

Leptin and insulin growth factor 1: diagnostic markers of the refeeding syndrome and mortality

Manal O. Elnenaei¹, Jamshid Alaghaband-Zadeh¹, Roy Sherwood¹, Mahmoud A. Awara², Cajé Moniz¹ and Carel W. le Roux^{1,3*}

¹Clinical Biochemistry, King's College Hospital, Denmark Hill, London SE5 9RS, UK

²Mental Health Unit, Basildon Hospital, South Essex Partnership University NHS Foundation Trust, Essex SS16 5NL, UK

³Imperial Weight Centre, Imperial College London, London W6 8RF, UK

(Received 19 November 2010 – Revised 12 January 2011 – Accepted 7 February 2011 – First published online 4 May 2011)

Abstract

Refeeding syndrome is difficult to diagnose since the guidelines for identifying those at risk are largely based on subjective clinical parameters and there are no predictive biochemical markers. We examined the suitability of insulin-like growth factor 1 (IGF1) and leptin as markers to identify patients at risk of the refeeding syndrome before initiation of parenteral nutrition (PN). A total of thirty-five consecutive patients referred for commencement of PN were included. Serum leptin and IGF1 were measured before starting PN. Electrolytes, liver and renal function tests were conducted before and daily for 1 week after initiating PN. The primary outcome was a decrease in phosphate 12–36 h after initiating PN. 'Refeeding index' (RI) was defined as leptin \times IGF1 divided by 2800 to produce a ratio of 1.0 in patients who are well nourished. RI had better sensitivity (78%; 95% CI 40, 97%) and specificity (78%; 95% CI 40, 97%) with a likelihood ratio of 3.4, at a cut-off value of 0.19 for predicting a $\geq 30\%$ decrease in phosphate concentration within 12–36 h after starting PN, compared with IGF1 or leptin alone. However, IGF1 was a better predictor of mortality than either leptin or the RI. The present study is the first to derive and test the 'RI', and find that it is a sensitive and specific predictor of the refeeding syndrome in hospitalised patients before starting PN.

Key words: Refeeding syndrome; Insulin-like growth factor 1; Leptin

Refeeding syndrome is a potentially fatal condition caused by rapid or excessive administration of feeding, whether oral, enteral or parenteral, after a period of relative or absolute starvation⁽¹⁾. A constellation of metabolic disturbances with the development of severe fluid and electrolyte imbalances, along with neurological, pulmonary, cardiac, neuromuscular and haematological complications, could result from the refeeding syndrome^(2,3). Hospitalised patients are those most at risk of developing this problem, while the true incidence remains unknown.

The hallmark biochemical feature of the refeeding syndrome is hypophosphataemia, but the syndrome may also feature abnormal Na and fluid balance, hypokalaemia, hypomagnesaemia, and changes in glucose, protein and fat metabolism⁽⁴⁾. Carbohydrate metabolism requires thiamin that is not stored in large amounts in the body, hence heart failure and encephalopathy secondary to thiamin deficiency may be precipitated by refeeding^(2,3). These features are often subtle before initiation of feeding, but once feeding is started, these may escalate into life-threatening biochemical abnormalities with major fluid and electrolyte shifts⁽³⁾. Morbidity and

mortality from the refeeding syndrome may be significantly reduced if reliable predictive markers could be identified. The evidence base for predicting the refeeding syndrome is poor and consists mainly of cohort studies, case series and consensus expert opinion⁽⁵⁾. It is not surprising therefore that the UK National Institute of Clinical Excellence (NICE) guidelines on the identification of patients at high risk of refeeding problems include recommendations based on expert opinion and 'good clinical practice'⁽⁶⁾. The difficulty in predicting those at high risk is further compounded by the need for a thorough and precise history obtained from the patient, spanning the preceding 3–6 months. Hospitalised patients may often be either too confused or simply unable to provide an accurate history. The NICE criteria also have the potential to identify a number of patients as at risk when they may not be, thus delaying the establishment of full nutritional support.

The well-known case of a healthy performance artist who did not consume any energy for 44 d revealed that while most of his biochemical measurements were unchanged before refeeding, there were striking alterations in leptin and

Abbreviations: IGF1, insulin-like growth factor 1; NBM, nil by mouth; NICE, National Institute of Clinical Excellence; PN, parenteral nutrition; RI, refeeding index.

* **Corresponding author:** Dr C. W. le Roux, fax +44 203 299 3140, email c.leroux@imperial.ac.uk

insulin-like growth factor 1 (IGF1) concentrations⁽⁷⁾. This is not surprising as several nutritional factors regulate leptin and IGF1 production^(8,9). Both decline significantly in conditions of starvation⁽⁷⁾, through potentially independent mechanisms⁽¹⁰⁾. Our hypothesis was that concentrations of leptin and IGF1 decline more significantly in patients who are at risk of the refeeding syndrome, and that, together, leptin and IGF1 could provide a useful 'refeeding index' (RI) as a predictive tool for this potentially serious and difficult to diagnose condition.

Materials and methods

Between January and April 2009, new inpatients referred to our team for initiation of parenteral nutrition (PN) were recruited. Exclusion criteria comprised patients who were on oral or enteral feeding at the same time as when the parenteral feed was started, as well as those who had already been commenced on PN before being referred to us. None of the patients received dextrose or Ca infusions with PN. The total number of patients eligible for the present study was thus thirty-five. The present study was conducted according to the guidelines laid down in the Declaration of Helsinki, and all procedures involving patients were approved by the King's College Hospital Ethics Committee (Denmark Hill, London, UK). Verbal informed consent was obtained from all patients, and this was witnessed and formally recorded. The study has been registered in the ClinicalTrials.gov registry. The registration number is NCT01227850. Routine blood samples were taken between 06.00 and 09.00 hours in the morning before initiation of PN. Patients were assessed for nutritional status to determine whether or not they were malnourished, according to the NICE guidelines⁽⁶⁾, while nutritional intake and the number of days a patient was nil by mouth (NBM) were recorded. The Schofield equation⁽¹¹⁾ was used to estimate BMR and adjusted for activity and stress to give the approximate energy requirements⁽¹²⁾. Patients were identified to be at 'high risk' of developing refeeding problems according to the guidelines set out by the NICE⁽⁶⁾ for identifying such patients. These patients were started on 25% of their daily energy requirements for the first 48 h with gradual increase to meet or exceed requirements by 4–7 d. Patients who were NBM for more than 5 d or clinically malnourished according to the NICE guidelines (either BMI <18.5 kg/m² or unintentional weight loss >10% within the last 3–6 months, or BMI <20 kg/m² and unintentional weight loss >5% within the last 3–6 months)⁽⁶⁾ were commenced on 50% of their daily energy requirements for the first 48 h⁽⁶⁾. Feeding rates were then increased to meet the nutritional needs, provided the patients did not show a significant decrease in phosphate, K or Mg. Kabiven[®] PN bags or part bags (Fresenius Kabi Limited, Runcorn, Cheshire, UK) were selected based on each patient's energy requirements with the addition of electrolytes, vitamins and minerals as required either in the PN bag or as separate infusions.

Blood tests were repeated daily between 08.00 and 09.30 hours, for at least a week once PN was started. Samples

were analysed for electrolytes (including phosphate, K or Mg), liver and renal function on the Advia 2400 (Siemens Healthcare Diagnostics, Camberley, Surrey, UK). Patients were subsequently followed up for a period of 1 month to determine mortality.

Insulin-like growth factor 1 and leptin measurements

Serum was separated and stored frozen at –20°C. IGF1 was analysed using an immunochemiluminescent assay run on the Immulite 2000 (Siemens Healthcare Diagnostics). The intra- and inter-assay CV were less than 8%. Since the lowest limit of detection for the IGF1 assay was 25 µg/l, we considered any value of less than this for purposes of analysis as 25 µg/l. Leptin was measured using an ELISA kit (Diagnostic Systems Laboratories, Inc., Webster, TX, USA). The intra-assay CV for leptin was less than 5% at all concentrations.

Deriving the 'refeeding index'

Age- and sex-unified mean values for leptin and IGF1 were calculated from reference ranges used in our laboratory^(13,14). This was done by deriving the means of the given age- and sex-adjusted reference ranges for each analyte, and then by calculating their mean values. This value was 21 µg/l for leptin and 134 µg/l for IGF1. The product of these two numbers is 2800 (rounded to two significant figures), which forms the basis for the denominator to derive a 'RI', defined as the product of leptin and IGF1 divided by 2800; in well-nourished patients, this should give a value of approximately 1.0. This index was therefore derived in a similar way to the homeostatic model assessment for insulin resistance index, which is based on multiplying the glucose concentration by the insulin concentration and then dividing by a factor (product of mean glucose and insulin values), which normalises to 1.0⁽¹⁵⁾.

Statistics

Statistical analysis was conducted using 'Analyse-It' version 2.21 (Analyse-It Limited, Leeds, UK). Receiver-operator curves were constructed; hence, sensitivity and specificity were obtained for each of IGF1, leptin and RI against a ≥30% phosphate drop as well as against death within 1 month of starting PN.

As the leptin and IGF1 data were not normally distributed, non-parametric statistical tests were used for all data analyses. The Mann–Whitney test was used to determine the relationship between IGF1, leptin and the RI and other variables such as patient's sex, days NBM ≥5 d, whether or not a patient was clinically malnourished and mortality. The Kruskal–Wallis ANOVA test was used to assess the association of the aforementioned parameters, as well as death, against patient location and estimated glomerular filtration rate, which consisted of three categories as given in Table 1. The association of the three parameters (IGF1, leptin and the RI) with age was determined using the Spearman rank correlation test, while Fisher's exact test was used to compare phosphate

Table 1. Summary of the patient data

Patient data	Frequency
Sex	
Male	19
Location	
General ward	17
Other ITU	5
Liver ITU	13
Phosphate drop (%)	
< 30	26
≥ 30	9
Days nil by mouth	
< 5	20
≥ 5	11
Unknown	4
Clinically malnourished	
No	15
Yes	15
Unsure	5
GFR (ml/min)	
< 40	4
40–60	10
> 60	21
Died	
Yes	8
No	27

ITU, intensive treatment units; GFR, glomerular filtration rate.

drop with mortality, whether a patient was malnourished and NBM ≥ or < 5 d.

Results

Demographics and clinical evaluation

A total of thirty-five patients, nineteen males and sixteen females, mean age 53 (SEM 3.29; range 18–83) years, were included in the present study. Of these patients, thirteen were admitted to a liver intensive treatment unit, five to other intensive treatment units and seventeen patients were from various non-intensive therapy wards.

We identified seven patients as at 'high risk' of refeeding problems. In total, eleven patients were NBM for 5 d or more, while four other patients had an unclear energy intake for the weeks preceding admission. After clinical examination, fifteen patients were deemed to be malnourished, while five other patients were difficult to categorise, based on vague history and inconclusive clinical evaluation.

Table 1 demonstrates that eight patients died (five from liver intensive treatment unit, two from general wards and one from other intensive treatment units); seven patients died within 10–14 d of initiation of PN, while one died 4 weeks after PN was started. None of the patients developed clinical features of the refeeding syndrome.

Electrolyte trends

Despite supplementation of phosphate, K or Mg in anticipation of suspected decreases after starting PN, we observed phosphate concentrations decreasing significantly between 12 and 36 h of starting PN in nine patients. The phosphate

decrease in these patients was observed to be >30% and fell below the lower limit of our laboratory's reference range of 0.80 mmol/l in all nine patients. The other patients had either no decrease of phosphate or a ≤21% decrease. Of the nine patients who exhibited a significant decrease in phosphate, five were identified before starting PN as at risk of the refeeding syndrome based on either being NBM for ≥5 d and/or being malnourished on clinical assessment. On the other hand, fourteen patients who were defined as either clinically malnourished⁽⁶⁾ and/or NBM for ≥5 d showed no significant decrease in their phosphate concentrations after starting PN. Table 2 shows the mean values and standard deviations of phosphate concentrations for all patients and for patients who died, before and after starting PN, with those who had a significant decrease in serum phosphate compared with the group with no significant decrease. K, Mg and Na showed minimal variations in the week following initiation of PN.

Leptin, insulin-like growth factor 1 and the refeeding index

Table 3 shows the results for leptin, IGF1 and the RI (calculated from the first two parameters as described earlier in the Materials and methods section). Relevant clinical details

Table 2. Comparison of phosphate concentrations in all patients *v.* those who died, with either a ≥30% or a <30% drop in phosphate before and after starting parenteral nutrition (PN), against the refeeding index measured before starting PN†

(Mean values, standard deviations, ranges and number of patients)

	Patients with a ≥30% phosphate decrease	Patients with a <30% phosphate decrease
Number of all patients	9	26
Phosphate before starting PN		
Range (mmol/l)	0.65–1.56	0.7–1.55
Mean (mmol/l)	1.08	1.1
SD	0.3	0.23
Phosphate after starting PN		
Range (mmol/l)	0.38–0.73	0.54–1.88
Mean (mmol/l)	0.55	1.05*
SD	0.12	0.35
Refeeding Index		
Range	0.06–0.64	0.02–3.00
Mean (mmol/l)	0.20	0.53*
SD	0.20	0.64
Number of patients who died	4	4
Phosphate before starting PN		
Range (mmol/l)	0.74–1.38	0.69–1.25
Mean (mmol/l)	0.98	1.00
SD	0.28	0.25
Phosphate after starting PN		
Range (mmol/l)	0.49–0.73	0.67–1.76
Mean (mmol/l)	0.59	1.10*
SD	0.10	0.48
Refeeding index		
Range	0.06–0.42	0.04–0.26
Mean (mmol/l)	0.17	0.18
SD	0.17	0.10

* Mean values were significantly different ($P < 0.05$; *t* test).

† Reference interval for phosphate is 0.80–1.40 mmol/l.

Table 3. Clinical details of individual patients, as well as results for insulin-like growth factor 1 (IGF1), leptin, the refeeding index (RI) and those who had a $\geq 30\%$ phosphate drop on day 2 or 3 of starting parenteral nutrition

Sex	Age (years)	Location	IGF1 ($\mu\text{g/l}$)	Leptin ($\mu\text{g/l}$)	RI	Maln	NBM (d)	Weight (kg)/BMI (kg/m^2)	PO ₄	Diagnosis	Death	Cause*	Time (d)†
M‡	19	Ward	25	13	0.12	Yes	7	39/20	Yes	AIH	No		
F‡	77	Ward	33	13	0.15	Yes	10	52/19	Yes	SBO	No		
M‡	83	Ward	35	7	0.09	Yes	10	49/16	Yes	Cholangio CA	No		
F	58	LITU	25	8	0.07	Yes	2	–	Yes	ALD	Yes	ALF	10
F	58	LITU	75	33	0.88	No	0	–	No	Venlafaxine OD	No		
M	59	ITU	89	11	0.35	No	0	–	No	SB perforation	No		
M	18	Ward	245	21	1.83	Yes	2	61/20	No	Aplastic anaemia	No		
F	45	LITU	32	5	0.06	No	0	52/22	Yes	POD	Yes	ALF	14
M	76	Ward	48	29	0.5	No	5	82/25	No	Rectal CA	No		
M	66	Ward	28	66	0.66	Yes	0	80/24	No	Acute pancreatitis	No		
F‡	50	Ward	44	7	0.11	Yes	10	51/20	Yes	LVH/megarectum	Yes	OCM	13
F	22	Ward	119	12	0.5	Yes	3	75/29	No	Aplastic anaemia	No		
M	41	LITU	98	28	1	No	3	–	No	Cholangio CA	No		
M	29	LITU	73	16	0.42	Yes	2	80/22	No	LT for ALF	No		
M	65	LITU	25	5	0.04	Yes	2	–	No	Cholangio CA	Yes	MOF + sepsis	14
M	68	LITU	25	5	0.04	Yes	UNK	66/21	No	LT/SB fistula	No		
F	83	Ward	72	9	0.23	Yes	1	64/22	No	CA colon	No		
M	26	LITU	30	2	0.02	Unsure	7	102/–	No	SB ischaemia	No		
F	26	Ward	25	17	0.15	No	4	52/21	No	Pancreatitis	No		
M	61	LITU	44	45	0.7	No	0	81/22	No	Colonic CA	No		
M	73	LITU	31	38	0.42	No	UNK	90/33	Yes	Cholangio CA	Yes	Metastasis	28
F	67	Ward	25	20	0.18	No	2	66/25	Yes	Cholangio CA	No		
F	78	LITU	27	30	0.29	No	2	55/23	No	Rectal CA	No		
M	47	Ward	35	60	0.8	Unsure	UNK	57/19	No	LT-PSC	No		
M	24	ITU	25	24	0.21	No	2	71/27	No	CF	Yes	MOF + sepsis	10
F	65	ITU	25	30	0.27	No	7	–	No	PPU	No		
M	32	Ward	168	50	3	No	3	78/25	No	AML	No		
F	45	LITU	25	24	0.21	Unsure	UNK	85/30	No	Adeno CA	Yes	MOF + sepsis	10
M	65	Ward	44	10	0.16	Unsure	2	–	No	NET	No		
F	58	ITU	49	37	0.64	No	2	64/24	Yes	Ischaemic colitis	No		
M	44	ITU	50	27	0.5	No	2	76/23	No	Ischaemic SBO	No		
F	48	LITU	25	21	0.19	Unsure	5	78/–	No	ALD	No		
F‡	78	Ward	25	29	0.26	Yes	7	45/17	No	Colonic CA	Yes	SBO + AF	14
F‡	50	Ward	49	23	0.4	Yes	10	50/18	No	Choledochal cyst	No		
M‡	55	Ward	56	8	0.16	Yes	7	54/19	No	AML	No		

Maln, malnourished; NBM, nil by mouth; PO₄, phosphate drop of $>30\%$; M, male; AIH, autoimmune hepatitis; F, female; SBO, small-bowel obstruction; CA, cancer; LITU, liver intensive care unit; ALD, alcoholic liver disease; ALF, acute liver failure; OD, overdose; SB, small bowel; ITU, other intensive care units; POD, paracetamol overdose; LVH, left ventricular hypertrophy; OCM, obstructive cardiomyopathy; LT, liver transplant; MOF, multiorgan failure; UNK, unknown; PSC, primary sclerosing cholangitis; CF, cystic fibrosis; PPU, perforated pyloric ulcer; AML, acute myeloid leukaemia; NET, neuroendocrine tumour; AF, atrial fibrillation.

* Cause of death as documented in the death certificate.

† Time of death from starting parenteral nutrition.

‡ The patients who were identified as being at 'high risk' of refeeding problems.

for each patient are also given in this table. Receiver-operator curves plotted against a $\geq 30\%$ phosphate drop showed that the RI had the greatest area under the curve compared with IGF1 or leptin alone (0.75 *v.* 0.64 and 0.65, respectively; 95% CI 0.55, 0.94; *P* 0.006), as indicated in Fig. 1. The cut-off value for the RI, which conveys the best sensitivity and specificity for a significant decrease in phosphate, is 0.19, with a likelihood ratio of 3.4. At this cut-off value, the sensitivity is 78% (95% CI 40, 97%), the specificity is 77% (95% CI 60, 90%), and the positive and negative predictive values for the occurrence of a $\geq 30\%$ phosphate drop are 54 and 91%, respectively.

Receiver-operator curves plotted against death within 1 month of starting PN showed that IGF1 had the greatest area under the curve compared with either leptin or the RI (0.79 *v.* 0.62 and 0.75, respectively; 95% CI 0.64, 0.94; *P* < 0.0001), as indicated in Fig. 2. The cut-off value for IGF1, which conveys the best specificity and sensitivity for

the occurrence of death, is 33 $\mu\text{g/l}$, with a likelihood ratio of 2.6. At this cut-off value, the sensitivity is 88% (95% CI 47, 99.7%) and the specificity is 67% (95% CI 46, 84%).

Other correlations

There was no significant association between the values of either IGF1 or the RI and the presence of malnourishment, but leptin was significantly lower in those who were defined as being malnourished as opposed to others (mean leptin was 16.1 (SEM 4.0) *v.* 28.3 (SEM 3.1) $\mu\text{g/l}$, respectively; *P* = 0.004). There were no significant correlations between the measured analytes (IGF1 and leptin) and, therefore, the RI and patient's sex, age, location or estimated glomerular filtration rate. A low RI, on the other hand, was significant regarding ≥ 5 d NBM (*P* = 0.04). There was no significant correlation between a decrease in phosphate of $\geq 30\%$ and whether a patient was clinically malnourished, the number

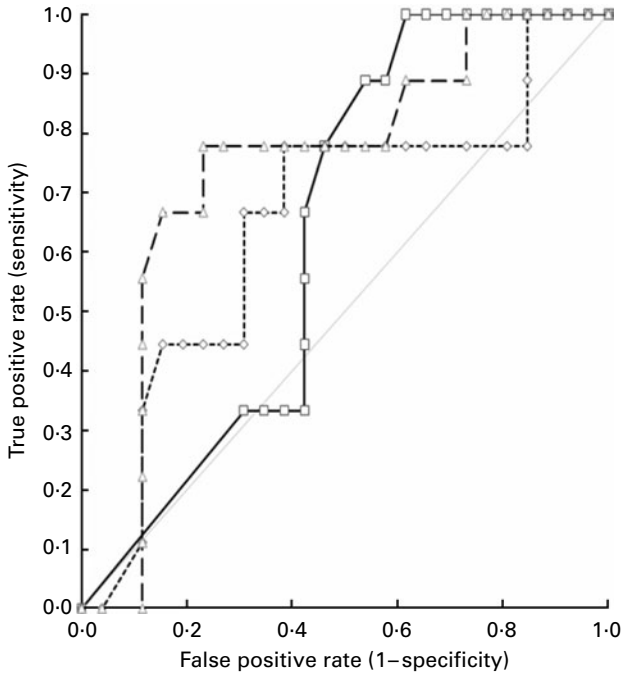


Fig. 1. Receiver-operator curves showing the area under the curve (AUC) for the thirty-five patients, when the outcome of the phosphate drop of $\geq 30\%$ on days 2–3 of starting parenteral nutrition is tested against insulin-like growth factor 1 (IGF1, $-\square-$), leptin ($-\diamond-$) and the 'refeeding index' (RI, $-\triangle-$), which employs the former two parameters. The AUC for IGF1 is 0.64, for leptin is 0.65 and for the RI is 0.75, indicating the superiority of the RI over IGF1 or leptin alone in predicting the occurrence of refeeding problems. —, No discrimination.

of days NBM before starting PN, estimated glomerular filtration rate, patient's location, age or sex. Mortality was also not found to be significantly related to the latter variables but was significantly associated with a phosphate drop of $\geq 30\%$ ($P=0.004$). IGF1 was not found to be correlated with C-reactive protein but was correlated with albumin concentrations (95% CI 0.21, 0.72; $P=0.0017$). Leptin did not correlate with either.

Discussion

We demonstrate, for the first time, that the 'RI', formulated from leptin and IGF1 concentrations, may predict a decrease in phosphate as observed in the refeeding syndrome, following the initiation of PN, and is a more powerful tool than either IGF1 or leptin alone. It therefore appears that a combination of a significantly low IGF1 and leptin presents a biochemical hallmark for predicting the refeeding syndrome.

Refeeding syndrome is commonly underdiagnosed and may present with minimal clinical indicators that could easily be masked by a multitude of pre-existing medical problems in hospitalised patients. Moreover, criteria for the identification of patients at high risk of developing this syndrome are based on a history from patients or relatives, which is often equivocal, difficult to obtain and plagued with subjectivity. Even though the nutritional state of the patient can be estimated through history and clinical examination, we have shown in the present study that the number of

days NBM and clinical signs of malnutrition, unlike the RI, are not significantly associated with a decrease in phosphate concentrations. These findings suggest that history and clinical assessment may be suboptimal predictive tools of the refeeding syndrome. This problem is compounded by the fact that electrolyte imbalances before initiating feeding are often minimal⁽⁶⁾ as we have demonstrated.

Significant reduction in phosphate concentrations after initiation of feeding appears to be the most reliable biochemical marker that the refeeding syndrome has occurred^(16–18). This reduction was observed in approximately 26% of our patients, consistent with the incidence reported in the literature that varies between 18 and 34%^(17,19). It is important to note that the acute decrease in serum phosphate observed was despite additional phosphate supplementation, in those predicted to develop the refeeding syndrome⁽⁶⁾. Moreover, only four patients had an initial phosphate concentration below the reference interval (0.80–1.40 mmol/l) before feeding; the lowest concentration in these patients was 0.65 mmol/l.

An interesting finding was the relationship between low IGF1 values and mortality; IGF1 alone being superior to the RI denotes that leptin is not as strongly linked to death. The cause of death for the majority of these patients was single or multiple organ failure. None of them had clinical signs of the refeeding syndrome; although such metabolically compromised patients may have had subclinical refeeding syndrome,

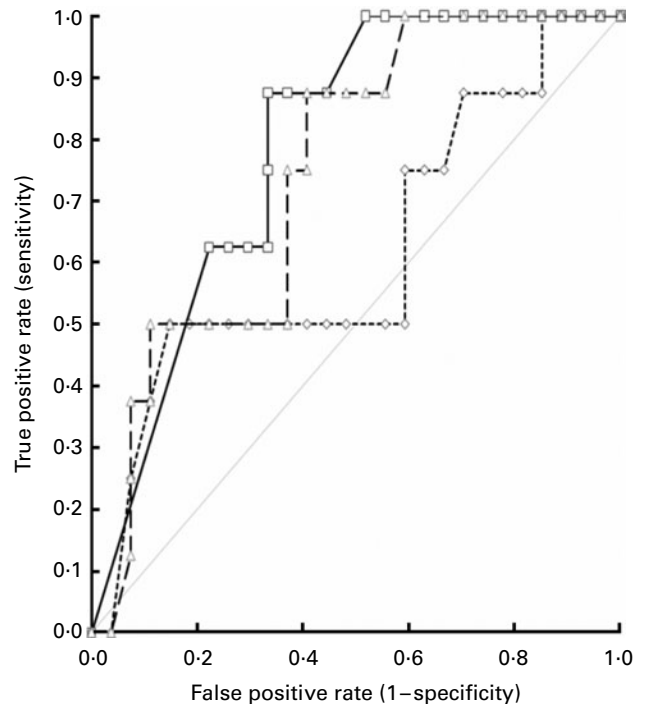


Fig. 2. Receiver-operator curves showing the area under the curve (AUC) for the thirty-five patients, when the outcome of death within 1 month (eight patients) of starting parenteral nutrition (PN) is tested against insulin-like growth factor 1 (IGF1, $-\square-$), leptin ($-\diamond-$) and the 'refeeding index' (RI, $-\triangle-$), which employs the former two parameters. The AUC for IGF1 is 0.79, for leptin is 0.62 and for the RI is 0.75, indicating that IGF1 alone is superior over leptin or the RI in predicting the occurrence of death. Note that seven out of eight patients died within the first 2 weeks of starting PN. —, No discrimination.

which could have tipped the delicate balance irreversibly. Another explanation could be that patients with organ failure, particularly liver failure, would be more prone to have lower concentrations of IGF1. The patient's location on a liver intensive care unit was not found to be significantly associated with mortality, suggesting that liver failure alone would not account for the present findings. Moreover, the fact that a decrease in phosphate of $\geq 30\%$ was significantly linked to mortality denotes that the refeeding syndrome may have played a role in these patients before death; though this syndrome remains clinically difficult to recognise.

Previous reports have observed the association of low IGF1 concentrations with cell death⁽²⁰⁾. The IGF1 signalling pathway has been established as a key modulator of ageing and longevity⁽²¹⁾. Yet, conflicting evidence surrounds the effect of a low IGF1 on death, as lifelong IGF1 deficiency causes early ageing, but not premature death⁽²¹⁾. Specifically, in the cardiovascular system, heart failure has been associated with reduced local expression of IGF1⁽²²⁾, denoting a cardioprotective role of IGF1, with low concentrations being detrimental to cardiac function. Moreover, a link between IGF1 and morbidity in hospitalised patients has previously been suggested, and it has been found to be one of the predictors of survival in cancer patients on palliative care^(23,24). Although IGF1 has been suggested to be a marker of the acute inflammatory response, particularly following surgery⁽²⁵⁾, we did not find a correlation between IGF1 and acute-phase reactants such as C-reactive protein in our patient cohort. However, a significant correlation has been found between IGF1 and albumin concentrations, suggesting that other factors besides inflammation can influence IGF1, most notably protein–energy undernutrition⁽²³⁾.

Despite abiding by the recommended guidelines^(6,12) for electrolyte replacement during PN in those suspected to be at risk of refeeding, a decrease in phosphate of $\geq 30\%$ occurred in several of our patients, even though only four patients had a low serum phosphate before starting PN. There remains a lack of clear guidance on effective management of potential refeeding syndrome. The decrease in phosphate in patients who were not identified to be at risk of the refeeding syndrome may be explained by the lack of additional supplemented phosphate and the normal rate of intravenous administration of the prescribed PN. The use of the RI may therefore help to identify more subtle cases and hence avoid the repercussions of inappropriate feeding or insufficient electrolyte replacement.

The main limitation of the present study was the relatively small study population, and hence further work is warranted in order to create better defined cut-off values for the RI to improve its positive predictive value by using more sensitive IGF1 assays and employing a larger study population. Other limitations were the inability of our IGF1 assay to accurately quantify concentrations below $25 \mu\text{g/l}$ and the fact that age- and sex-unified values of IGF1 and leptin were employed to obtain a single factor to calculate the RI. The latter would be particularly affected by the IGF1 values since it was derived from a larger set of reference ranges influenced by both age and sex; thus, expansion of our work is needed to create

age- and sex-tailored cut-off values that could introduce greater sensitivity and specificity to the RI. However, the value of the RI in the present study was found to be surprisingly unaffected by age and sex on statistical analysis. It must also be acknowledged that there is still no 'gold standard' for defining the refeeding syndrome, and thus a significant decrease in phosphate is currently the best surrogate marker to be used as a benchmark for further studies on the RI^(16–18).

The present findings suggest that the 'RI' as a normalised product of IGF1 and leptin is a novel and specific marker that can be helpful in identifying patients before starting PN who are at risk of developing the refeeding syndrome.

Acknowledgements

We would like to thank Tracy Dew from the clinical biochemistry team at King's College Hospital, for her technical assistance in some parts of the present study. The authors declare that there is no conflict of interest. The present study received no specific grant from any funding agency in the public, commercial or not-for-profit sectors. The contribution of each author was as follows: M. O. E. supervised the design of the study, collection and analysis of the data and writing of the manuscript. J. A.-Z. and C. W. I. R. were responsible for the experimental design, editing of the manuscript and provision of significant consultation. C. M. edited the manuscript and provided significant consultation. R. S. and M. A. A. were involved in the statistical analysis of the data and editing of the manuscript.

References

1. Weinsier RL & Krumdieck CL (1980) Death resulting from overzealous total parenteral nutrition: the refeeding syndrome revisited. *Am J Clin Nutr* **34**, 393–399.
2. Solomon SM & Kirby DF (1990) The refeeding syndrome: a review. *J Parenter Enteral Nutr* **14**, 90–97.
3. Mehanna HM, Moledina J & Travis J (2008) Refeeding syndrome: what it is, and how to prevent and treat it. *BMJ* **336**, 1495–1498.
4. Klein CJ, Stanek GS & Wiles CE (1998) Overfeeding macronutrients to critically ill adults: metabolic complications. *J Am Diet Assoc* **98**, 795–806.
5. Oxford Centre for Evidence-based Medicine (2009) Levels of evidence. <http://www.cebm.net/index.aspx?o=1025> (accessed 17 April 2010).
6. National Institute for Health and Clinical Excellence (2006) Nutrition support in adults. Clinical guideline CG32.
7. Korbonits M, Blaine D, Elia M, *et al.* (2007) Metabolic and hormonal changes during the refeeding period of prolonged fasting. *Eur J Endocrinol* **157**, 157–166.
8. Villanueva EC & Myers MG Jr (2008) Leptin receptor signalling and the regulation of mammalian physiology. *Int J Obes* **32**, S8–S12.
9. Nindl BC & Pierce JR (2010) Insulin-like growth factor I as a biomarker of health, fitness, and training status. *Med Sci Sports Exerc* **42**, 39–49.
10. Gómez JM, Maravall FJ, Gómez N, *et al.* (2003) Interactions between serum leptin, the insulin-like growth factor-I system, and sex, age, anthropometric and body composition

- variables in a healthy population randomly selected. *Clin Endocrinol* **58**, 213–219.
11. Schofield WN (1985) Predicting basal metabolic rate, new standards and review of previous work. *Hum Nutr Clin Nutr* **39**, 5–41.
 12. Dewar H & Horvath R (2007) Refeeding syndrome. In *A Pocket Guide to Clinical Nutrition*, 3rd ed., [VE Todorovic and A Micklewright, editors]. Worksop, Nottinghamshire: British Dietetic Association.
 13. Brabant G, von zur Mühlen A, Wüster C, *et al.* (2003) Serum insulin-like growth factor I reference values for an automated chemiluminescence immunoassay system: results from a multicenter study. *Horm Res* **60**, 53–60.
 14. Unal M, Eskandari G, Muşlu N, *et al.* (2006) Serum leptin levels in patients with allergic rhinitis. *Otolaryngol Head Neck Surg* **134**, 331–333.
 15. Holzinger U, Kitzberger R, Fuhrmann V, *et al.* (2007) Correlation of calculated indices of insulin resistance (QUICKI and HOMA) with the euglycaemic hyperinsulinaemic clamp technique for evaluating insulin resistance in critically ill patients. *Eur J Anaesthesiol* **24**, 966–970.
 16. Marvin VA, Brown D, Portlock J, *et al.* (2008) Factors contributing to the development of hypophosphataemia when refeeding using parenteral nutrition. *Pharm World Sci* **30**, 329–335.
 17. Martinez MJ, Matrinez MA, Montero M, *et al.* (2006) Hypophosphatemia in postoperative patients on total parenteral nutrition: influence of nutritional support teams. *Nutr Hosp* **21**, 657–660.
 18. Adkins SM (2009) Recognizing and preventing refeeding syndrome. *Dimens Crit Care Nurs* **28**, 53–58.
 19. Marik PE & Bedigan MK (1996) Refeeding hypophosphataemia in an intensive care unit: a prospective study. *Arch Surg* **131**, 1043–1047.
 20. Sanchez C, Oskowitz A & Pochampally RR (2009) Epigenetic reprogramming of IGF1 and leptin genes by serum deprivation in multipotential mesenchymal stromal cells. *Stem Cells* **27**, 375–382.
 21. Laron Z (2008) The GH-IGF1 axis and longevity. The paradigm of IGF1 deficiency. *Hormones (Athens)* **7**, 24–27.
 22. Hambrecht R, Schulze PC, Gielen S, *et al.* (2005) Effects of exercise training on insulin-like growth factor-I expression in the skeletal muscle of non-cachectic patients with chronic heart failure. *Eur J Cardiovasc Prev Rehabil* **12**, 401–406.
 23. Sullivan DH & Carter WJ (1994) Insulin-like growth factor I as an indicator of protein-energy undernutrition among metabolically stable hospitalized elderly. *J Am Coll Nutr* **13**, 184–191.
 24. Fouladiun M, Körner U, Bosaeus I, *et al.* (2005) Body composition and time course changes in regional distribution of fat and lean tissue in unselected cancer patients on palliative care – correlations with food intake, metabolism, exercise capacity, and hormones. *Cancer* **103**, 2189–2198.
 25. Holdaway IM, Lethaby AE, Mason BH, *et al.* (2001) Effect of breast surgery on serum levels of insulin-like growth factors (IGF-I, IGF-II, and IGF binding protein-3) in women with benign and malignant breast lesions. *Ann Surg Oncol* **8**, 25–31.