

Availability of lysine in vegetable protein concentrates as determined by the slope-ratio assay with growing pigs and rats and by chemical techniques

BY E. S. BATTERHAM, R. D. MURISON* AND L. M. ANDERSEN

*Department of Agriculture, Agricultural Research Centre, Wollongbar,
New South Wales 2480, Australia*

(Received 18 October 1982 - Accepted 3 August 1983)

1. The availability of lysine in nine vegetable-protein concentrates was assessed using the slope-ratio assay for growing pigs and rats. Diets were equalized for crude fibre using solka floc to minimize any possible effects of variation in fibre content on availability estimates.

2. The availability of lysine in the nine proteins for pigs, using food conversion efficiency (FCE) on a carcass basis as the criterion of response were (proportion of total): cottonseed meal 0.39, lupin (*Lupinus angustifolius*) seed meal no. 1 0.37, no. 2 0.65, no. 3 0.54, no. 4 0.54, field peas (*Pisum sativum*) 0.93, peanut (groundnut) meal 0.57, soya-bean meal no. 1 0.98, no. 2 0.89.

3. Estimates of available lysine for rats as assessed by the slope-ratio assay using FCE on a carcass basis were in close agreement with the pig estimates for cottonseed meal (0.35) and soya-bean meal no. 1 (0.91) and no. 2 (0.89), higher for lupin-seed meals (range 0.70-0.94 with a mean of 0.81) and peanut meal (0.76) and lower for field peas (0.76).

4. The differences in available lysine were not detected by the chemical Silcock available-lysine test (Roach *et al.* 1967) or by the direct 1-fluoro-2,4-dinitrobenzene procedure (Carpenter, 1960).

Previous work (Batterham *et al.* 1979) showed that, for growing pigs, the availability of lysine in cottonseed meal and meat-and-bone meal was approximately half that in fish meal, skim-milk powder and soya-bean meal. Similar differences were detected with a slope-ratio assay with rats but not with the chemical Silcock available-lysine assay as developed by Roach *et al.* (1967).

Further studies (Batterham *et al.* 1981) on the availability of lysine in vegetable-protein concentrates confirmed that lysine availability varied widely and was not detected by chemical techniques. However, there was a wide variation in the crude fibre content of the diets (up to 50 g/kg), as a result of the large quantities of vegetable-protein concentrates needed to supply the desired concentrations of total lysine in the assays (up to 250 g/kg). Amino acid digestibility is negatively correlated with fibre content in cereals (Taverner & Farrell, 1981) and the addition of fibre to diets can depress the digestibility and utilization of lysine and other nutrients (Just, 1982). Thus the high crude fibre contents in some of the diets used by Batterham *et al.* (1981) could have depressed lysine digestibility and hence availability, yielding estimates which may not apply in diets of lower crude fibre content.

In the present paper the results of two assays conducted with pigs to determine the availability of lysine in vegetable-protein concentrates using diets equalized for crude fibre content are reported. Following a very low assay value for a sample of lupin (*Lupinus angustifolius*)-seed meal in Expt 1, additional samples of lupin-seed meal were assayed in Expt 2 to assess the range in lysine availability in this meal. Lysine availability was also estimated by a slope-ratio assay with rats, the Silcock available-lysine technique and by the direct 1-fluoro-2,4-dinitrobenzene (FDNB)-available-lysine assay (Carpenter, 1960).

* Present address: Agricultural Research and Veterinary Centre, Orange, New South Wales 2800, Australia.

Table 1. Composition (g/kg) of the nine protein concentrates

	Expt 1					Expt 2				
	Cotton-seed meal	Lupin (<i>Lupinus angustifolius</i>) meal no. 1 (cv. Uniharvest)	Peanut (groundnut) meal	Soya-bean meal no. 1	Lupin-seed meal no. 2 (cv. Unicrop)	Lupin-seed meal no. 3 (cv. Uniharvest)	Lupin-seed meal no. 4 (cv. Uniharvest)	Field peas (<i>Pisum sativum</i>)	Soya-bean meal no. 2	
Crude protein (nitrogen \times 6.25)	424	337	448	439	309	323	301	216	449	
Dry matter	932	895	913	883	891	906	903	889	891	
Light petroleum (b.p. 40-60°) extract	54	39	60	11	42	54	58	9	17	
Fibre										
Crude	106	137	63	64	153	168	162	33	64	
Acid-detergent	179	174	121	81	223	211	221	76	85	
Neutral-detergent	187	201	141*	112	247	231	237	103	110	
Hemicellulose	8	27	20	31	24	20	16	27	25	
Ash	71	28	46	65	29	30	26	25	59	
Gross energy (MJ/kg)	19.8	18.5	19.2	18.0	18.2	18.4	18.8	18.7	18.0	
Alkaloid	—	≤ 0.14	—	—	≤ 0.1	≤ 0.1	≤ 0.1	—	—	
Essential amino acids										
Threonine	15	11	13	17	11	11	11	8	18	
Valine	19	11	19	20	11	12	11	10	23	
Methionine + cystine	12	3	9	8	6	8	9	5	9	
Isoleucine	13	12	15	20	13	13	13	9	19	
Leucine	27	23	30	35	24	25	25	16	35	
Phenylalanine + tyrosine	36	26	40	37	22	23	23	17	38	
Histidine	12	8	10	13	8	9	9	6	13	
Lysine	18	15	15	28	14	15	14	16	26	
Arginine	45	40	60	41	33	35	34	23	34	

* Difficult to filter.

EXPERIMENTAL

Protein concentrates

The chemical composition of the nine protein concentrates is presented in Table 1. The cottonseed meal and peanut (groundnut) meal were expeller processed and the soya-bean meals 'prepress' solvent extracted. The field peas (*Pisum sativum* cultivar Early Dun) and lupin (cultivars Unicrop and Uniharvest)-seed meal were coarsely crushed through a hammer mill. The lupin-seed meal samples were analysed for alkaloids, aflatoxins and for the presence of *Phomopsis leptostromiformis*, a fungus which produces a mycotoxin on the leaves of lupin plants. As an additional precaution against *P. leptostromiformis* infection, the lupin-seed meals for Expt 2 (nos 2, 3 and 4) were stored at $15 \pm 1^\circ$ and $55 \pm 5\%$ relative humidity before incorporation into diets.

Pig slope-ratio assay

A slope-ratio assay was used to determine the availability of lysine in proteins. Linear regression coefficients of response (say to food conversion efficiency) to increasing dose level of test protein and standard lysine are calculated and the ratio of the test protein's linear regression coefficient to the standard lysine's linear regression coefficient provides the potency of the lysine in the test protein. In our assays the dose levels for the test proteins were formulated to contain the same total lysine as that of the standard lysine doses so that the potency estimate for lysine in the test protein was an expression of lysine availability as a proportion of total lysine. The statistical analysis of the slope-ratio assay was as outlined in Chapter 7 of Finney (1964).

There are a number of criteria that can be used to assess response. Food conversion efficiency is preferred to weight gain as it takes into account any differences in food intake (Carpenter, 1973). Previous work (Batterham *et al.* 1979, 1981) had shown that results expressed on a carcass basis were preferred to those expressed on a live-weight basis as the former eliminates any effect of variation in gut content on availability estimates. This is important, as in the formulation of diets the test protein was included at the expense of highly digestible wheat starch.

Expt 1. For this assay, four protein concentrates were assayed in the one experiment. This involved the use of twenty-six diets: the basal diet (blanks), five diets to determine the pig's response to standard lysine and twenty for the four protein concentrates (five for each protein concentrate). The basal diet (Tables 2 and 3) was formulated using a medium-protein wheat (Timgalen cultivar) which, in combination with the wheat gluten, supplied adequate quantities of all the amino acids except lysine, which was added to bring the basal level up to 5.2 g/kg, and methionine and threonine, which were added to ensure adequacy according to estimates of Lewis & Cole (1976). The five levels of lysine used to determine the pig's response to standard lysine were in 500 mg increments of L-lysine/kg and were obtained by the addition to the basal diet of L-lysine monohydrochloride, anhydrous, feed grade, supplied by Miwon Co. Ltd, Korea. The protein concentrates were incorporated into the basal diets to provide five levels of total lysine, again in 500 mg/kg increments, at the expense of wheat starch. The cottonseed meal contained 15100 and 940 mg total and free gossypol/kg respectively and ferrous sulphate was added to inactivate any effects it may have had (Husby & Kroening, 1971).

Solka floc was included in the basal diet and the level reduced to make allowance for the crude fibre content of the vegetable-protein concentrates so that all diets were fed on an equal crude fibre basis. Maize oil was included in the basal diet to improve the texture of the diet. The digestible energy content of the components was calculated using results of previous determinations at this Agricultural Research Centre or literature values. Dietary

Table 2. *Composition (g/kg) of the basal diets used for the slope-ratio assays with pigs*

	Expt 1	Expt 2
Wheat	670	650
Wheat gluten	100	90
L-Lysine monohydrochloride	1.54	1.79
DL-Methionine	0.70	0.30
L-Threonine	0.60	0.70
Mineral and vitamin premix*	5.5	5.5
Bone meal	30	30
Solka floc	32	39
Maize oil	20	25
Wheat starch	139.66	157.71

* Contributed the following (per kg diet): iron 60 mg, zinc 100 mg, manganese 30 mg, copper 5 mg, iodine 2 mg, selenium 150 µg, sodium chloride 2.5 g, retinol equivalent 960 µg, cholecalciferol 12 µg, α -tocopherol 20 mg, thiamin 1 mg, riboflavin 3 mg, nicotinic acid 12 mg, pantothenic acid 10 mg, pyridoxine 1.5 mg, cyanocobalamin 15 µg, pteroylmonoglutamic acid 2 mg, choline 500 mg, ascorbic acid 10 mg, biotin 100 µg, arsenilic acid 9 mg.

Table 3. *Composition (g/kg) of the wheat, wheat gluten and basal diets used for the growth assays with pigs*

	Expt 1			Expt 2		
	Wheat	Wheat gluten	Basal diet	Wheat	Wheat gluten	Basal diet
Crude protein (nitrogen \times 6.25)	147	814	179	156	841	177
Dry matter	905	923	905	904	925	903
Light petroleum (b.p. 40–60°) extract	17	7	34	14	6	41
Fibre						
Crude	23	0	38	25	0	44
Acid-detergent	42	—*	55	45	—*	64
Neutral-detergent	96	3	94	105	5	105
Hemicellulose	54	—	39	60	—	41
Essential amino acids						
Threonine	4.3	21	5.6	4.4	22	5.6
Valine	6.0	28	6.8	6.3	32	7.0
Methionine + cystine	4.4	30	6.7	4.6	39	6.8
Isoleucine	5.0	28	6.2	5.2	29	6.0
Leucine	9.7	57	12.2	10.2	62	12.2
Phenylalanine + tyrosine	10.6	63	13.4	10.9	65	12.9
Histidine	3.2	17	3.8	3.1	19	3.7
Lysine	4.0	13	5.2	3.9	14	5.2
Arginine	7.5	39	8.9	7.5	27	7.3

* Assay appeared inapplicable as abnormally high values recorded (198 and 161 g/kg), possibly due to high glutamic acid content in wheat gluten.

energy was maintained at 14.5 MJ digestible energy/kg diet using wheat starch and maize oil as non-protein energy sources.

The twenty-six diets were arranged in a randomized block design. The pigs were blocked on 7-week weight, sex and position in the experimental facilities. There were four blocks, two containing males and two females, all of the Large White breed. The pigs were penned individually and water supplied by 'nipple' drinkers. Dietary treatments were introduced when the pigs reached 20 kg live weight.

Table 4. *Expt 1. Analysis of variance*

Source of variation	Degrees of freedom	Source of variation	Degrees of freedom
Lysine (linear)	1	Curvature	
Test 1 (linear)	1	Lysine	3
Test 2 (linear)	1	Test 1	3
Test 3 (linear)	1	Test 2	3
Test 4 (linear)	1	Test 3	3
Blanks	1	Test 4	3
Intersection		Error	78
Test 1-lysine	1	Total	103
Test 2-lysine	1		
Test 3-lysine	1		
Test 4-lysine	1		

The diets were offered at a daily rate of 1000 g at 20 kg live weight, with 100 g increments/2.5 kg live-weight gain. The pigs were fed eight times daily, at intervals of 3 h, with a solenoid-controlled automatic frequent feeder to ensure the utilization of added free amino acids (Batterham & Murison, 1981). The food was offered dry. Rations were adjusted after the weekly weighings of the pigs.

The pigs were slaughtered after reaching a minimum weight of 45 kg and hot eviscerated carcass weights recorded. The ham was dissected and the lean content used as an indicator of carcass leanness. Pig response was assessed in terms of daily live-weight gain, food conversion efficiency (FCE; kg live-weight gain/kg food eaten), dressing proportion (hot carcass weight as a proportion of live weight), lean content of the ham, carcass gain/d (kg hot carcass weight - (kg initial live weight \times 0.69) \div period (d) on experiment) and FCE on a carcass basis (kg hot carcass weight - (kg initial live weight \times 0.69) \div kg food intake). The factor of 0.69 for estimated carcass weight was previously determined with ten piglets (five males and five females) slaughtered at 20 kg live weight.

The results for daily live-weight gain, FCE, carcass gain/d and FCE on a carcass basis were analysed by the slope-ratio technique of Finney (1964) for multiple assays. The analysis of variance was as shown in Table 4. The error degrees of freedom were reduced to seventy-six as there were two missing plots (see Results section).

The results for dressing proportion and lean content of the hams were regressed *v.* lysine for each protein concentrate. These analyses were conducted to determine if there was any effect of inclusion level of protein concentrate on dressing proportion or dietary lysine concentration on lean deposition.

Expt 2. This was conducted in a similar manner to Expt 1 except that five protein concentrates were assayed. This involved the use of thirty-one diets: the basal diet (blanks), five diets to determine the pig's response to standard lysine and twenty-five for the five protein concentrates (five/protein concentrate). The basal diet (Tables 2 and 3) was formulated using a medium-protein wheat (Condor cultivar) which, in combination with the wheat gluten, supplied adequate quantities of all the amino acids except lysine, which was added to bring the basal level up to 5.2 g/kg, and methionine and threonine, which were added to ensure adequacy according to estimates of Lewis & Cole (1976).

Inclusion levels of lysine, equalization of dietary crude fibre and digestible energy content were as in Expt 1 except that the digestible energy content of the field peas and soya-bean meal no. 2 (13.9 and 14.8 MJ/kg, air-dry basis) were determined by the method outlined by Batterham (1979).

The design of the experiment and allocation of animals was as for Expt 1 with four pigs allotted to each diet except for diets 1 (blanks) and 6 (2500 mg standard lysine/kg) where six pigs were allotted (three males and three females). Feeding rates, method of procedure, assessment of performance and statistical analyses of the results were as for Expt 1 except that with the extra protein concentrate and additional pigs on diets 1 and 6 there were ninety-seven degrees of freedom in the analysis of variance.

Rat slope-ratio assay

Single separate assays were conducted for each protein concentrate. A total of seven diets were used for each assay: the basal diet (blanks), three diets to determine the rat's response to standard lysine and three diets to determine the rat's response to the protein concentrate. The basal diet contained (g/kg): wheat 650, wheat gluten 100, DL-methionine 1.5, L-threonine 0.7, maize oil 20, bone flour 25, mineral and vitamin premix 5 (composition in Table 2) and wheat starch 197.8. The combination of wheat and gluten supplied adequate levels of all amino acids except lysine (4.7 mg/kg), methionine and threonine. The latter two were added to ensure adequacy according to estimates of the (US) National Research Council (1972). The three levels of L-lysine used to determine the rat's response to standard lysine were 0.75, 1.5 and 2.25 g/kg (same batch of lysine as used for the pig assay). The protein concentrates were incorporated into the diets to supply the same three levels of total lysine as used to determine the standard lysine response. This was done at the expense of wheat starch. Solka floc was incorporated into the diets to equalize dietary crude fibre content to that supplied by the highest level of protein concentrate. Additional maize oil was used with some protein concentrates to maintain the estimated digestible energy content of the diets.

For the rat assays, two female and two male albino rats, approximately 24–26 d old, were used per dose and were blocked on the basis of litter and sex (block size seven). The rats were individually caged in a room where the temperature and relative humidity were maintained at $21 \pm 1^\circ$ and $50 \pm 5\%$ respectively. Lighting was provided for 12 h daily. Food was supplied in 'self-feeders'.

At the completion of a 14 d test, the rats were weighed, killed with chloroform and the alimentary tract, heart and lungs removed. The weight of the eviscerated carcass was recorded. Performance was assessed in terms of weight gain, FCE (g weight gain/g food eaten), carcass gain (g eviscerated carcass weight – (g initial live weight \times 0.79)) and FCE on a carcass basis (g eviscerated carcass weight – (g initial live weight \times 0.79) \div g food eaten). The factor of 0.79 for estimated eviscerated carcass weight was previously determined with eight rats (four male and four female) of similar live weight and age to those used for the assays.

The results were analysed by the slope-ratio technique of Finney (1964) for single assays. The analysis of variance for each protein was as follows:

Source of variation	Degrees of freedom
Lysine (linear)	1
Test (linear)	1
Blanks	1
Intersection	1
Curvature of lysine	1
Curvature of test	1
Error	21
Total	27

Table 5. Expt 1.* Live-weight gain, food conversion efficiency and lean content of hams of pigs during the 20–45 kg growth phase when fed on the diets for the slope-ratio assay for lysine

Lysine dose level (g/kg)	Form of lysine addition				
	Free lysine	Cottonseed meal	Lupin (<i>Lupinus angustifolius</i>) seed meal no. 1	Peanut (groundnut) meal	Soya-bean meal no. 1
	Live-wt gain (g/d)				
0	478	—	—	—	—
0.5	447	494	495	465	495
1.0	558	532	514	543	549
1.5	599	486	534	575	608
2.0	597	527	542	555	609
2.5	631	579	548	591	643
	SEM 24				
	Food conversion efficiency†				
0	0.381	—	—	—	—
0.5	0.370	0.375	0.393	0.367	0.388
1.0	0.426	0.413	0.405	0.427	0.418
1.5	0.449	0.402	0.420	0.443	0.457
2.0	0.453	0.412	0.418	0.431	0.471
2.5	0.479	0.445	0.439	0.458	0.485
	SEM 0.012				
	Lean in ham (g/kg)				
0	594	—	—	—	—
0.5	606	593	587	590	568
1.0	609	592	606	583	603
1.5	610	590	610	600	580
2.0	627	599	623	649	626
2.5	633	614	617	609	619
	SEM 12				

* For details, see p. 87.

† Live-weight gain (kg) ÷ food intake (kg).

Chemical analyses

The techniques used were as reported by Batterham *et al.* (1979) except for acid-detergent fibre (Van Soest, 1963), neutral-detergent fibre (Van Soest & Wine, 1967, as modified by King & Taverner, 1975), alkaloids (Ruiz, 1976) and aflatoxins (Romer, 1975).

RESULTS

Performance results of the pigs for Expt 1 are presented in Tables 5 and 6 and for Expt 2 in Tables 7 and 8. Two pigs died in Expt 1 (diets nos 2 and 5) with post-mortem symptoms of *Escherichia coli* infections. There was a smaller amount of feed rejection by most pigs in Expt 1 and lesser amounts by pigs in Expt 2.

Lean in the ham of pigs increased slightly as the level of dietary lysine increased. In Expt 1 the slope of the increase was greater ($P < 0.05$) in pigs given soya-bean meal than with those given cottonseed meal but neither was significantly different ($P > 0.05$) from the standard lysine response. In Expt 2 the slope of the increase in pigs given field peas was

Table 6. *Expt 1.* Dressing proportion, carcass gain and food conversion efficiency on a carcass basis of pigs during the 20–45 kg growth phase when fed on the diets for a slope-ratio assay for lysine*

Lysine dose level (g/kg)	Form of lysine addition				
	Free lysine	Cottonseed meal	Lupin (<i>Lupinus angustifolius</i>) seed meal no. 1	Peanut (groundnut) meal	Soya-bean meal no. 1
	Dressing proportion (g/kg)†				
0	752	—	—	—	—
0.5	770	754	748	769	755
1.0	758	756	742	733	755
1.5	751	746	718	735	748
2.0	762	738	744	738	751
2.5	758	734	736	736	741
	SEM 8				
	Carcass gain (g/d)‡				
0	381	—	—	—	—
0.5	373	397	391	386	398
1.0	450	428	401	415	441
1.5	478	384	394	442	481
2.0	488	408	426	428	487
2.5	509	444	423	454	500
	SEM 18				
	Food conversion efficiency (carcass basis)§				
0	0.304	—	—	—	—
0.5	0.309	0.301	0.311	0.305	0.311
1.0	0.344	0.332	0.316	0.327	0.336
1.5	0.358	0.318	0.310	0.340	0.362
2.0	0.370	0.320	0.328	0.332	0.377
2.5	0.386	0.341	0.339	0.353	0.378
	SEM 0.010				

* For details, see p. 87.

† Hot carcass weight as a proportion of live weight before slaughter.

‡ Hot carcass weight (kg) – (initial live weight (kg) × 0.69) ÷ period (d) on experiment.

§ Hot carcass weight (kg) – (initial live weight (kg) × 0.69) ÷ food intake (kg).

greater and those given soya-bean meal smaller ($P < 0.05$) compared with pigs given the standard lysine or other protein concentrates.

The level of inclusion of protein concentrate depressed dressing proportion in Expt 1, but this was only significant ($P < 0.05$) for cottonseed and peanut meals. In Expt 2 the addition of standard lysine increased dressing proportion ($P < 0.05$), field peas had no effect and the lupin-seed and soya-bean meals had small but non-significant ($P > 0.05$) depressing effects.

The availabilities of lysine in the nine protein concentrates, as determined by the four production criteria, are presented in Table 9. Availability estimates were lower on a carcass compared to live-weight basis and in the majority of cases marginally higher on a FCE compared to weight-gain basis. Using FCE on a carcass basis as the criterion of response, availability estimates were very low in cottonseed meal, low and variable in lupin-seed meal and high in field peas and soya-bean meal.

Table 7. Expt 2.* Live-weight gain, food conversion efficiency and lean content of hams of pigs during the 20–45 kg growth phase when fed on the diets for the slope-ratio assay for lysine

Lysine dose level (g/kg)	Form of lysine addition					
	Free lysine	Lupin (<i>Lupinus angustifolius</i>) seed meal no. 2	Lupin-seed meal no. 3	Lupin-seed meal no. 4	Field peas (<i>Pisum sativum</i>)	Soya-bean meal no. 2
	Live-wt gain (g/d)					
0	493	—	—	—	—	—
0.5	535	541	502	515	523	502
1.0	543	550	510	536	563	541
1.5	587	574	562	572	589	576
2.0	587	555	585	544	564	601
2.5	590	598	566	567	614	631
	SEM 15					
	Food conversion efficiency†					
0	0.371	—	—	—	—	—
0.5	0.397	0.406	0.388	0.383	0.391	0.382
1.0	0.413	0.406	0.391	0.399	0.418	0.406
1.5	0.441	0.429	0.423	0.425	0.441	0.443
2.0	0.440	0.417	0.440	0.424	0.430	0.453
2.5	0.444	0.444	0.428	0.435	0.463	0.475
	SEM 0.015					
	Lean in ham (g/kg)					
0	590	—	—	—	—	—
0.5	585	602	599	601	589	631
1.0	604	608	617	618	606	614
1.5	631	625	632	635	634	638
2.0	640	652	651	655	620	627
2.5	628	636	653	645	671	649
	SEM 9					

* For details, see p. 89.

† Live-weight gain (kg) ÷ food intake (kg).

Rat slope-ratio estimates on the nine protein concentrates are presented in Table 10. There was no consistent pattern of gain *v.* FCE but estimates were generally lower on a carcass basis. Using FCE on a carcass basis as the criterion of response, there was close agreement with the pig estimates for cottonseed and soya-bean meals, higher values for lupin-seed meal and peanut meal and a lower value for field peas.

The Silcock available-lysine values ranged from 0.98 for lupin-seed meal no.1 and field peas to 0.87 for cottonseed meal (Table 11). Values for the direct-FDNB procedure were lower and more variable and ranged from 0.57 for lupin-seed meal no. 1 to 0.94 for soya-bean meal no. 2. For comparative purposes, the slope-ratio estimates for pigs and rats (based on FCE on a carcass basis) are also presented in Table 11. There was little relationship between these values and the chemical techniques.

All samples of lupin-seed meal had low levels of aflatoxins ($\leq 30 \mu\text{g}/\text{kg}$), alkaloids ($\leq 0.10\text{--}0.14 \text{ g}/\text{kg}$) and presence of *P. leptostromiformis* (nos 1 and 4 not detectable, nos 2 and 3 one seed per 200 seeds examined).

Table 8. *Expt 2.* Dressing proportion, carcass gain and food conversion efficiency on a carcass basis of pigs during the 20–45 kg growth phase when fed on the diets for a slope-ratio assay for lysine*

Lysine dose level (g/kg)	Form of lysine addition					
	Free lysine	Lupin (<i>Lupinus angustifolius</i>) seed meal no. 2	Lupin-seed meal no. 3	Lupin-seed meal no. 4	Field peas (<i>Pisum sativum</i>)	Soya-bean meal no. 2
	Dressing proportion (g/kg)†					
0	749	—	—	—	—	—
0.5	746	743	751	749	749	755
1.0	745	742	737	737	740	736
1.5	760	736	738	746	751	739
2.0	759	751	714	731	769	731
2.5	768	729	745	729	736	741
	SEM 10					
	Carcass gain (g/d)‡					
0	392	—	—	—	—	—
0.5	421	422	400	407	413	404
1.0	428	428	395	414	436	417
1.5	475	441	434	450	468	447
2.0	475	442	428	413	467	457
2.5	487	452	444	431	471	490
	SEM 12					
	Food conversion efficiency (carcass basis)§					
0	0.295	—	—	—	—	—
0.5	0.313	0.317	0.310	0.303	0.309	0.307
1.0	0.325	0.317	0.302	0.308	0.324	0.314
1.5	0.357	0.330	0.326	0.335	0.350	0.344
2.0	0.356	0.332	0.322	0.322	0.356	0.344
2.5	0.366	0.336	0.336	0.330	0.356	0.368
	SEM 0.017					

* For details, see p. 89.

† Hot carcass weight as a proportion of live weight before slaughter.

‡ Hot carcass weight (kg) – (initial live weight (kg) × 0.69) ÷ period (d) on experiment.

§ Hot carcass weight (kg) – (initial live weight (kg) × 0.69) ÷ food intake (kg).

DISCUSSION

The results confirm earlier findings (Batterham *et al.* 1979) that the availability of lysine in cottonseed meal is low for pigs, and that of soya-bean meal high. There appears to be little effect of the method of processing on cottonseed meal in that the low availability in the cottonseed meal in Expt 1 (0.39, expeller processed) was similar to that of a 'prepress' solvent-extracted meal (0.43, Batterham *et al.* 1979). The results also indicate that the availability of lysine in peanut meal is low to medium (0.57) and in field peas high (0.93).

The low and variable results for lupin-seed meal were unexpected as no heat processing was used in their preparation. Consequently, an availability similar to field peas (also given raw) was anticipated. The low availabilities were unlikely to be due to the presence of aflatoxins, mycotoxins or alkaloid content. The production history of meal no. 1 (availability 0.37) was traced and there was no evidence of pesticides or chemicals used during

Table 9. Expts 1 and 2. Availability of lysine (proportion of total) in the protein concentrates as assessed with pigs using daily live-weight gain, food conversion efficiency (FCE), daily carcass gain and FCE on a carcass basis as the criteria for availability
(Mean values and standard deviations)

Protein concentrate	Daily live-wt gain (kg)		FCE*		Daily† carcass gain		FCE‡ (carcass basis)	
	Mean	SD	Mean	SD	Mean	SD	Mean	SD
Expt 1								
Cottonseed meal	0.51	0.12	0.57	0.09	0.36	0.11	0.39	0.09
Lupin (<i>Lupinus angustifolius</i>) seed meal no. 1	0.51	0.12	0.62	0.09	0.31	0.11	0.37	0.09
Peanut (groundnut) meal	0.74	0.12	—§	0.10	0.54	0.11	0.57	0.09
Soya-bean meal no. 1	1.08	0.14	1.10	0.12	0.97	0.12	0.98	0.11
Expt 2								
Lupin-seed meal no. 2	0.94	0.11	0.89	0.10	0.68	0.09	0.65	0.08
Lupin-seed meal no. 3	—§	—	0.82	0.10	0.49	0.09	0.54	0.08
Lupin-seed meal no. 4	0.74	0.10	0.81	0.10	0.48	0.09	0.54	0.08
Field peas (<i>Pisum sativum</i>)	1.06	0.12	1.08	0.11	0.91	0.10	0.93	0.09
Soya-bean meal no. 2	—§	—	—§	—	0.85	0.10	0.89	0.09

* Live-weight gain (kg) ÷ food intake (kg).

† Hot carcass weight (kg) - (initial live weight (kg) × 0.69) ÷ period (d) on experiment.

‡ Hot carcass weight (kg) - (initial live weight (kg) × 0.69) ÷ food intake (kg).

§ Slope-ratio values not calculated as curvature (peanut meal) and intersection (lupin-seed meal no. 3 and soya-bean meal no. 2) significant ($P \leq 0.05$).

Table 10. Availability of lysine (proportion of total) in the protein concentrates as assessed with rats using live-weight gain, food conversion efficiency (FCE), carcass gain and FCE on a carcass basis as the criteria for availability
(Mean values and standard deviations)

Protein concentrate	Live-wt gain		FCE*		Carcass gain†		FCE‡ (carcass basis)	
	Mean	SD	Mean	SD	Mean	SD	Mean	SD
Cottonseed meal	0.35	0.11	0.47	0.10	0.26	0.10	0.35	0.09
Lupin (<i>Lupinus angustifolius</i>) seed meal no. 1	0.96	0.12	0.93	0.14	0.96	0.29	0.81	0.08
Peanut (groundnut) meal	0.87	0.15	0.92	0.12	0.74	0.10	0.76	0.09
Soya-bean meal no. 1	0.80	0.14	1.00	0.15	0.78	0.13	0.91	0.10
Lupin-seed meal no. 2	1.02	0.05	1.05	0.22	0.93	0.07	0.94	0.13
Lupin-seed meal no. 3	1.07	0.15	0.80	0.13	0.96	0.15	0.70	0.13
Lupin-seed meal no. 4	1.13	0.22	0.97	0.17	0.90	0.15	0.80	0.19
Field peas (<i>Pisum sativum</i>)	0.76	0.07	—§	—	0.74	0.08	0.76	0.08
Soya-bean meal no. 2	0.85	0.13	0.90	0.10	0.91	0.12	0.89	0.11

* Live-weight gain (g) ÷ food intake (g).

† Hot eviscerated weight (g) - (initial live weight (g) × 0.79)

‡ Hot eviscerated weight (g) - (initial live weight (g) × 0.79) ÷ food intake (g).

§ Slope-ratio values not calculated as responses were not linear ($P > 0.05$).

Table 11. Total lysine (g/kg) and the availability of lysine (proportion of total) in the protein concentrates as assessed by the chemical Silcock technique (Roach et al. 1967), the direct 1-fluoro-2,4-dinitrobenzene (FDNB) assay (Carpenter, 1960) and by the slope-ratio assay with pigs and rats using food conversion efficiency on a carcass basis as the criterion for availability (Mean values and standard deviations)

Protein concentrate	Total lysine		Silcock assay	Direct FDNB assay	Slope-ratio assay			
	Mean	SD			Pigs		Rats	
					Mean	SD	Mean	SD
Cottonseed meal	18	0.03	0.87	0.83	0.39	0.09	0.35	0.09
Lupin (<i>Lupinus angustifolius</i>) seed meal								
No. 1	15	0.07	0.96	0.57	0.37	0.09	0.81	0.08
No. 2	14	0.03	0.98	0.68	0.65	0.08	0.94	0.13
No. 3	15	0.03	0.97	0.76	0.54	0.08	0.70	0.13
No. 4	14	0.02	0.97	0.90	0.54	0.08	0.80	0.19
	—	—	—	—	0.53	0.04	0.81	0.06
Peanut (groundnut) meal	15	0.03	0.91	0.80	0.57	0.09	0.76	0.09
Field peas (<i>Pisum sativum</i>)	16	0.01	0.98	0.83	0.93	0.09	0.76	0.08
Soya-bean meal no. 1	28	0.04	0.94	0.82	0.98	0.11	0.91	0.10
Soya-bean meal no. 2	26	0.09	0.93	0.94	0.89	0.09	0.89	0.11

production or storage that was likely to affect pig performance. There does not appear to be a toxic factor in the meal as a high inclusion level of meal no. 2 (430 g/kg) was given to weaner pigs with no adverse effect (Barnett & Batterham, 1981) and the response to inclusion level of the meals in the slope-ratio assay was linear. Nor is there evidence in the literature of anti-nutritional factors associated with cultivars of *L. angustifolius* (Hove et al. 1978; Hove & King, 1979; Hudson, 1979).

The true digestibility of lysine at the terminal ileum of pigs was determined for lupin-seed meal no. 1 (availability 0.37) by M. R. Taverner (personal communication) and was high (0.86). This indicates that the low availability is not associated with low digestibility. Nor was the low availability detected by the slope-ratio assays with rats or by the two chemical techniques. Whilst the values for the direct-FDNB procedure were lower than the Silcock values for lupin-seed meal, this may only be a reflection of the instability of dinitrophenyl-lysine in the presence of carbohydrates. Over-all, the lower availability of lysine in the lupin-seed meal for pigs may reflect either the presence of an unidentified growth depressant (which has a linear effect on performance with increasing inclusion level) or to the lysine being in a form that is digested but inefficiently utilized. Furthermore, the causal factor (or factors) appears specific to pigs. These aspects are currently being investigated.

Whilst crude fibre does not measure any definable chemical fraction of a feed (Van Soest & Wine, 1967) it does appear to be a suitable basis on which to equalize diets for fibre content using solka floc, as variation in dietary acid-detergent and neutral-detergent fibre levels was also much reduced (levels of 55–63 and 94–104 g/kg in Expt 1 and 64–70 and 105–115 g/kg in Expt 2 respectively). Nor did equalizing diets for fibre content affect the lowering of availability estimates based on carcass relative to live-weight performance. A limited number of viscera (including the trachea but excluding the kidneys) were examined and this indicated that whilst there was a slight increase in the weight of viscera minus gut contents in some pigs given the protein concentrates, the main effect was due to an increase in gut contents. In Expt 2 there was also an increase in the dressing proportion in pigs given standard

lysine and this contributed to the lowering of availability estimates when based on a carcass basis. The reason for such an effect is unclear; it did not occur in Expt 1 or in previous slope-ratio assays (Batterham *et al.* 1979, 1981). It is desirable to express availability estimates on a carcass basis as it avoids the effect of variation in the weight of viscera and gut contents. It is also desirable to examine the effect of expressing availability estimates on a protein deposition basis to determine if the small differences in the slopes for lean in the ham of pigs given the different protein concentrates reflected differences in availability.

The slope-ratio estimate of 0.98 for lysine availability in soya-bean meal no. 1 (Table 9) was higher than the estimate for soya-bean meal no. 2 (0.89, Table 9) and that of a previously reported soya-bean meal (0.84; Batterham *et al.* 1979). The high value for soya-bean meal no. 1 may reflect variation in the availability estimate as the standard deviation was 0.11; it may also reflect variation in the total lysine determination. Whilst the standard deviation of the total lysine estimate was small (0.04), this reflects only within-laboratory variation, and between-laboratory variation may be greater (Porter *et al.* 1968; Williams *et al.* 1980). However, the estimates of lysine availability that were greater than 1.0 for a number of meals when gain and FCE were assessed on a live-weight basis (Tables 9 and 10) do not appear to be due to underestimation of total lysine but rather to overestimation of the availability estimates (due to the effect of variation in dressing proportion, as discussed earlier). The estimates on a carcass basis were considerably lower for these meals and less than 1.0.

The slope-ratio estimates with rats indicated that for some meals (cottonseed and soya-bean meal) there is a close agreement with the pig estimates. This also applies for sunflower meal (Batterham *et al.* 1981). For others, such as lupin-seed meal, the rat assay is inapplicable for predicting pig response. Additional assays need to be made with peanut meal and field peas before firm conclusions for these meals can be made. As in previous work (Batterham *et al.* 1981), there appeared to be little relationship between the values from the two chemical procedures and the slope-ratio values. Until the cause (or causes) of the variation in available lysine in the different vegetable proteins for pigs and rats is elucidated it is difficult to suggest suitable *in vitro* or *in vivo* techniques for predicting lysine availability in these meals for pigs.

The authors thank Messrs N. R. Thompson, A. W. Davis, H. M. Essery and E. R. Layton for management of the pigs and skilled technical assistance, Mr R. F. Lowe for chemical analyses, Dr M. R. Taverner, Animal Research Institute, Werribee, for the determination of the ileal digestibility of lysine in lupin-seed meal no. 1, Dr H. S. Saini, Agricultural Research Institute, Wagga Wagga, for the alkaloid analyses, Mr E. J. Major, Chemist, Poultry Research Station, Seven Hills, for the aflatoxin analyses and Mr P. M. Woods, Plant Pathologist, Department of Agriculture, Western Australia, for the examination of the lupin seeds for *P. leptostromiformis*.

REFERENCES

- Barnett, C. W. & Batterham, E. S. (1981). *Animal Feed Science and Technology* **6**, 27–34.
 Batterham, E. S. (1979). *Australian Journal of Agricultural Research* **30**, 369–375.
 Batterham, E. S. & Murison, R. D. (1981). *British Journal of Nutrition* **46**, 87–92.
 Batterham, E. S., Murison, R. D. & Lewis, C. E. (1979). *British Journal of Nutrition* **41**, 383–391.
 Batterham, E. S., Murison, R. D. & Lowe, R. F. (1981). *British Journal of Nutrition* **45**, 401–410.
 Carpenter, K. J. (1960). *Biochemical Journal* **77**, 604–610.
 Carpenter, K. J. (1973). *Nutrition Abstracts & Reviews* **43**, 423–451.
 Finney, D. J. (1964). *Statistical Method in Biological Assay*, 2nd ed. London: Griffin.
 Hove, E. L. & King, S. (1979). *New Zealand Journal of Agricultural Research* **22**, 41–42.
 Hove, E. L., King, S. & Hill, G. D. (1978). *New Zealand Journal of Agricultural Research* **21**, 457–462.
 Hudson, B. J. F. (1979). *Qualitas Plantarum—Plant Foods for Human Nutrition* **29**, 245–251.
 Husby, F. M. & Kroening, G. H. (1971). *Journal of Animal Science* **33**, 592–594.
 Just, A. (1982). *Livestock Production Science* **9**, 717–729.

- King, R. H. & Taverner, M. R. (1975). *Animal Production* **21**, 275–284.
- Lewis, D. & Cole, D. J. A. (1976). *Proceedings of the Nutrition Society* **35**, 87–91.
- National Research Council (1972). *Nutrient Requirements of Laboratory Animals*, 2nd ed. Washington, DC: National Academy of Sciences.
- Porter, J. W. G., Westgarth, D. R. & Williams, A. P. (1968). *British Journal of Nutrition* **22**, 437–450.
- Roach, A. G., Sanderson, P. & Williams, D. R. (1967). *Journal of the Science of Food and Agriculture* **18**, 274–278.
- Romer, T. R. (1975). *Journal of the Association of Official Analytical Chemists* **58**, 500–506.
- Ruiz, L. P. Jr (1976). *New Zealand Journal of Agricultural Research* **20**, 51–52.
- Taverner, M. R. & Farrell, D. J. (1981). *British Journal of Nutrition* **46**, 181–192.
- Van Soest, P. J. (1963). *Journal of the Association of Official Agricultural Chemists* **46**, 829–835.
- Van Soest, P. J. & Wine, R. H. (1967). *Journal of the Association of Official Analytical Chemists* **50**, 50–55.
- Williams, A. P., Hewitt, D., Cockburn, J. E., Harris, D. A., Moore, R. A. & Davies, M. G. (1980). *Journal of the Science of Food and Agriculture* **31**, 474–480.