

Review: Endocrine pathways to regulate calcium homeostasis around parturition and the prevention of hypocalcemia in periparturient dairy cows

L. E. Hernández-Castellano^{1,2†} , L. L. Hernandez³ and R. M. Bruckmaier¹

¹Veterinary Physiology, Vetsuisse Faculty, University of Bern, 3001 Bern, Switzerland; ²Department of Animal Science, AU-Foulum, Aarhus University, 8830 Tjele, Denmark; ³Department of Dairy Science, University of Wisconsin, Madison, WI 53706, USA

(Received 11 January 2019; Accepted 18 June 2019; First published online 24 July 2019)

Calcium homeostasis is crucial for the normal function of the organism. Parathyroid hormone, calcitriol and calcitonin play critical roles in the homeostatic regulation of calcium. Serotonin and prolactin have also been shown to be involved in the regulation of calcium homeostasis. In modern dairy cows, the endocrine pathways controlling calcium homeostasis during non-lactating and non-pregnant physiological states are unable to fully support the increased demand of calcium required for milk synthesis at the onset of lactation. This review describes different endocrine systems associated with the regulation of calcium homeostasis in mammalian species around parturition with special focus on dairy cows. Additionally, classic and novel strategies to reduce the incidence of hypocalcemia in parturient dairy cows are discussed.

Keywords: transition period, metabolism, metabolic diseases, mammary gland, mineral balance

Implications

Calcium is essential in multiple physiological processes. Under normal physiological conditions, calcium is tightly regulated by parathyroid hormone, calcitriol and calcitonin. Dairy cows have been selected for high milk production. At the onset of lactation, the high demand of calcium for milk production often causes decreased calcium concentrations in blood, which leads to subclinical or clinical hypocalcemia. Besides the impact on performance and health, hypocalcemia increases the risk of suffering from further metabolic diseases. This review describes factors associated to calcium regulation around parturition as well as established and novel strategies to reduce hypocalcemia incidence in dairy cows.

Introduction

Calcium is the most abundant mineral in all vertebrate animals. Most of the body's calcium (99%) is stored in the bones and teeth, with only 1% stored in blood and other tissues. Calcium is a critical regulator of multiple physiological processes, including blood coagulation, nerve conduction, membrane permeability, muscle contraction, enzyme activity and hormone release. Based on the multiple important functions

of calcium, maintenance of calcium concentrations in the normal physiological range is tightly regulated in all organisms (Brown, 1991). The classic calciotropic hormones – parathyroid hormone (PTH), calcitriol and calcitonin (CT) – regulate calcium homeostasis under normal physiological conditions in mammals by coordinating the availability of calcium in the blood at the level of the bones, intestine and kidneys (Brommage and DeLuca, 1985; Hoenderop *et al.*, 2005b).

Lactation is part of the reproductive strategy employed by mammals to improve the survival of their offspring. Calcium is the most abundant mineral present in milk. Because calcium is involved in many functional systems of the organism, and despite the enormous loss of calcium into milk, calcium homeostasis is crucial for the survival of the lactating animal. Therefore, calcium homeostasis is differentially regulated during pregnancy and lactation compared to non-pregnant and non-lactating physiological states (Bauman and Currie, 1980; Salari and Abdollahi, 2014). The calcium uptake by the mammary gland for milk production decreases circulating calcium concentrations, which triggers the homeostatic mechanisms to restore calcium concentrations in blood. The homeostatic regulation of calcium in lactating rodents and humans is known to be largely regulated by parathyroid hormone-related protein (PTHrP) (Kovacs, 2011), rather than PTH, which is crucial in non-lactating physiological states.

† E-mail: lhc@anis.au.dk

Table 1. Calcium dynamics during lactation in human, rat and dairy cow species¹

	Human	Rat	Dairy cow (Holstein)
BW at partum (kg)	75.0	0.35	680
Calcium in blood in normal conditions (mmol/l)	2.37	3.01	2.30
Calcium in blood at partum (mmol/l)	2.14	2.78	1.72
Colostrum yield (kg/day)	0.07	5×10^{-3}	8.23
Colostrum yield per kg of BW (kg)	9×10^{-4}	0.01	0.01
Calcium in colostrum (mmol/l)	5.35	16.3	54.2
Milk yield (kg/day)	0.77	0.01	35.4
Milk yield per kg of BW (kg)	0.01	0.02	0.05
Calcium in milk (mmol/l)	6.25	24.5	28.7

¹ Data obtained from Keen *et al.*, 1981; Saint *et al.*, 1984; Neville *et al.*, 1988; Goldstein, 1990; Thorsdottir and Birgisdottir, 1998; LaBorde *et al.*, 1999; VandeHaar and St-Pierre, 2006; Tsioulpas *et al.*, 2007; Capper and Cady, 2012; Hernández-Castellano *et al.*, 2017b).

Unlike other mammalian species, dairy cows have been genetically selected for high milk production beyond neonatal requirements (Table 1). Overall, the produced amount of milk does usually not exceed the nutritional requirements of the offspring. As shown in Table 1, calcium concentrations are higher in both colostrum and milk from dairy cows than those for humans. Therefore, the enormous demand of the mammary gland for calcium around parturition makes dairy cows unique to study different physiological processes involved in calcium homeostasis at the onset of lactation.

This review describes diverse endocrine systems and other factors associated with the regulation of calcium homeostasis in mammalian species at the onset of lactation, focusing on dairy cows. Additionally, we review traditional strategies such as those based on the manipulation of the mineral balance, the dietary availability of calcium and the administration of either calcitriol or its precursors to reduce the incidence of hypocalcemia (HC) in parturient dairy cows. We also review novel players in calcium homeostasis such as serotonin and prolactin, which may be implemented in future strategies to prevent HC in dairy cows.

Endocrine regulation of calcium during pregnancy and lactation

Calcium concentrations in blood are tightly maintained by the regulation of intestinal calcium absorption, renal calcium reabsorption and bone calcium resorption. Calcium fluxes in these three organs are primarily regulated by the hormones PTH, calcitriol and CT. Decreased calcium concentrations activate calcium-sensing receptors located on the parathyroid chief cells, which cause PTH release into the bloodstream

(Kumar and Thompson, 2011). Under normal physiological conditions, PTH signals via its receptor on the osteoblast membrane, which increases osteoclast proliferation, increasing bone resorption activity. In addition to osteoclastic bone resorption, osteocytes seem to contribute to calcium transferred from bones through a mechanism known as osteocytic osteolysis (Wysolmerski, 2012). Osteocytes are able to remodel bone matrix contained in the lacunae, releasing calcium into the bloodstream through the osteocyte canalicular network. Increased PTH levels stimulate osteocytes to resorb perilacunar mineral (mainly calcium) and collagen, which increases the size of the lacunar area (Belanger and Migicovsky, 1963; Tazawa *et al.*, 2004; van Bezooijen *et al.*, 2004). Wysolmerski (2012) described increased lacunar area in lactating rats compared to virgin rats. However, these differences were not evidenced in rats with osteocyte-specific disruption of the PTH receptor 1 gene. These results suggest that osteocytic osteolysis could play an important role in calcium homeostasis during lactation; however, further studies will be necessary to know how quantitatively important is osteocytic bone remodeling during lactation. During bone resorption, collagen is degraded by osteoclasts and osteocytes, and factors such as pyridinoline (PYD), deoxypyridinoline (DPD) and collagen type I C telopeptide (CTX) are released into the bloodstream (Urena *et al.*, 1995). Therefore, PYD, DPD and CTX are often used as markers for calcium release from the bone to the bloodstream not only in humans (Urena *et al.*, 1995; Eastell *et al.*, 1997) but also in dairy ruminants (Liesegang *et al.*, 1998; Liesegang, 2000; Holtenius and Ekelund, 2005; Wilkens *et al.*, 2014; Hernández-Castellano *et al.*, 2017a).

In addition to increased bone resorption, PTH also stimulates the secretion of the renal 1- α -hydroxylase, the enzyme that catalyzes the hydroxylation of calcidiol to calcitriol. During pregnancy and lactation, other endocrine factors such as prolactin, estradiol and placental lactogen are also able to stimulate the activity of 1- α -hydroxylase (Kovacs and Kronenberg, 1997), contributing to increased calcitriol concentrations. Although similar studies do not exist in ruminants, several authors have described increased calcitriol concentrations in cows (Özçelik *et al.*, 2017), goats (Starke *et al.*, 2016) and sheep (Klinger *et al.*, 2016) during late pregnancy and early lactation. Therefore, it seems that the regulation of calcitriol is expanded during this period due to the increased demand of calcium. Calcitriol, also known as the active form of vitamin D (1,25-(OH)₂D₃), increases calcium concentrations in blood by increasing both intestinal calcium absorption and renal calcium reabsorption (Horst *et al.*, 1994). In the intestine, calcitriol regulates calcium absorption mainly through the active transcellular pathway. Intestinal transcellular transport consists of calcium entry through specific calcium channels such as transient receptor potential vanilloid type 5 (TRPV5) and transient receptor potential vanilloid type 6 (TRPV6) present in membranes of the brush border, followed by intracellular transport mediated by the calcium-binding protein, calbindin-D_{9k}. Calcium is then actively transported to the bloodstream by the plasma membrane calcium ATPase (PMCA1b) (Lieben *et al.*, 2011).

Calcitriol also increases the efficiency of passive paracellular transport of calcium by increasing the expression of claudin-2 and claudin-12 in the gut (Lieben *et al.*, 2011); however, this mechanism is still not fully understood. In the kidney, calcitriol regulates calcium reabsorption which is mediated by active transcellular transport in the distal nephron (Hoenderop *et al.*, 2005a). Calcium enters the kidney through TRPV5, binding to calbindin-D_{9k} and calbindin-D_{28k}. Then, calcium enters the bloodstream through the Na⁺/Ca²⁺ exchanger (NCX1) and PMCA1b (Hoenderop *et al.*, 2005a). In the intestine and the kidney, calcitriol increases the expression of calcium transporters (i.e., calbindin-D_{9k}, calbindin-D_{28k}, PMCA1b, TRPV5 and TRPV6) (Fleet and Wood, 1999; Li *et al.*, 2001; Lieben *et al.*, 2011). In humans, Ardawi *et al.* (1997) showed how calcitriol concentrations are increased two- to threefold during pregnancy (333 pmol/l) and lactation (267 pmol/l) compared to normal physiological conditions (93 pmol/l). However, in rodents, the role of calcitriol in calcium homeostasis during lactation seems to be secondary, as mice and rats deficient in the calcitriol receptor are able to lactate and provide adequate milk for their pups (Fudge and Kovacs, 2010; Kovacs, 2012).

When calcium concentrations increase above normal physiological levels, increased cytoplasmic calcium concentrations in the parafollicular cells of the thyroid (C cells) cause CT release from C cells into the bloodstream (Garrett *et al.*, 1995). Calcitonin primarily acts on the osteoclasts, reducing calcium resorption from bones. Specifically, CT promotes dephosphorylation of Pyk2 and phosphorylation of focal adhesion kinase, which reduce osteoclast adhesion. Additionally, CT also disrupts the cytoskeletal organization of the osteoclasts, reducing osteoclast mobility and causing cell retraction (Zhang *et al.*, 2002; Takahashi *et al.*, 2007; Del Fattore *et al.*, 2008). As observed in a study in humans, CT concentrations increase during pregnancy (20 pmol/l) and early lactation (25 pmol/l) compared to non-pregnant physiological states (9.2 pmol/l) (Ardawi *et al.*, 1997). These data suggest that CT may have an important role protecting the maternal skeleton from excessive calcium resorption. Similarly, mice deficient for the CT gene lose twice as much bone mineral content (−53.6%) compared to wild-type mice (−23.6%) during the first 21 days of lactation (Woodrow *et al.*, 2006), suggesting again the protective role of CT on bones during the periods of high calcium demand, such as pregnancy and lactation.

Prolactin and its role in calcium homeostasis

As described above, calcium homeostasis is regulated by other endocrine factors in addition to PTH, calcitriol and CT. Prolactin is secreted by lactotrophs in the anterior pituitary (Lamberts and Macleod, 1990). This hormone has a predominant role in milk secretion during lactogenesis, and it has also been demonstrated to regulate calcium homeostasis. At the onset of lactation, prolactin concentrations are increased about 5- to 20-fold in humans, rats and cows compared to non-lactating concentrations (Pepperell *et al.*, 1977; Johke,

1979; Jahn *et al.*, 1993; Mazor *et al.*, 1996; Tong *et al.*, 2018). In addition to the role of prolactin in lactogenesis, it has also been demonstrated to be critical for intestinal calcium transport in mice, rats and humans (Pahuja and DeLuca, 1981; Ajibade *et al.*, 2010; Christakos *et al.*, 2011), regulating diverse calcium transport proteins such as TRPV6 and calbindin-D_{9k} (Ajibade *et al.*, 2010). As demonstrated by Wongdee *et al.* (2016), prolactin increases intestinal calcium absorption in rats by increasing the absorptive surface area and enhancing the expression of other calcium transporters (PMCA1b and NCX1) in addition to TRPV6. Less research is available regarding the role of prolactin on renal calcium reabsorption. Lotinun *et al.* (1998) observed that pregnant rats treated with prolactin reduced urinary calcium excretion (5 μmol/kg of BW per day) compared to control pregnant rats (10 μmol/kg of BW per day). Additionally, Robinson *et al.* (1982) suggested that prolactin indirectly reduces renal calcium excretion by increasing the activity of 1-α-hydroxylase, which increases calcitriol concentrations.

In addition to the effects of prolactin on the intestine and kidneys, prolactin increases calcium resorption from bones and reduces renal calcium excretion in humans and rats during pregnancy and lactation (Mainoya, 1975; Ritchie *et al.*, 1998; Lotinun *et al.*, 2003; Charoenphandhu and Krishnamra, 2007). Prolactin's effects on bone calcium metabolism are predominantly on the primary trabecular sites (Puntheeranurak *et al.*, 2006; Charoenphandhu and Krishnamra, 2007). Prolactin acts by decreasing mRNA expression of runt-related transcription factor (Runx) 2 (Charoenphandhu *et al.*, 2008), which is essential for osteoblast maturation and differentiation (Komori *et al.*, 1997; Xiao *et al.*, 2005). When osteoblast maturation and formation are suppressed, calcium absorption from blood into bones is reduced in favor of increased bone resorption from bones.

Serotonin and its role in calcium homeostasis

Serotonin (5-Hydroxytryptamine; 5-HT), a biogenic amine and tryptophan derivative, is synthesized in the CNS and regulates physiological processes related to mood or appetite (Jenkins *et al.*, 2016). However, 98% of 5-HT is synthesized in organs such as the intestine, lung, pancreas, prostate, salivary gland, thyroid, liver and the mammary gland (Lauder, 2004; Hernandez *et al.*, 2009; Pai and Marshall, 2011). Serotonin has been demonstrated to contribute to calcium homeostasis in humans (Modder *et al.*, 2010), mice (Laporta *et al.*, 2014) and dairy cows (Laporta *et al.*, 2013). During lactation, prolactin stimulates the expression of genes essential for 5-HT biosynthesis (tryptophan hydroxylase, **TPH**; and aromatic amine decarboxylase, **AADC**) by mammary epithelial cells in response to filling of the mammary gland (Matsuda *et al.*, 2004). Consequently, mammary epithelial cells increase the secretion of 5-HT into the bloodstream, contributing to an elevation in circulating 5-HT concentrations

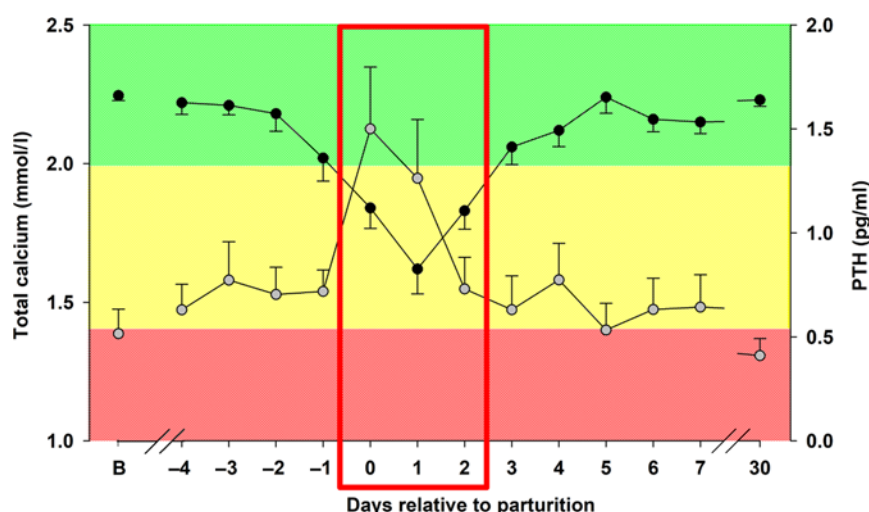


Figure 1 (Color online) Total calcium (●) and parathyroid hormone (PTH; ○) concentrations in dairy cows around parturition. Total calcium was measured in blood serum. PTH was measured in blood plasma. The red box shows time around parturition, when circulating calcium concentrations abruptly drop, causing an increase in PTH concentrations. B, baseline obtained 2 weeks before parturition. Cows with calcium concentrations within the green area are within the normal circulating calcium levels. Cows with calcium concentrations within the yellow area are under subclinical hypocalcemia. Cows with calcium concentrations within the red area are under clinical hypocalcemia. Adapted from Hernández-Castellano *et al.* (Hernández-Castellano *et al.*, 2017a and 2017b).

(Matsuda *et al.*, 2004; Hernandez *et al.*, 2012; Laporta *et al.*, 2014). Additionally, 5-HT has been demonstrated to stimulate the production of mammary-derived PTHrP (Hernandez *et al.*, 2012). As reviewed by Cowin and Wysolmerski (2010), PTHrP is necessary for full differentiation of the mammary mesenchyme and regulates the expression of at least 40 molecules in the mammary gland. During lactation, the mammary gland secretes PTHrP into milk at concentrations exceeding 10 000 times the amount of PTHrP present in the blood (Riond *et al.*, 1995; Kovacs, 2011). Despite the function of PTHrP in milk remains unknown, the concentration of this peptide in milk is correlated to circulating calcium concentrations in cows (Kocabagli *et al.*, 1995). At the onset of lactation, PTHrP concentrations in the circulation increase to detectable levels in humans, rats and cows (Thiede and Rodan, 1988; Onda *et al.*, 2006; Kovacs, 2011), for the purpose of coordinating calcium metabolism during lactation (Rakopoulos *et al.*, 1992; Onda *et al.*, 2006). It has been demonstrated that PTHrP utilizes the PTH receptor, suggesting that it has the same biological activity as PTH, although it does not regulate calcitriol (Philbrick *et al.*, 1996; Strewler, 2000). Therefore, the interaction of 5-HT with PTHrP implicates the possibility of 5-HT in regulating calcium homeostasis during lactation (Hernandez, 2017). Schilling *et al.* (1993) demonstrated that PTHrP does not stimulate renal 1-hydroxylase activity in patients with humoral hypercalcemia of malignancy. This is in contrast to the action of PTH during primary hyperparathyroidism in which PTH stimulates renal 1-hydroxylase activity and consequently increases calcitriol concentrations. Although PTH increases intestinal calcium absorption and decreases renal calcium excretion through increased calcitriol concentrations, the effect of PTHrP on these mechanisms remains unclear. Therefore, it is possible that PTHrP may only act on the bone to regulate calcium metabolism.

Calcium regulation in dairy cows and hypocalcemia during the periparturient period

The periparturient period involves considerable metabolic adaptations in mammals. In high-yielding dairy cows, the sudden high demand of nutrients for milk production at the onset of lactation and the associated metabolic load required for the production of this copious amount of milk increasingly exceeds the adaptive capacity of the animal, leading to increased incidence of several metabolic diseases (Paudyal *et al.*, 2018), HC being one of the most relevant (Figure 1). There are a variety of factors that can modulate and affect the metabolic adaptation around parturition.

Lactation, breed and milk yield

Lactation number is a critical factor in the etiology of HC. Reinhardt *et al.* (2011) described that plasma calcium concentrations decreased much less during the first 48 h postpartum in primiparous cows compared to multiparous cows (2.15 v. 1.85 mmol/l, respectively). These data suggest that the mechanisms for calcium homeostasis become less effective as the cow undergoes subsequent lactations and may contribute to greater or prolonged HC in older animals. In agreement with this observation, Fleischer *et al.* (2001) and Caixeta *et al.* (2017) described a positive correlation between HC and lactation number. In primiparous cows, remodeling processes in the bones are very active, and therefore calcium resorption from the bones is rapid and efficient compared to multiparous cows (Iwama *et al.*, 2004). Aging also results in a decline in the active transport of calcium in the intestine and impaired production of calcitriol (Woodrow *et al.*, 2006), affecting the capacity of the animal to rapidly modulate imbalances in blood calcium concentrations.

In addition to lactation number, breed also contributes to HC susceptibility in dairy cows. Henderson (1938) reported significant differences in HC incidence between cow breeds (15.3% in Brown Swiss, 13.3% in Shorthorn, 8.6% in Guernsey and 6.0% in Ayrshire). Lean *et al.* (2006) demonstrated that HC incidence is 2.25 times higher in Jersey cows than in Holstein cows. Similar results were reported by Chiwome *et al.* (2017) on HC incidence in Jersey and Holsteins cows (14.78% and 4.82%, respectively). Despite several studies having described HC incidence among breeds, the mechanism involved in these differences remains unclear. Goff *et al.* (1995) determined that Jersey cows have fewer intestinal calcitriol receptors than Holstein cows (95 v. 142 fmol/mg protein, respectively). The relative amount of intestinal calcitriol receptors in Jersey cows compared to Holsteins cows may affect the amount of calcium that can be absorbed in the intestine, which could be a potential explanation for the differences in HC susceptibility between breeds. It has also been suggested that differences in HC incidence between Jerseys and Holsteins may be related to the amount of milk produced by kilogram of BW (Blake *et al.*, 1986; Chiwome *et al.*, 2017), which is higher in Jerseys (average BW = 454 kg; average milk production = 31.7 kg/day) compared to Holsteins (average BW = 680 kg; average milk production = 35 kg/day) (Capper and Cady, 2012). However, not only the relation between milk yield and BW, but milk yield alone appears to be positively correlated with HC incidence in dairy cows. Jawor *et al.* (2012) demonstrated that cows suffering HC during the first 48 h postpartum are those that produced approximately 6.4 kg of milk more per day compared to cows without HC. Similarly, Gild *et al.* (2015) described that cows with subclinical HC produced 3.17, 2.71 and 1.90 kg more milk compared to normocalcemic cows on the first, second and third days of lactation, respectively. It is assumed that cows with greater capacity for milk production may also be prone to greater net loss of calcium into the milk shortly after parturition (Jawor *et al.*, 2012), exceeding the already saturated adaptive capacity of the animal.

Mineral balance

Adequate mineral balance is crucial for the normal function of the metabolic and physiological processes in animal systems. Deficiencies or imbalances in the mineral content can lead to abnormal nerve function, muscle contraction, immune response, the blood coagulation cascade and other physiological processes. Minerals present in organisms in large amounts are calcium, phosphorus, magnesium, sodium, potassium, chloride and sulfur. In the specific case of calcium, several studies have detected a correlation between low circulating magnesium concentrations and low circulating calcium concentrations in humans (Classen *et al.*, 1986; Paunier, 1992), mice (Zimmermann *et al.*, 2000) and dairy cows (Kronqvist *et al.*, 2011 and 2012). Low magnesium concentrations have been shown to reduce the secretion of PTH (Suh *et al.*, 1973), as well as decrease tissue responsiveness to PTH (Reddy *et al.*, 1973; Rude,

1998). Phosphorous status also affect cow's ability to regulate calcium concentrations in blood around parturition. Indeed, Cohrs *et al.* (2018) showed how reduced dietary phosphorus intake in late pregnancy increased the calcium resorption from bones, which in turn increase circulating calcium concentrations around parturition.

Based on these facts, it can be assumed that blood mineral status (i.e., magnesium and phosphorous) directly affects the efficiency of the mechanisms involved in calcium homeostasis during the periparturient period in cows, which may result in increased susceptibility to HC.

Hypocalcemia: Traditional treatments and new alternatives

At the onset of lactation, the demand for calcium by the mammary gland often causes decreased circulating calcium concentrations in dairy cows between 12 and 24 h postpartum (Goff, 2008) (Figure 1). As described by Reinhardt *et al.* (2011), 50% of dairy cows are affected by subclinical HC (blood calcium concentrations within 1.4 to 2.0 mmol/l) postpartum. Furthermore, it was determined that 5% to 7% of dairy cows suffer clinical HC (calcium concentration <1.4 mmol/l) (Roche and Berry, 2006). In most of the cases, cows with clinical HC are not able to restore normal calcium concentrations postpartum. If the cow is not treated rapidly, clinical HC can lead to the death of the animal (Oetzel, 1988). Therefore, cows with clinical HC are often treated with intravenous infusions of calcium gluconate (23%, 500 ml = 10.8 g of calcium) (Oetzel, 1988). Alternatively, the administration of a ruminal calcium bolus (approximately 43 g of calcium) can be used to restore blood calcium concentrations; however, calcium absorption is slower compared to intravenous administration of calcium gluconate. When cows undergo clinical and subclinical HC, they are more susceptible to other metabolic and infectious diseases (Compton *et al.*, 2007). Therefore, most strategies for HC prevention focus on the compensation of circulating calcium deficiencies during the first 24 to 48 h postpartum. Prevention of HC is crucial in order to increase the health and welfare in dairy cows as well as the economic benefit for dairy farmers.

Traditional management practices for the prevention of hypocalcemia

The use of diets formulated with a negative dietary cation-anion difference (DCAD) prior to parturition has long been established to increase calcium concentrations around parturition (Ender *et al.*, 1962; Dishington, 1975; Block, 1988). Shohl and Sato (1923) were the first to hypothesize that mineral interrelationships were related to acid-base status. Then, Shohl (1939) proposed that the body excretes dietary cations and anions excess in order to maintain normal acid-base balance. Based on these findings, Dishington (1975) established that the net acid intake in dairy cows, based on dietary difference between cations and anions,

was mostly influenced by sodium, potassium, chloride and sulfur. Therefore, DCAD diets for dairy cows are calculated based on the following formula: $\text{DCAD} = (\text{sodium} + \text{potassium}) - (\text{chloride} + \text{sulfur}) \text{ mEq/kg of DM}$ (Charbonneau *et al.*, 2006). Negative DCAD diets aim to reduce the charge ion difference in blood, which decreases blood and urine pH, resulting in strong ion metabolic acidosis. This metabolic acidosis is compensated in part by the bone accepting hydrogen ions (H^+) in exchange for calcium (Lemann *et al.*, 2003). Thus, the metabolic acidosis created by feeding negative DCAD diets inhibits renal calcium reabsorption prepartum, increasing urinary calcium excretion. This fact triggers the homeostatic mechanisms that increase calcium influx (Gaynor *et al.*, 1989; Phillipppo *et al.*, 1994; DeGaris and Lean, 2008) and target tissue responsiveness to parathyroid hormone (Horst *et al.*, 1997). In addition, feeding negative DCAD diets (i.e., -59 mEq/kg DM) results in increased calcitriol concentrations in blood compared to cows being fed positive DCAD diets ($68 \text{ v. } 40 \text{ pmol/l}$ on day -3 relative to parturition, respectively) (Phillippo *et al.*, 1994). However, decreased palatability of anionic salts is thought to be a limitation of this strategy, which can dramatically affect the already reduced feed intake in dairy cows around parturition (Moore *et al.*, 2000). More recently, the combination of diets with reduced basal cations in combination with more palatable anion sources have been used to increase circulating calcium concentrations around parturition without negative effects on feed intake (DeGroot *et al.*, 2010; Weich *et al.*, 2013; Weiss *et al.*, 2015). However, the relation between decreased feed intake and DCAD composition is still controversial. According to a recent publication by Zimpel *et al.* (2018), reduced feed intake in cows fed negative DCAD diets prepartum (-114 and -113 mEq/kg DM) seems to be a direct consequence of the metabolic acidosis status of the animal and not a consequence of the addition of acidogenic products or salts containing chloride in the DCAD diet. The number of days that cows need to be fed with DCAD diets is another factor that may influence the capacity of these diets to regulate calcium homeostasis at parturition. Lopera *et al.* (2018) fed two different negative DCAD diets (-70 and -180 mEq/kg DM) to dairy cows for either 42 or 21 days prepartum. They showed that cows fed for 21 days prepartum showed a similar metabolic acidosis at calving as those fed for 42 days postpartum. Therefore, it seems that feeding cows with a negative DCAD for 21 days prepartum is enough to cause a metabolic acidosis at calving and prevent or reduce HC incidence. Metabolic acidosis level prepartum can be easily monitored by measuring either urinary calcium concentrations or urinary pH (Martin-Tereso and Martens, 2014). As described by Leno *et al.* (2017) cows fed a negative DCAD diet prepartum (-74 mEq/kg DM) increased urinary calcium excretion ($8.2 \text{ v. } 0.7 \text{ g/day}$, in negative DCAD diet and control cows, respectively) and maintained urine pH between 5.5 and 6.0 compared to the control cows (urine pH 7.89). Additionally, circulating calcium concentrations ($2.27 \text{ v. } 2.16 \text{ mmol/l}$), feed intake ($3.07\% \text{ v. } 2.88\%$ of BW) and milk yield ($43.9 \text{ v. } 40.8 \text{ kg/day}$)

increased postpartum in cows fed a negative DCAD diet compared to control cows. Similar results have been recently shown by Diehl *et al.* (2018) using a negative DCAD diet (-222 mEq/kg DM) during the last 21 days prepartum.

As an alternative to negative DCAD, feeding low calcium diets prepartum have also been used to induce homeostatic adaptation of calcium metabolism in dairy cows. Calcium requirements for dry cows are minimal (22 g/day for a 500 kg cow; NRC, 2001), being necessary only for maintenance and fetal skeletal development. Feeding $<20 \text{ g}$ of calcium per day during the dry period decreases calcium concentrations in blood, which activates calcium homeostatic mechanisms involving calcium mobilization from bones, intestinal calcium absorption and renal calcium resorption (Kichura *et al.*, 1982). This prepartum feeding strategy results in increased calcium mobilization from bones into the bloodstream and contributes to compensate calcium requested for milk production (Thilising-Hansen *et al.*, 2002a). Additionally, cows fed low calcium diets before parturition increase intestinal efficiency for calcium absorption at parturition. In order to activate calcium homeostatic mechanisms before parturition, low calcium diets should provide $<20 \text{ g}$ of calcium per day. Therefore, high calcium forages such as alfalfa (12 g of calcium/kg DM) need to be exchanged by low calcium forages such as corn silage (1.7 g of calcium/kg DM) and barley silage (3.4 g of calcium/kg DM) in dry cow diets in order to reduce calcium intake. However, this is difficult to achieve in some places of Europe and North America, due to the predominant forages available for dairy cows. Therefore, an alternative strategy was explored in instances where calcium content of forages is relatively high, which is the inclusion of rumen-protected rice bran in the diet (Martin-Tereso *et al.*, 2016). The low calcium content of this product (5.5 g/kg DM) and the calcium-binding capacity of the phytic acid present in rice bran (61.5 g of phytic acid/kg DM) reduce the dietary calcium available for intestinal absorption, activating the homeostatic calcium mechanisms. Indeed, Martin-Tereso *et al.* (2016) showed that cows receiving 140 g of rumen-protected rice bran/kg DM prepartum exhibited higher calcium concentrations compared to controls at parturition ($1.98 \text{ v. } 1.87 \text{ mmol/l}$, respectively), 6 h postpartum ($2.05 \text{ v. } 1.85 \text{ mmol/l}$, respectively) and 12 h postpartum ($2.15 \text{ v. } 1.92 \text{ mmol/l}$, respectively). As described above, these extraordinary physiological conditions for efficient calcium absorption can be used in cows after parturition by feeding them a diet without zeolite and with high calcium content. Large amounts of calcium absorbed in the intestine will contribute to the required amount of calcium for milk production and will maintain circulating calcium concentration constant during the first days after parturition. An additional strategy was developed by Thilising-Hansen *et al.* (2002b) to maintain low calcium concentrations in dairy cow diets prepartum. These authors aimed to increase the efficiency the intestinal calcium absorption by feeding dietary zeolite (sodium aluminum silicate) during the last 2 weeks prepartum. Similar to rice bran, zeolite acts by binding dietary calcium, reducing the amount of calcium available

for intestinal absorption. Therefore, the functional calcium deficiency prepartum caused by the dietary zeolite stimulates the intestine to increase calcium absorption efficiency at parturition. The use of zeolite appears to be effective in the prevention of HC as cows supplemented with zeolite prepartum had higher calcium concentrations compared to the controls around parturition (2.35 v. 2.05 mmol/l, respectively) (Thilting-Hansen *et al.*, 2002b). Similar results were shown by Khachlouf *et al.* (2019) in cows supplemented with 200 g of sodium aluminum silicate per day during the last 40 days prepartum. In this study, supplemented cows showed higher circulating calcium concentrations at calving than the control cows (~2.34 v. 2.23 mmol/l, respectively).

Another strategy for the prevention of HC consists on the administration of exogenous calcitriol or its precursors in order to increase calcitriol concentrations around parturition, resulting in increased calcium concentrations in blood. Cholecalciferol is one of the precursors of calcitriol. Cholecalciferol is converted to calcidiol, the substrate for 1- α -hydroxylase and renal synthesis of calcitriol. Consequently, the role of cholecalciferol in the prevention of HC has been thoroughly investigated. Hibbs *et al.* (1947) demonstrated that supplementation with 1 to 5 million IU of cholecalciferol per day during 2 to 4 weeks before parturition was not sufficient to prevent HC in Jersey cows. In subsequent studies, Hibbs and Pouden (1955) demonstrated that daily supplementation with 30 million IU of cholecalciferol between 8 and 3 days prevented HC in Jersey cows with previous HC problems. However, several studies have shown that the amount of dietary cholecalciferol required for increasing calcium concentrations is close to the toxic dose (Swan, 1951; Capen *et al.*, 1966; Julien *et al.*, 1977), causing marked anorexia, loss of BW, dyspnea, tachycardia, recumbency and severe cardiovascular calcifications (Thilting-Hansen *et al.*, 2002a). According to nutrient requirements for dairy cattle published by the NRC (2001), approximately 20 000 IU of cholecalciferol per day are necessary to support calcium and phosphorus homeostasis during lactation. In another study, a single subcutaneous injection of calcitriol (300 μ g) within the first 6 h postpartum successfully increased calcitriol concentrations from 58.5 to 1271 pmol/l 24 h, resulting in increased total calcium concentrations from 2.52 to 3.02 mmol/l (Vieira-Neto *et al.*, 2017). Additionally, the combination of DCAD diets together with dietary supplementation of either cholecalciferol or calcidiol prepartum in dairy cows has been explored (Martinez *et al.*, 2018). Feeding prepartum cows a diet containing a negative DCAD diet (-130 mEq/kg DM) regardless of the source of vitamin D resulted in increased calcium concentrations around parturition (2.18 mmol/l) compared to cows fed a positive DCAD (+130 mEq/kg DM) diet (1.96 mmol/l). This suggests that the source of vitamin D supplementation does not matter prepartum as long as it is in combination with a negative DCAD diet.

Novel treatments for the prevention of hypocalcemia

In the last decade, the knowledge surrounding the endocrine regulation of calcium during lactation has increased. Based on this new knowledge, factors such as 5-HT have been manipulated to develop novel and potentially more efficient strategies to prevent HC in parturient dairy cows.

In addition to the ability of the mammary gland to synthesize 5-HT, the mammary gland is able to respond to 5-HT as the bovine mammary epithelium expresses multiple 5-HT receptors such as 5-HT-1B, 5-HT-2A, 5-HT-2B, 5-HT-4 and 5-HT-7 (Hernandez *et al.*, 2009). At the level of the mammary gland, 5-HT is thought to increase PTHrP production (Hernandez *et al.*, 2012); however, it is not known how 5-HT induces the production of PTHrP in the mammary gland. Based on these findings in rodents, research has been performed to evaluate the role of 5-HT on calcium homeostasis in dairy cows (Weaver *et al.*, 2016; Hernández-Castellano *et al.*, 2017a and 2017c; Weaver *et al.*, 2017). In these studies, 5-hydroxy-l-tryptophan (5-HTP), a hydroxylated derivative of the amino acid tryptophan, has been intravenously infused to promote the synthesis of 5-HT by the peripheral tissues in dairy cows prior to parturition. Furthermore, 5-HTP has been used as an oral supplement to increase 5-HT concentrations in dairy calves (Hernández-Castellano *et al.*, 2017b). In Holstein cows, increased 5-HT concentrations in the prepartum period resulted in a less dramatic decrease in calcium concentrations compared to control cows on day 1 (1.93 v. 1.62 mmol/l, respectively) and day 2 postpartum (2.07 v. 1.83 mmol/l, respectively) (Hernández-Castellano *et al.*, 2017c). Similar results were observed in Holstein cows in Weaver *et al.* (2016). However, these authors observed that the administration of 5-HTP prepartum resulted in decreased calcium concentrations in Jersey cows before parturition (2.60, 2.25, 2.15 and 2.10 mmol/l from day 4 to 1 prior to parturition, respectively). Jersey cows treated with 5-HTP prepartum increased circulating calcium concentrations earlier postpartum compared to controls (2.25 v. 2.00 mmol/l on day 1 postpartum, respectively). Therefore, it appears that Holstein and Jersey cows differentially respond to 5-HTP administration prepartum. It is possible that in Jersey cows, decreased circulating calcium concentrations prepartum may trigger an earlier recovery of circulating calcium concentrations at parturition. The intravenous infusion of 5-HTP prepartum also resulted in an increase of PYD concentrations in blood on day 1 postpartum (Hernández-Castellano *et al.*, 2017a). These results suggest that cows with increased 5-HT concentrations prepartum potentially mobilized calcium from bone tissue prior to control cows. Furthermore, the administration of 5-HTP did not result in a change of either PTH (Hernández-Castellano *et al.*, 2017a) or calcitriol concentrations (54 and 48 pmol/l in 5-HTP and control groups, respectively; Hernández-Castellano and Bruckmaier, unpublished data). Therefore, the current data suggest that 5-HT possibly increases calcium resorption from the bones through a different mechanism than PTH, potentially via PTHrP secretion by the mammary gland. The administration of 5-HTP prepartum also resulted in increased 5-HT concentrations

in colostrum (37 v. 24 nmol/l in 5-HTP and control cows, respectively) (Hernández-Castellano *et al.*, 2017c). As 5-HTP infusions ceased at parturition, no differences in milk 5-HT concentrations were detected between groups on day 7 postpartum (12 nmol/l). Additionally, Slater *et al.* (2018) combined the use of negative DCAD (−55 v. +14 mEq/kg DM) with the administration of 5-HTP (1 mg 5-HTP/kg BW) prior to parturition, resulting in further increases of both ionized and total calcium concentrations compared to negative DCAD or 5-HTP treatment alone.

Concluding remarks

The present work summarizes the endocrine mechanisms involved in the regulation of calcium homeostasis in mammalian species during lactation, with a special focus on dairy cows. Furthermore, it describes novel research on calcium homeostasis as well as new strategies for the prevention of HC at parturition in dairy cows. Research and knowledge present in this review may be used in the near future to reduce HC incidence. Reduced HC incidence will not only increase farmer's benefit but also improve animal welfare and reduce the risk of other metabolic diseases during the transition period.

 L. E. Hernández-Castellano 0000-0003-2729-0434

Declaration of interest

The authors declare that there are no conflicts of interest.

Ethics statement

None.

Software and data repository resources

None of the data were deposited in an official repository.

Supplementary material

To view supplementary material for this article, please visit <https://doi.org/10.1017/S1751731119001605>

References

Ajibade DV, Dhawan P, Fechner AJ, Meyer MB, Pike JW and Christakos S 2010. Evidence for a role of prolactin in calcium homeostasis: regulation of intestinal transient receptor potential vanilloid type 6, intestinal calcium absorption, and the 25-hydroxyvitamin D(3) 1 α hydroxylase gene by prolactin. *Endocrinology* 151, 2974–2984.

Ardawi MS, Nasrat HA and BA'Aqueel HS 1997. Calcium-regulating hormones and parathyroid hormone-related peptide in normal human pregnancy and postpartum: a longitudinal study. *European Journal of Endocrinology* 137, 402–409.

Charoenphandhu N and Krishnamra N 2007. Prolactin is an important regulator of intestinal calcium transport. *Canadian Journal of Physiology and Pharmacology* 85, 569–581.

Charoenphandhu N, Teerapornpantakit J, Methawasin M, Wongdee K, Thongchote K and Krishnamra N 2008. Prolactin decreases expression of Runx2, osteoprotegerin, and RANKL in primary osteoblasts derived from tibiae of adult female rats. *Canadian Journal of Physiology and Pharmacology* 86, 240–248.

Chiwome B, Kandiwa E, Mushonga B, Sajeni S and Habarugira G 2017. A study of the incidence of milk fever in Jersey and Holstein cows at a dairy farm in Beatrice, Zimbabwe. *Journal of the South African Veterinary Association* 88, e1–e6.

Cowin P and Wysolmerski J 2010. Molecular mechanisms guiding embryonic mammary gland development. *Cold Spring Harbor Perspectives in Biology* 2, a003251.

DeGaris PJ and Lean IJ 2008. Milk fever in dairy cows: a review of pathophysiology and control principles. *Veterinary Journal* 176, 58–69.

DeGroot MA, Block E and French PD 2010. Effect of prepartum anionic supplementation on periparturient feed intake, health, and milk production. *Journal of Dairy Science* 93, 5268–5279.

Dishington IW 1975. Prevention of Milk Fever (Hypocalcemic Paresis Puerperalis) by Dietary Salt Supplements. *Acta Veterinaria Scandinavica* 16, 503–512.

Ender F, Dishington IW, Helge-Bostad A and Martinsons E 1962. Parturient paresis and related forms of hypocalcemic disorders induced experimentally in dairy cows. 2. Studies on the etiological importance of feeding prepartal diets high in calcium and low or normal in phosphorus in relation to development of milk fever. Effect of high and low alkalinity of diets. *Acta Veterinaria Scandinavica* 3 (suppl. 1), 5–52.

Goff J, Reinhardt T, Beitz D and Horst R 1995. Breed affects tissue vitamin D receptor concentration in periparturient dairy cows: a milk fever risk factor. *Journal of Dairy Science* 78 (suppl. 1), 184–184.

Goff JP 2008. The monitoring, prevention, and treatment of milk fever and subclinical hypocalcemia in dairy cows. *Veterinary Journal* 176, 50–57.

Hernandez LL 2017. Triennial lactation symposium/bolfa: serotonin and the regulation of calcium transport in dairy cows. *Journal of Animal Science* 95, 5711–5719.

Hernandez LL, Gregerson KA and Horseman ND 2012. Mammary gland serotonin regulates parathyroid hormone-related protein and other bone-related signals. *American Journal of Physiology-Endocrinology and Metabolism* 302, E1009–E1015.

Hernandez LL, Limesand SW, Collier JL, Horseman ND and Collier RJ 2009. The bovine mammary gland expresses multiple functional isoforms of serotonin receptors. *Journal of Endocrinology* 203, 123–131.

Hernández-Castellano LE, Hernandez LL, Sauerwein H and Bruckmaier RM 2017a. Endocrine and metabolic changes in transition dairy cows are affected by prepartum infusions of a serotonin precursor. *Journal of Dairy Science* 100, 5050–5057.

Hernández-Castellano LE, Özçelik R, Hernandez LL and Bruckmaier RM 2017b. Supplementation of colostrum and milk with 5-hydroxy-L-tryptophan affects immune factors but not growth performance in newborn calves. *Journal of Dairy Science* 101 794–800.

Hernández-Castellano LE, Weaver SR, Hernandez LL and Bruckmaier RM 2017c. Increased serum serotonin improves parturient calcium homeostasis in dairy cows. *Journal of Dairy Science* 100, 1580–1587.

Hibbs JW and Pounden WD 1955. Studies on milk fever in dairy cows. 4. Prevention by short-time, prepartum feeding of massive doses of vitamin-D. *Journal of Dairy Science* 38, 65–72.

Horst RL, Goff JP and Reinhardt TA 1994. Calcium and vitamin D metabolism in the dairy cow. *Journal of Dairy Science* 77, 1936–1951.

Khachlouf K, Hamed H, Gdoura R and Gargouri A 2019. Effects of dietary Zeolite supplementation on milk yield and composition and blood minerals status in lactating dairy cows. *Journal of Applied Animal Research* 47, 54–62.

Kovacs CS 2011. Calcium and bone metabolism disorders during pregnancy and lactation. *Endocrinology and Metabolism Clinics of North America* 40, 795–826.

Kovacs CS 2012. The role of vitamin D in pregnancy and lactation: insights from animal models and clinical studies. *Annual Review of Nutrition* 32, 97–123.

Kovacs CS and Kronenberg HM 1997. Maternal-fetal calcium and bone metabolism during pregnancy, puerperium, and lactation. *Endocrine Reviews* 18, 832–872.

Kronqvist C, Emanuelson U, Sporndly R and Holtenius K 2011. Effects of prepartum dietary calcium level on calcium and magnesium metabolism in periparturient dairy cows. *Journal of Dairy Science* 94, 1365–1373.

Kronqvist C, Emanuelson U, Traven M, Sporndly R and Holtenius K 2012. Relationship between incidence of milk fever and feeding of minerals during the last 3 weeks of gestation. *Animal* 6, 1316–1321.

- Laporta J, Keil KP, Vezina CM and Hernandez LL 2014. Peripheral serotonin regulates maternal calcium trafficking in mammary epithelial cells during lactation in mice. *PLoS ONE* 9, e110190.
- Laporta J, Moore SA, Peters MW, Peters TL and Hernandez LL 2013. Short communication: Circulating serotonin (5-HT) concentrations on day 1 of lactation as a potential predictor of transition-related disorders. *Journal of Dairy Science* 96, 5146–5150.
- Leno BM, Ryan CM, Stokol T, Kirk D, Zanzalari KP, Chapman JD and Overton TR 2017. Effects of prepartum dietary cation-anion difference on aspects of peripartum mineral and energy metabolism and performance of multiparous Holstein cows. *Journal of Dairy Science* 100, 4604–4622.
- Lieben L, Carmeliet G and Masuyama R 2011. Calcemic actions of vitamin D: effects on the intestine, kidney and bone. *Best Practice & Research: Clinical Endocrinology & Metabolism* 25, 561–572.
- Lotinun S, Limlomwongse LC, Sirikulchayanonta V and Krishnamra N 2003. Bone calcium turnover, formation, and resorption in bromocriptine- and prolactin-treated lactating rats. *Endocrine* 20, 163–170.
- Martinez N, Rodney RM, Block E, Hernandez LL, Nelson CD, Lean IJ and Santos JEP 2018. Effects of prepartum dietary cation-anion difference and source of vitamin D in dairy cows: health and reproductive responses. *Journal of Dairy Science* 101, 2563–2578.
- Martin-Tereso J, Martens H, Deiner C, van Laar H, den Hartog LA and Verstegen MWA 2016. Pre-calving feeding of rumen-protected rice bran to multiparous dairy cows improves recovery of calcaemia after calving. *Journal of Dairy Research* 83, 281–288.
- Matsuda M, Imaoka T, Vomachka AJ, Gudelsky GA, Hou Z, Mistry M, Bailey JP, Nieport KM, Walther DJ, Bader M and Horseman ND 2004. Serotonin regulates mammary gland development via an autocrine-paracrine loop. *Developmental Cell* 6, 193–203.
- Oetzel GR 1988. Parturient paresis and hypocalcemia in ruminant livestock. *Veterinary Clinics of North America: Food Animal Practice* 4, 351–364.
- Özçelik R, Bruckmaier RM and Hernández-Castellano LE 2017. Prepartum daylight exposure increases serum calcium concentrations in dairy cows at the onset of lactation. *Journal of Animal Science* 95, 4440–4447.
- Reinhardt TA, Lippolis JD, McCluskey BJ, Goff JP and Horst RL 2011. Prevalence of subclinical hypocalcemia in dairy herds. *Veterinary Journal* 188, 122–124.
- Roche JR and Berry DP 2006. Periparturient climatic, animal, and management factors influencing the incidence of milk Fever in grazing systems. *Journal of Dairy Science* 89, 2775–2783.
- Salari P and Abdollahi M 2014. The influence of pregnancy and lactation on maternal bone health: a systematic review. *Journal of Family and Reproductive Health* 8, 135–148.
- Slater CJ, Endres EL, Weaver SR, Cheng AA, Lauber MR, Endres SF, Olstad E, DeBruin A, Crump PM, Block E and Hernandez LL 2018. Interaction of 5-hydroxy-L-tryptophan and negative dietary cation-anion difference on calcium homeostasis in multiparous peripartum dairy cows. *Journal of Dairy Science* 101, 5486–5501.
- Thilsing-Hansen T, Jorgensen RJ, Enemark JM and Larsen T 2002a. The effect of zeolite A supplementation in the dry period on periparturient calcium, phosphorus, and magnesium homeostasis. *Journal of Dairy Science* 85, 1855–1862.
- Thilsing-Hansen T, Jorgensen RJ and Ostergaard S 2002b. Milk fever control principles: a review. *Acta Veterinaria Scandinavica* 43, 1–19.
- Urena P, Ferreira A, Kung VT, Morieux C, Simon P, Ang KS, Souberbielle JC, Segre GV, Druke TB and Devernejoul MC 1995. Serum pyridinoline as a specific marker of collagen breakdown and bone metabolism in hemodialysis-patients. *Journal of Bone and Mineral Research* 10, 932–939.
- van Bezooijen RL, Roelen BAJ, Visser A, van der Wee-Pals L, de Wilt E, Karperien M, Hamersma H, Papapoulos SE, ten Dijke P and Lowik CWGM 2004. Sclerostin is an osteocyte-expressed negative regulator of bone formation, but not a classical BMP antagonist. *Journal of Experimental Medicine* 199, 805–814.
- Vieira-Neto A, Lima IRP, Lopes F, Jr., Lopera C, Zimpel R, Sinedino LDP, Jeong KC, Galvao K, Thatcher WW, Nelson CD and Santos JEP 2017. Use of calcitriol to maintain postpartum blood calcium and improve immune function in dairy cows. *Journal of Dairy Science* 100, 5805–5823.
- Weaver SR, Prichard AP, Endres EL, Newhouse SA, Peters TL, Crump PM, Akins MS, Crenshaw TD, Bruckmaier RM and Hernandez LL 2016. Elevation of circulating serotonin improves calcium dynamics in the peripartum dairy cow. *Journal of Endocrinology* 230, 105–123.
- Weaver SR, Prichard AS, Maerz NL, Prichard AP, Endres EL, Hernandez-Castellano LE, Akins MS, Bruckmaier RM and Hernandez LL 2017. Elevating serotonin pre-partum alters the Holstein dairy cow hepatic adaptation to lactation. *PLoS ONE* 12, e0184939.
- Weiss WP, Azem E, Steinberg W and Reinhardt TA 2015. Effect of feeding 25-hydroxyvitamin D3 with a negative cation-anion difference diet on calcium and vitamin D status of periparturient cows and their calves. *Journal of Dairy Science* 98, 5588–5600.
- Wilkens MR, Liesegang A, Richter J, Fraser DR, Breves G and Schroder B 2014. Differences in peripartal plasma parameters related to calcium homeostasis of dairy sheep and goats in comparison with cows. *Journal of Dairy Research* 81, 325–332.
- Wysolmerski JJ 2012. Osteocytic osteolysis: time for a second look? *Bonekey Reports* 1, 229.