May A, Hayes SL, Jackson HA & Worwood M (2002) Hereditary haemochromatosis: only 1% of adult HFE C282Y homozygotes in South Wales have a clinical diagnosis of iron overload. *Hum Genet* **111**, 538–543.
- Merryweather-Clarke AT, Pointon JJ, Jouanolle AM, Rochette J & Robson KJ (2000) Geography of HFE C282Y and H63D mutations. *Genet Test* **4**, 183–198.
- Nemeth E, Tuttle MS, Powelson J, Vaughn MB, Donovan A, Ward DM, Ganz T & Kaplan J (2004) Hepcidin regulates cellular iron efflux by binding to ferroportin and inducing its internalization. *Science* **306**, 2090–2093.
- Rasmussen ML, Folsom AR, Catellier DJ, Tsai MY, Garg U & Eckfeldt JH (2001) A prospective study of coronary heart disease and the hemochromatosis gene (HFE) C282Y mutation: the Atherosclerosis Risk in Communities (ARIC) study. *Atherosclerosis* **154**, 739–746.
- Reunanen A, Takkunen H, Knekt P, Seppanen R & Aromaa A (1995) Body iron stores, dietary iron intake and coronary heart disease mortality. *J Intern Med* **238**, 223–230.
- Roe MA, Heath AL, Oyston SL, Macrow C, Hoogewerff JA, Foxall R, Dainty JR, Majsak-Newman G, Willis G & Fairweather-Tait SJ (2005) Iron absorption in male C282Y heterozygotes. *Am J Clin Nutr* **81**, 814–821.
- Roest M, van der Schouw YT, de Valk B, Marx JJ, Tempelman MJ, de Groot PG, Sixma JJ & Banga JD (1999) Heterozygosity for a hereditary hemochromatosis gene is associated with cardiovascular death in women. *Circulation* **100**, 1268–1273.
- Roetto A, Papanikolaou G, Politou M, Alberti F, Girelli D, Christakis J, Loukopoulos D & Camaschella C (2003) Mutant antimicrobial peptide hepcidin is associated with severe juvenile hemochromatosis. *Nat Genet* **33**, 21–22.
- Salonen JT, Nyyssonen K, Korpela H, Tuomilehto J, Seppanen R & Salonen R (1992) High stored iron levels are associated with excess risk of myocardial infarction in eastern Finnish men. *Circulation* **86**, 803–811.
- Sempos CT, Looker AC, Gillum RF & Makuc DM (1994) Body iron stores and the risk of coronary heart disease. *N Engl J Med* **330**, 1119–1124.
- Sullivan JL (1981) Iron and the sex difference in heart disease risk. *Lancet* **i**, 1293–1294.
- Swinkels DW, Janssen MC, Bergmans J & Marx JJM (2006) Hereditary hemochromatosis: genetic complexity and new diagnostic approaches. *Clin Chem* **52**, 950–968.
- The UK Haemochromatosis Consortium (1997) A simple genetic test identifies 90% of UK patients with haemochromatosis. *Gut* **41**, 841–844.
- Tuomainen TP, Kontula K, Nyyssonen K, Lakka TA, Helio T & Salonen JT (1999) Increased risk of acute myocardial infarction in carriers of the hemochromatosis gene Cys282Tyr mutation: a prospective cohort study in men in eastern Finland. *Circulation* **100**, 1274–1279.
- Tuomainen TP, Punnonen K, Nyyssonen K & Salonen JT (1998) Association between body iron stores and the risk of acute myocardial infarction in men. *Circulation* **97**, 1461–1466.
- van der A DL, Grobbee DE, Roest M, Marx JJM, Voorbij HA & van der Schouw YT (2005) Serum ferritin is a risk factor for stroke in postmenopausal women. *Stroke* **36**, 1637–1641.
- van der A DL, Marx JJM, Grobbee DE, Kamphuis MH, Georgiou NA, van Kats-Renaud JH, Breuer W, Cabantchik ZI, Roest M & Voorbij HA (2006) Non-transferrin-bound iron and risk of coronary heart disease in postmenopausal women. *Circulation* **113**, 1942–1949.
- Willis G, Wimperis JZ, Smith KC, Fellows IW & Jennings BA (1999) Haemochromatosis gene C282Y homozygotes in an elderly male population. *Lancet* **354**, 221–222.
- Wolff B, Volzke H, Ludemann J, Robinson D, Vogelgesang D, Staudt A, Kessler C, Dahm JB, John U & Felix SB (2004) Association between high serum ferritin levels and carotid atherosclerosis in the Study of Health in Pomerania (SHIP). *Stroke* **35**, 453–457.