

Review Paper

Cite this article: Baskin JM, Baskin CC (2023). A classification system for seed (diaspore) monomorphism and heteromorphism in angiosperms. *Seed Science Research* **33**, 193–202. <https://doi.org/10.1017/S0960258523000193>

Received: 19 July 2023
Revised: 9 September 2023
Accepted: 12 September 2023
First published online: 22 February 2024


Keywords:

amphi-basiscarpy; amphicarpy; basicarpy; chasmogamy; cleistogamy; geocarpy; heteroarthrocarpy; seed (diaspore) heteromorphism

Corresponding author:

Carol C. Baskin;
Email: Carol.Baskin@uky.edu

A classification system for seed (diaspore) monomorphism and heteromorphism in angiosperms

Jerry M. Baskin¹ and Carol C. Baskin^{1,2} 

¹Department of Biology, University of Kentucky, Lexington, KY40506, USA and ²Department of Plant and Soil Sciences, University of Kentucky, Lexington, KY 40546, USA

Abstract

‘Seed heteromorphism’ is a broadly- and loosely-defined term used to describe differences in size/mass, morphology, position on mother plants and ecological function (e.g. dispersal, dormancy/germination) of two or more seeds or other diaspores produced by an individual plant. The primary aim of this review paper was to characterize via an in-depth classification scheme the physical structural design (‘architecture’) of diaspore monomorphism and diaspore heteromorphism in angiosperms. The diaspore classification schemes of Mandák and Barker were expanded/modified, and in doing so some of the terminology that Zohary, Ellner and Shmida, and van der Pijl used for describing diaspore dispersal were incorporated into our system. Based on their (relative) size, morphology and position on the mother plant, diaspores of angiosperms were divided into two divisions and each of these into several successively lower hierarchical layers. Thus, our classification scheme, an earlier version of which was published in the second edition of ‘Seeds’ by Baskin and Baskin, includes not only heteromorphic but also monomorphic diaspores, the *Division* to which the diaspores of the vast majority of angiosperms belong. The scheme will be useful in describing the ecology, biogeography and evolution of seed heteromorphism in flowering plants.

Introduction

‘Seed (diaspore) heteromorphism’ is a broadly- and loosely-defined term used to describe differences in size/mass, morphology, position on mother plant and ecological function (e.g. dispersal, dormancy/germination) of two or more seeds or other diaspores produced by an individual plant. Thus, the term is applied to a variety of situations concerning degree of distinctness of differences in size/mass, morphology and position of diaspores on a plant. For example, *Aethionema arabicum* (Brassicaceae) produces two morphologically distinct aerial diaspores with no intermediates (Arshad et al., 2019), *Heterosperma pinnatum* (Asteraceae) produces two morphologically distinct diaspores connected by a series of morphologically intermediate ones (Venable et al., 1987, 1995; Martorell and Martínez-López, 2014) and *Ceratocarpus arenarius* (Amaranthaceae) produces two morphologically distinct ground-level diaspores and a series of aerial diaspores that differ continuously in size and morphology from top to bottom of the plant canopy (Lu et al., 2013). It is no wonder, then, that in a recent paper Scholl et al. (2020) stated that defining seed heteromorphism is a challenge.

The aim of this review paper is to provide an in-depth classification scheme, an earlier version of which was published in the second edition of ‘Seeds’ by Baskin and Baskin (2014), based on size, morphology and position on the mother plant that will give more exactness to use of the term ‘seed heteromorphism’. More generally, our aim was to characterize the diversity of structural design (‘architecture’) of diaspore monomorphism and heteromorphism in angiosperms.

Methods

Our classification scheme is based on information in the literature on the size/mass, morphology and position (e.g. aerial, basal and subterranean) on the mother plants of the diaspores of angiosperm taxa. Basically, we greatly expanded/modified the diaspore classification schemes of Mandák (1997) and Barker (2005), sometimes using terminology that Zohary (1937, 1962), Ellner and Shmida (1981) and van der Pijl (1982) applied to diaspore dispersal. Seeds (diaspores) were first divided into two major categories (monomorphic and heteromorphic) called divisions and each *Division* into several successively lower hierarchical layers.

Our scheme does not include the terms that Zohary (1937, 1962), Ellner and Shmida (1981), van der Pijl (1982) and/or Gutterman (1993, 1994b) used to describe agents/modes of dispersal such as anemochory (wind), hydrochory (water), ombrohydrochory (rain) and

© The Author(s), 2024. Published by Cambridge University Press. This is an Open Access article, distributed under the terms of the Creative Commons Attribution licence (<http://creativecommons.org/licenses/by/4.0/>), which permits unrestricted re-use, distribution and reproduction, provided the original article is properly cited.

zoochory (animals). Neither does it include genetic polymorphism, in which (1) two or more kinds of diaspores are produced by a species, (2) a population may be diaspore-monomorphic or dimorphic/polymorphic and (3) individual plants produce only one kind of diaspore, which is determined by Mendelian inheritance, i.e. a genetic segregation, not a somatic differentiation (see [Appendix A](#)).

In general, we follow Scholl et al.'s (2020) definition of 'seed heteromorphism.' According to these authors, variation in the morphology of heteromorphic diaspores can be discrete or continuous. However, if the variation is continuous the most extreme diaspores need to be widely divergent morphologically, such as occurs in *H. pinnatum* (see Introduction), for them to qualify as heteromorphic. Diaspores that do not meet one of these two criteria are, by default, considered to be monomorphic.

Results and discussion

The classification scheme we assembled for diaspore monomorphism and heteromorphism is shown in [Table 1](#). However, as pointed out by Imbert (2002) and Scholl et al. (2020) seed heteromorphism (and thus monomorphism) is not easy to define. According to Harper et al. (1970), most individual plants and populations have a normal or skewed (continuous) distribution for seed size or shape, but that 'It is, however, characteristic of some species to produce two or more sharply defined types of seed.' Thus, the word 'monomorphic' does not mean that all seeds on an individual plant have a single size/mass or morphology (or that they have the same degree of dormancy). In fact, there is considerable variation in these traits (especially mass) among monomorphic seeds on an individual plant and even among those on the same infructescence or within a fruit as various authors have reported (e.g. Janzen, 1977a, b; Ernst, 1981; Thompson, 1981; Pitelka et al., 1983; Stanton, 1984; Thompson, 1984; Wolf et al., 1986; Wulff, 1986; Michaels et al., 1988; McGinley, 1989; Winn, 1991; Fenner, 1992; Susko and Lovett-Doust, 2000). Monomorphic simply means that seeds cannot easily be sorted into two or more clearly-defined (distinct) groups based on traits such as size/mass and/or morphology. Monomorphic seeds may show differences in dormancy/germination: cryptic polymorphism (i.e. ecological differentiation in the absence of obvious morphological differences) of Venable (1985). Heteromorphic seeds, on the other hand, means that seeds on an individual plant readily can be sorted into two or more distinct groups that differ in size/mass and/or morphology.

An excellent example of cryptic seed heteromorphism has been reported by Liyanage et al. (2016) for two species of Fabaceae (*Bossiaea heterophylla* and *Viminaria juncea*) whose seeds have physical dormancy. They found that individual plants of the two species, which occur in fire-prone ecosystems in southeastern Australia, produce seeds with a high threshold temperature and a low threshold temperature for physical dormancy break. There were no significant differences in mass or visible differences in shape or color of high- and low threshold temperature seeds from individual plants of either species. Seeds with a low threshold temperature for dormancy break could germinate after exposure to temperatures of a low-intensity fire (40–60°C), whereas those with a high threshold temperature for dormancy break could germinate only after exposure to temperatures of a high-intensity fire (≥80°C). Further, under competition with seedlings of *Acacia linifolia* (Fabaceae), a co-occurring species, seedlings of *B. heterophylla*

emerging after low-intensity fire temperature grew better than those emerging after high-intensity fire temperature. Competition would be more intense following low- than high-intensity fire due to the survival of more plants in the low- than high-intensity-burned community. Thus, although low and high threshold temperature seeds did not exhibit differences in mass or morphology they differed in their dormancy-breaking response to fire intensity and in seedling growth response to competition resulting from different fire intensities.

Overall, we show that there is considerable structural diversity in design ('architecture') of diaspore-heteromorphic angiosperm species based on diaspore size, morphology and position on the mother plant and of diaspore-monomorphic angiosperms based on diaspore position (e.g. aerial, basal and subterranean) on the mother plant. To date, only a few hundred of the >250,000 extant angiosperm species have been reported to produce heteromorphic diaspores (Imbert, 2002; Wang et al., 2010; Baskin et al., 2014; Scholl et al., 2020; Zhang et al., 2020), apparently meaning that the vast majority of flowering plants produce monomorphic diaspores.

In a recent survey for homomorphic (=our monomorphic) and heteromorphic species in the North American deserts, using information in various floras for the area, Scholl et al. (2020) identified 458 monomorphic species and 101 heteromorphic taxa, of which 75 of the latter were annuals. They also reported that their study brought the total number of known seed heteromorphic species to 378. The flora of this area contains many annuals, the climate is arid/semiarid and amount and timing of rainfall is unpredictable, which are conditions that favor bet-hedging via diaspore heteromorphism as a life history strategy (Scholl et al., 2020; Gianella et al., (2021). Thus, the North American deserts undoubtedly are highly over-represented in proportion of heteromorphic species compared to other bioclimatic regions. In which case, we should expect that the proportion of diaspore-heteromorphic species in the world's flora is much lower than that in the North American deserts.

Mandák's (1997) classification scheme for seed (diaspore) heteromorphism divides heteromorphic diaspores into Amphicarp and Heterodiaspory and distinguishes three subgroups for the latter category: heterocarp (Heteromericarp of van der Pijl, 1982), heteroarthrocarpy and heterospermy. (See [Table 1](#) for definition of each of these three terms and of those mentioned in the following.) Barker's (2005) diaspore classification scheme deals only with basicarp, geocarp and amphicarp. His scheme includes full geocarp, with three subtypes, i.e. active geocarp, geophytic geocarp and passive geocarp; and basicarp, also with three subtypes, i.e. aerial amphicarp, amphi-geocarp and amphi-basicarp. Imbert (2002) recognized two categories of diaspore heteromorphism: heterocarp and heterospermy.

Many species of grasses (Poaceae) produce cleistogamous (CL) axillary spikelets within leaf sheaths at nodes on flowering culms (Connor, 1979, 1981; Campbell et al., 1983). Some grasses, e.g. the well-studied species *Triplaris purpurea* (e.g. Chase, 1908, 1918; Cheplick, 1996; Cheplick and Sung, 1998), produce these clandestine spikelets at all nodes on the flowering culm, which is terminated by an inflorescence of chasmogamous (CH) spikelets. Caryopsis mass decreases continuously [in a log-linear (or nearly so) fashion] from the lowermost to one of the upper leaf sheaths,

Table 1. A classification system for monomorphic and heteromorphic diaspores in angiosperms based on size/mass, morphology and position on mother plant.

DIVISION I. Monomorphic. One type of fruit and one type of seed produced by an individual plant (see Discussion for further explanation).
Supergroup 1. Monomorphic aerial. Fruits/seeds produced on aerial stems (not including basal part of aerial stem at ground-level).
Group A. All fruits/seeds produced by chasmogamous (CH) flowers; includes most angiosperm species.
Group B. Fruits/seeds produced by both CH and cleistogamous (CL) flowers; includes a few hundred angiosperm species, e.g. many, but not all, <i>Viola</i> species (Violaceae) (Uphof, 1938; Lord, 1981; Culley and Klooster 2007).
Group C. All fruits/seeds produced by CL flowers (complete cleistogamy) (Uphof, 1938; Lord, 1981; Culley and Klooster, 2007), e.g. species in the mycoheterotropic genus <i>Gastrodia</i> (Orchidaceae) (Suetsuga, 2013, 2014, 2016).
Supergroup 2. Monomorphic basal or subterranean. One type of fruit and one type of seed produced at or near ground-level or underground.
Group A. Basicarpy. Flowers and fruits at or near ground-level but not subterranean, e.g. <i>Asteriscus pygmaeus</i> (Asteraceae) (Zohary, 1962; Ellner and Shmida, 1981), <i>Oenothera triloba</i> (Onagraceae) (Walck and Hidayati, 2007), <i>Trigonella stellata</i> (Fabaceae) (Zohary, 1962; Ellner and Shmida, 1981), holoparasitic plants [with ‘dust seeds’ and undifferentiated embryo (Baskin and Baskin 2022)] in Balanophoraceae and Cytinaceae (Barker, 2005).
Group B. Geocarpy. Subterranean fruits from subterranean or aboveground flowers.
Subgroup a. Full geocarpy (protogeocarpy sensu Zohary, 1962). Flowers and fruits subterranean, e.g. <i>Rhizanthella gardneri</i> (Orchidaceae) (Warcup, 1985).
Subgroup b. Active geocarpy (hysterogeocarpy sensu Zohary, 1962). Flowers aboveground. After fertilization, ovary is pushed or pulled underground, where the fruit develops, e.g. <i>Arachis hypogaea</i> (Fabaceae) (Smith, 1950), <i>Holocarpa</i> spp. (Asteraceae) (Barker, 2005) and <i>Trifolium subterraneum</i> (Fabaceae) (Darwin, 1888; Katznelson and Morley, 1965).
Subgroup c. Geophytic geocarpy. Stigma aboveground but ovary remains belowground at all times, e.g. the holoparasitic genera [with ‘dust seeds’ and undifferentiated embryo (Baskin and Baskin 2022)] <i>Hydnora</i> (Hydnoraceae) (Bolin et al., 2009) and <i>Lophophytum</i> (Balanophoraceae) (Borchsenius and Olesen, 1990) and Iridaceae species (Barker, 2005). In some geophytic species in the monocot families Amaryllidaceae, Colchicaceae and Iridaceae with bulbs or corms, the flower is raised into the air by elongation of the perianth tube, which acts as a pedicel, and the ovary remains belowground. The peduncle elongates after flowering and raises the fruit above the level of the soil surface. In <i>Syringodea leipoldtii</i> (Iridaceae), the upper part of the ovary is raised aboveground, and the lower part remains belowground. In <i>Crinum acaule</i> (Amaryllidaceae), the short peduncle may, or may not, lift the ovary above ground (Burt, 1970, 1977).
Subgroup d. Dioecious geocarpy. Male flowers produced on aerial stems of male plants and single-flowered female inflorescences produced belowground (or below ocean-sediment surface in seagrasses) on horizontal rhizomes of female plants. The ephemeral stigma protrudes aboveground (or above ocean-sediment surface in seagrasses), where it is pollinated by wind, e.g. <i>Alexgeorgea</i> (Restionaceae) (Carlquist, 1976; Meney et al., 1990), or water, e.g. geocarpy species in the seagrass genera <i>Cymodocea</i> and <i>Halodule</i> (Cymodoceaceae) and <i>Halophila</i> (Hydrocharitaceae) (Kuo and Kirkman, 1992; Ackerman, 2006; Orth et al., 2006).
Subgroup e. Passive geocarpy. Flowers and fruits at ground-level, but fruits become covered via natural soil (sand) disturbance, e.g. the sea strand plant <i>Arctotheca populifolia</i> (Asteraceae) (Barker, 2005)
DIVISION II. Heteromorphic [somatic polymorphism sensu Harper et al. (1970) and Harper (1977), which Harper (1977, p. 69) defines as ‘... the production of seeds of different morphologies or behavior on different parts of the same plant – not a genetic segregation but a somatic differentiation.’]. Two or more distinct kinds of fruits and/or seeds (produced by an individual plant) that differ in size/mass and/or morphology; fruits sometimes with accessory parts such as phyllary, bracteoles or perianth; different morphs usually show definite differences in dispersal and/or dormancy. The proportion of diaspore morphs produced can vary between years, habitats, populations and individuals within a population, and the variation may have a genetic, progeny- environmental and/or a maternal (and even a great grandmother or grandmother) environmental component (transgenerational plasticity) (Baker and O’Dowd, 1982; Cheplick and Quinn, 1983; Venable et al., 1987; Venable and Búrquez, 1989; 1995; Mandák and Pyšek, 1999; Imbert and Ronce, 2001; Wang and Wei, 2007; Lu et al., 2012, 2021; Talavera et al., 2012; Baskin and Baskin, 2014; Martorell and Martínez-López, 2014; Yang et al., 2015a,b; Zhang et al., 2017; Gan et al., 2020; Wang et al., 2021). For <i>Heterotheca subaxillaris</i> , an invasive species (from the USA) on the eastern shore of the Mediterranean Sea in Israel, Robinson et al. (2023) found that not only was the proportion of dispersing (disc) cypselae greater in the leading-edge populations than in the core populations but the investment in pappus of the dispersing cypselae was greater in the leading-edge populations. Some genetic aspects of diaspore heteromorphism are discussed by Baskin and Baskin (2014, see Table 8.14 and text page 351).
Group A. Heterodiaspory. Two or more types of diaspores (or of fruits within diaspores) produced aboveground (in some species including a few at or near ground-level, i.e. basal).
Subgroup a. Heterocarpy. Two, three or more kinds of fruits/seeds.
Type 1. Between-diaspore (inter-diaspore) variation. e.g. <i>Atriplex</i> spp. (Amaranthaceae) (Flores-Olvera et al., 2011), <i>Garhadiolus papposus</i> (Asteraceae) (Sun et al., 2009) and other Asteraceae species (Imbert, 2002), some of which have morphologically distinct diaspores connected by a series of morphologically intermediate ones [e.g. <i>Heterosperma pinnatum</i> (Asteraceae) (Venable et al., 1987)] and the <i>Rumex bucephalophorus</i> species complex (five subspecies) (Polygonaceae) [for plants of this species complex that produce aerial diaspores only (Talavera et al., 2010, 2011, 2012)]. See additional information about the <i>R. bucephalophorus</i> species complex under Amphicarpy sensu lato. Note that in addition to within-diaspore variation (see below), at least some species of <i>Aegilops</i> (Poaceae) exhibit between diaspore (inter-diaspore) variation within a spike, e.g. <i>A. cylindrica</i> (Donald and Ogg, 1991; Fandrich and Mallory-Smith, 2006) and <i>A. tauschii</i> (Wang et al., 2023). Thus, some <i>Aegilops</i> species fit into both between-diaspore and within-diaspore heteromorphism.
Type 2. Within-diaspore (intra-diaspore) variation (synaptospermy, i.e. two or more seeds or fruits joined together forming a compound diaspore, sensu Ellner and Shmida, 1981), e.g. <i>Aegilops</i> spp. (diaspore is spikelet or whole spike, depending on the species) (Waisel and Adler, 1959; Wurzbürger and Leshem, 1967; Datta et al., 1970; Wurzbürger et al., 1974; Wurzbürger and Koller, 1976; Marañón, 1989; Donald and Ogg, 1991; Dyer 2004; Fandrich and Mallory-Smith, 2006; Guzzon et al., 2018; Gianella et al., 2020, 2022; Wang et al., 2022, 2023); <i>Eremopyrum distans</i> (Poaceae, diaspore is a spikelet) (Wang et al., 2010); wild emmer wheat (<i>Triticum dicoccoides</i>) (Poaceae, diaspore is a spikelet) (Horovitz et al., 2013; Nave et al., 2016; Volis, 2016); and <i>Xanthium</i> spp. (Asteraceae, diaspore is an indehiscent bur with hooked prickles that develops from the involucre and contains two cypselae) (Arthur, 1895, 1906; Crocker, 1906; Wareing and Foda, 1957; Esashi and Leopold, 1968; Martin and Carnahan, 1983; Hockling and Liddle, 1986). If the seeds in the compound diaspore differ in dormancy-breaking/germination requirements, it is referred to as heteroblasty (Zohary and Imber, 1963). Thus, all the species mentioned in this section are heteroblastic.

Subgroup b. Heteroarthrocarpy (heteromericarpy *sensu van der Pijl, 1982*). One type of fruit with distinct proximal and distal segments, each typically bearing a single seed that is a distinct morph, e.g. *Cakile* spp. (Barbour, 1970; Rodman, 1974; Keddy, 1980; Payne and Maun, 1981; 1982; Hocking, 1982; Maun and Payne, 1989; Maun et al., 1990; Zhang, 1994; Donohue, 1997) and other species of Brassicaceae, tribe Brassiceae (Hall et al., 2011).

Subgroup c. Heterospermy. One or more types of fruits (F), without proximal/distal segments, containing seeds (S) that differ within and/or between fruits. Examples include (1) two types of fruits, each with a different type of seed (2S : 2F), e.g. annual species of *Aethionema* (Brassicaceae) (Zohary and Fahn, 1950; Arshad et al., 2019), *Diptychocarpus strictus* (Brassicaceae) (Lu et al., 2010) and *Suaeda aralocaspica* (Amaranthaceae) (Wang et al., 2008); (2) two types of fruits with three types of seeds (3S : 2F), e.g. *Ceratocarpus heterocarpa* (Fumariaceae) (Ruiz de Clavijo, 1994); and (3) one type of fruit with two types of seeds in same fruit (2S : 1F), e.g., *Capsella bursa-pastoris* (Brassicaceae) (Toorop et al., 2012; Gomez-Cabellos et al., 2022) and *Plantago coronopus* (Plantaginaceae) (Dowling, 1933; Schat, 1981).

Group B. Amphibasicarpy. Flowers and fruits produced at or near ground-level and on aerial parts of stem. Aerial and ground-level diaspores may differ in morphology and/or size.

Subgroup a. *sensu stricto*. Diaspores on aerial part of stem do not exhibit a continuous change in morphology or size, e.g. *Trionptiles solitaria* and some other Cyperaceae (Haines, 1971; Haines & Lye, 1977; Browning, 1992; Bruhl, 1994; Smith et al., 2006); and *Nassella clarazii* (Poaceae) (Lerner et al., 2008).

Subgroup b. *sensu lato*. Diaspores on aerial part of stem (canopy) exhibit a continuous change in morphology and size, e.g. *Ceratocarpus arenarius* (Amaranthaceae) (Lu et al., 2013). Cleistogamous grasses such as *Triplasis purpurea* (see Discussion), *Amphibromus scabrivalvis* (Cheplick and Clay, 1989) and *Achnatherum brachychaetum* (Lerner et al., 2008) might fit here.

Group C. Amphicarpy. One type of fruit or more than one type of fruit produced both above- and belowground (Note exception in the *Rumex bucephalophorus* species complex, Amphicarpy Subgroup b. *sensu lato*.) that differ in ecological function, e.g., dispersal and dormancy (Baskin and Baskin, 2014; Zhang et al., 2020).

Subgroup a. *sensu stricto* (amphi-geocarpy *sensu Barker, 2005*). Flowers aerial (CH or CH/CL) and subterranean (CL only); fruits aerial and subterranean. Examples include *Amphicarpeae bracteata* (Fabaceae), *Amphicarpum amphicarpon* (Poaceae), *Cardamine chenopodifolia* (Brassicaceae), *Commelina benghalensis* (Commelinaceae) and *Vigna minima* (Fabaceae) (Zhang et al., 2020). In *Amphicarpum* and three other genera of grasses, the subterranean spikelets with large caryopses are produced on specialized rhizomes called rhizanthogenes (Dobrenz and Beetle, 1966; Campbell et al., 1983).

Subgroup b. *sensu lato*. Flowers aerial (CH) and at or near ground-level (CH); fruits aerial and subterranean [or aerial and (basal → subterranean) in the *Rumex bucephalophorus* species complex (see below)], e.g., *Catananche lutea* (Asteraceae) (Ruiz de Clavijo, 1995; Ruiz de Clavijo and Jiménez, 1998), *Emex spinosa* (Polygonaceae) (Ortiz et al., 2009), *Gymnarrhena micrantha* (Asteraceae) (Koller and Roth, 1964) [Various authors have reported erroneously that subterranean fruits of *C. lutea*, *E. spinosa* and *G. micrantha* are produced by CL flowers, when, in fact, they are produced by CH flowers that are insect pollinated (see Baskin and Baskin, 2017).] and plants of the *R. bucephalophorus* species complex that produce both buried and aerial types of diaspores. Plants in some populations of the *R. bucephalophorus* species complex that form a basal rosette may produce female flowers and mature diaspores in the leaf axils that are buried by the action of contractile roots (buried diaspores) as well as aerial hermaphrodite flowers and diaspores. Plants that do not form a rosette produce only aerial flowers and diaspores. Altogether, one buried and three aerial diaspore types are produced by this species complex (Talavera et al., 2010, 2011, 2012). *C. lutea* and *G. micrantha* are examples of Asteraceae species with two types of capitula, i.e. aerial and basal (partly underground) (Burt, 1977).

above which there is little or no change in mass of caryopses, including those in the terminal CH spikelets. Chase (1908, 1918) used the term ‘cleistogene’ to describe the solitary sessile single floret with palea and lemma but without glumes in the lower leaf sheaths of *T. purpurea*. Chase (1918) applied the term ‘chasmogene’ to the terminal (‘ordinary’) spikelet. Her illustrations clearly show that the caryopsis from the cleistogene is much larger than that from the chasmogene. We have (cautiously) suggested that *T. purpurea* might fit subgroup B of amphibasicarpy (Table 1).

Monomorphic aerial CH/CL plants/populations may produce only CH or only CL flowers (Wilken, 1982; Sun, 1999; Hayamizu et al., 2014). *Corallorhiza bentleyi* (Orchidaceae), is an example of a species in which some populations produce only CL and others both CH and CL (see Freudenstein, 1999; Culley and Klooster, 2007). Small or young individuals of some CL species or individuals growing under unfavorable conditions produce only CL (Coker, 1962; Minter and Lord, 1983; Oakley et al., 2007; Hayamizu et al., 2014). *Epifagus virginiana* (Orobanchaceae) is an annual CL holoparasite with ‘dust seeds’ and an undifferentiated embryo that is host-specific on the roots of American beech (*Fagus grandifolia*). However, most, and sometimes all, of the flowers are CL, and the CH flowers usually are sterile. Thus, most seeds are produced by CL flowers (Schrenk, 1894; Cooke and Schively, 1904; Thieret, 1969, 1971; Musselman, 1982).

Lloyd and Schoen (1992) state that CH and CL seeds in most CL species differ in size, dispersal and germination. However, seeds from CH and CL flowers of our Group B of Supergroup I (monomorphic aerial) may or may not differ in these respects

(Baskin and Baskin, 2014, 2017). When there is a difference in mass of CH and CL seeds, the mass of CH seeds usually is greater than that of CL seeds (e.g. Cope, 1966; Hiratsuka and Inoue, 1988; Cheplick, 2005; Eckstein et al., 2006; Albert et al., 2011; Huebner, 2011; Munguía-Rosas et al., 2012). Differences in germination of CH and CL diaspores are more likely to occur in amphicarpy *sensu stricto* species than in aerial CL species. In 58 of 65 case studies (89.2%) for amphicarpy *sensu stricto* species, seeds from CH and CL flowers differed in germination percentage, whereas in aerial CL species 83 of 132 case studies (62.9%) of seeds from CH and CL flowers differed in germination percentage (Baskin and Baskin, 2017). Further study may show that a new category needs to be split out of Group B (Supergroup I, Division I) and incorporated into Division II. The new category would include species in which fruits/seeds produced by CH and CL flowers are morphologically distinct and (presumably) differ in ecology.

In *Ceratocarpus arenarius*, the basicarps differ in morphology and mass (and in embryo mass) from the aerial dispersal/germination units, which show continuous variation (increase or decrease) in morphology (see Supplementary Table S1 in Lu et al., 2013). Thus, there is discontinuous variation in this species between the basicarps (a) and aerial dispersal units (b)–(f) (Lu et al., 2013). Gao and Wei (2007) and Gao et al., (2008) recognized only two morphological types of fruits on plants of this species, namely subterranean (the two basicarps, which are, in fact, basal and not subterranean) and aerial.

In *Pisum fulvum* (Fabaceae), there is a gradient from amphicarpy plants (*sensu stricto*) with both aerial and subterranean

flowers and fruits to plants that produce only aerial flowers and fruits (Mattatia, 1977). One of the stages in the gradient is a basicarpic form that produces CH flowers near the soil surface, which Mattatia (1977) called 'subamphicarpic.' Thus, this species consists of both monomorphic (e.g. basicarpic form) and heteromorphic (e.g. amphicarpic form) plants.

Some species may exhibit plasticity as to the diaspore classification category to which they belong. For example, species that are amphicarpic and produce both aerial and subterranean diaspores under favorable environmental conditions may produce only underground fruits under stressful conditions and thus be 'facultatively geocarpic', e.g. *Amphicarpaea edgeworthii* (Fabaceae) (Zhang et al., 2017), *Amphicarpum amphicarpon* (Poaceae) (Cheplick and Quinn, 1983) and *Gymnarrhena micrantha* (Asteraceae) (Koller and Roth, 1964; Zeide, 1978; Loria and Noy-Meir, 1979/80). In which cases, individuals of these two annual species have the capacity to be either diaspore-heteromorphic (amphicarpic) or monomorphic (geocarpic).

Unlike these amphicarpic species, the cold desert annual diaspore-polymorphic amphibasicarpic species *Ceratocarpus arenarius* produces both basal (typically two) and aerial diaspores in the most stressful conditions in which it grows in its cold desert habitat. In the cold deserts of northwest China, we have observed that the smallest (5 cm tall, no branches) and largest (35 cm tall, bushy) plants of this species produce both basal and aerial diaspores, albeit in different basal morph:aerial morph and within-aerial morph ratios. Detailed experimental garden studies on the effect of abiotic (e.g. soil physicochemistry) and biotic (i.e. inter- and intraspecific competition) stress on phenotypic plasticity of the growth and reproduction of *C. arenarius*, including variation in diaspore morph ratios, have been published by Gan et al. (2020) and Lu et al. (2021).

Concluding remarks

We have documented via a hierarchical-based classification system the considerable diversity in structural design ('architecture') of diaspore monomorphism and heteromorphism in angiosperms. The scheme will enable investigators working on the broad topic of 'seed heteromorphism' to more precisely communicate their research to others, in part at least by giving more exactness to the term. It also will aid plant biologists in the preparation of a profile (spectrum) of the kinds (hierarchical categories) of diaspore monomorphism and heteromorphism for the various ecological and biogeographical units on earth. Finally, a detailed classification scheme that includes both diaspore monomorphism and heteromorphism is required for addressing the phylogenetic/evolutionary aspects of 'seed heteromorphism' in angiosperms (e.g. see Fernández et al., 2001; Cruz-Mazo et al., 2009, 2010). All that being said, however, it is likely that more hierarchical categories will need to be added to our system and/or existing ones revised/refined as literature and field research continues on diaspore monomorphism and heteromorphism in angiosperms.

Conflict of interest. The authors declare that they have no competing interests.

References

Abbott RJ (1986) Life history variation associated with the polymorphism for capitulum type and outcrossing rate in *Senecio vulgaris* L. *Heredity* **56**, 381–391.

- Abbott RJ** (1992) Plant invasions, interspecific hybridization and the evolution of new plant taxa. *Trends in Ecology and Evolution* **7**, 401–406.
- Abbott RJ and Horrill JC** (1991) Survivorship and fecundity of the radiate and non-radiate morphs of groundsel, *Senecio vulgaris* L., raised in pure stand and mixture. *Journal of Evolutionary Biology* **4**, 241–257.
- Abbott RJ, Horrill JC and Noble GDG** (1988) Germination behavior of the radiate and non-radiate morphs of groundsel, *Senecio vulgaris* L. *Heredity* **60**, 15–20.
- Abbott RJ, Ashton PA and Forbes DG** (1992) Introgressive origin of the radiate groundsel, *Senecio vulgaris* L. var. *hibernicus* Syme: Aat-3 evidence. *Heredity* **68**, 425–436.
- Abbott RJ, Bretagnolle FD and Thébaud C** (1998) Evolution of a polymorphism for outcrossing rate in *Senecio vulgaris*: influence of germination behavior. *Evolution* **52**, 1593–1601.
- Ackerman JD** (2006) Sexual reproduction of seagrasses: pollination in the marine context, pp. 89–109 in Larkum AWD; Orth RJ; Duarte CM (Eds) *Seagrasses: Biology, Ecology and Conservation*, Dordrecht, Springer.
- Albert LP, Campbell LG and Whitney KD** (2011) Beyond simple reproductive assurance: cleistogamy allows adaptive plastic responses to pollen limitation. *International Journal of Plant Sciences* **172**, 862–869.
- Arshad A, Sperber K, Steinbrecher T, Nichols B, Jansen VAA, Leubner-Metzger G and Mummenhoff K** (2019) Dispersal biophysics and adaptive significance of dimorphic diaspores in the annual *Aethionema arabicum* (Brassicaceae). *New Phytologist* **221**, 1434–1446.
- Arthur JC** (1895) Delayed germination of cocklebur and other paired seeds. *Proceedings of the Sixteenth Annual Meeting of the Society for the Promotion of Agricultural Science* **16**, 70–79.
- Arthur JC** (1906) The paired seeds of cocklebur. *The Plant World* **9**, 227–232.
- Baker GA and O'Dowd DJ** (1982) Effects of parent plant density on the production of achene types in the annual *Hypochoeris glabra*. *Journal of Ecology* **70**, 201–215.
- Barbour MG** (1970) Germination and early growth of the strand plant *Cakile maritima*. *Bulletin of the Torrey Botanical Club* **97**, 13–22.
- Barker NP** (2005) A review and survey of basicarpy, geocarpy, and amphicarpy in the African and Madagascan flora. *Annals of the Missouri Botanical Garden* **92**, 445–462.
- Baskin CC and Baskin JM** (2014) *Seeds: Ecology, Biogeography, and Evolution of Dormancy and Germination* (2nd Edn). San Diego, Elsevier/Academic Press.
- Baskin JM and Baskin CC** (2017) Seed germination in cleistogamous species: theoretical considerations and a literature survey of experimental results. *Seed Science Research* **27**, 84–98.
- Baskin JM and Baskin CC** (2022) Germination and seed/embryo size in holoparasitic flowering plants with "dust seeds" and an undifferentiated embryo. *The Botanical Review* **88**, 1–49.
- Baskin JM, Lu JJ, Baskin CC, Tan DY and Wang L** (2014) Diaspore dispersal ability and degree of dormancy in heteromorphic species of cold deserts of northwest China: a review. *Perspectives in Plant Ecology, Evolution and Systematics* **16**, 93–99.
- Belyayev A and Raskina O** (2013) Chromosomes evolution in marginal populations of *Aegilops speltoides* and consequences. *Annals of Botany* **111**, 531–538.
- Bolin JF, Maass E, Tennakoon KU and Musselman L** (2009) Host-specific germination of the root holoparasite *Hydnora triceps* (Hydnoraceae). *Botany* **87**, 1250–1254.
- Borchsenius F and Olesen JM** (1990) The Amazonian root holoparasite *Lophohytum mirabile* (Balanophoraceae) and its pollinators and herbivores. *Journal of Tropical Ecology* **6**, 501–505.
- Browning J** (1992) Hypogynous bristles and scales in basal florets in amphicarpous *Schoenoplectus* species (Cyperaceae). *Nordic Journal of Botany* **12**, 171–175.
- Bruhl JJ** (1994) Amphicarpy in the Cyperaceae, with novel variation in the wetland sedge *Eleocharis caespitosissima* Baker. *Australian Journal of Botany* **42**, 441–448.
- Burt BL** (1970) The evolution and taxonomic significance of a subterranean ovary in certain monocotyledons. *Israel Journal of Botany* **19**, 77–90.
- Burt BL** (1977) Aspects of diversification in the capitulum, pp. 41–59 in Heywood VH; Harborne JB; Turner BL (Eds) *The Biology and Chemistry of the Compositae*, London, Academic Press.

- Campbell CS, Quinn JA, Cheplick GP and Bell TJ** (1983) Cleistogamy in grasses. *Annual Review of Ecology and Systematics* **13**, 411–441.
- Carey K and Ganders FR** (1980) Heterozygote advantage at the fruit wing locus in *Plectritis congesta* (Valerianaceae). *Evolution* **34**, 601–607.
- Carlquist S** (1976) *Alexgeorgea*, a bizarre new genus of Restionaceae from Western Australia. *Australian Journal of Botany* **24**, 281–295.
- Ceynowa-Gieldon M** (1993) The variability of seeds in *Spergularia marina* (L.) Griseb. (= *S. salina* J. et C. Presl) growing on seacoast and inland saline soils in Poland. *Acta Societatis Botanicorum Poloniae* **62**, 185–191.
- Chapman MA and Abbott RJ** (2010) Introgression of fitness genes across a ploidy barrier. *New Phytologist* **186**, 63–71.
- Chase A** (1908) Notes on cleistogamy of grasses. *Botanical Gazette* **45**, 135–136.
- Chase A** (1918) Axillary cleistogenes in some American grasses. *American Journal of Botany* **5**, 254–258.
- Cheplick GP** (1996) Cleistogamy and seed heteromorphism in *Triplasis purpurea* (Poaceae). *Bulletin of the Torrey Botanical Club* **123**, 25–33.
- Cheplick GP** (2005) Biomass partitioning and reproductive allocation in the invasive, cleistogamous grass *Microstegium vimineum*: influence of the light environment. *Bulletin of the Torrey Botanical Society* **132**, 214–224.
- Cheplick GP and Clay K** (1989) Convergent evolution of cleistogamy and seed heteromorphism in two perennial grasses. *Evolutionary Trends in Plants* **3**, 127–136.
- Cheplick GP and Quinn JA** (1983) The shift in aerial/subterranean fruit ratio in *Amphicarpum purshii*: causes and significance. *Oecologia* **57**, 374–379.
- Cheplick GP and Sung LY** (1998) Effects of maternal nutrient environment and maturation position on seed heteromorphism, germination, and seedling growth in *Triplasis purpurea* (Poaceae). *International Journal of Plant Sciences* **159**, 338–350.
- Coker PD** (1962) Biological flora of the British Isles. *Corrigiola litoralis* L. *Journal of Ecology* **50**, 833–840.
- Connor HE** (1979) Breeding systems in the grasses: a survey. *New Zealand Journal of Botany* **17**, 547–574.
- Connor HE** (1981) Evolution of reproductive systems in the Gramineae. *Annals of the Missouri Botanical Garden* **68**, 48–74.
- Cope WA** (1966) Growth rate and yield in sericea lespedeza in relation to seed size and outcrossing. *Crop Science* **6**, 566–568.
- Cooke E and Schively AF** (1904) *Observations on the Structure and Development of Epiphegus virginiana*, vol. 2. *Contributions from the Botanical Laboratory of the University of Pennsylvania*. Philadelphia, University of Pennsylvania Press. pp. 352–398. + plates XXIX–XXXII.
- Crocker W** (1906) Role of seed coats in delayed germination. *Botanical Gazette* **42**, 265–291.
- Cruz-Mazo G, Buide ML, Samuel R and Narbona E** (2009) Molecular phylogeny of *Scorzoneroides* (Asteraceae): evolution of heterocarpy and annual habit in unpredictable environments. *Molecular Phylogenetics and Evolution* **53**, 835–847.
- Cruz-Mazo G, Narbona E and Buide ML** (2010) Germination patterns of dimorphic achenes in three related species of *Scorzoneroides* (Asteraceae, Lactuceae) growing in different environments. *Annales Botanici Fennici* **47**, 337–345.
- Culley TM and Klooster MR** (2007) The cleistogamous breeding system: a review of its frequency, evolution, and ecology in angiosperms. *The Botanical Review* **73**, 1–30.
- Darwin C** (1888) *The Power of Movement in Plants*. New York, D. Appleton and Company.
- Datta SC, Evenari M and Gutterman Y** (1970) The heteroblasty of *Aegilops ovata* L. *Israel Journal of Botany* **19**, 463–483.
- Dobrenz AK and Beetle AA** (1966) Cleistogones in *Danthonia*. *Journal of Range Management* **19**, 292–296.
- Donald WE and Ogg AG Jr** (1991) Biology and control of jointed goatgrass (*Aegilops cylindrica*), a review. *Weed Technology* **5**, 3–17.
- Donohue K** (1997) Seed dispersal in *Cakile edentula* var. *lacustris*: decoupling the fitness effects of density and distance from the home site. *Oecologia* **110**, 520–527.
- Dowling RE** (1933) The reproduction of *Plantago coronopus*: an example of morphological and biological seed dimorphism. *Annals of Botany* **47**, 861–872.
- Dyer AR** (2004) Maternal and sibling factors induce dormancy in dimorphic seed pairs of *Aegilops triuncialis*. *Plant Ecology* **172**, 211–218.
- Eckstein RL, Hölzel N and Danihelka J** (2006) Biological flora of Central Europe: *Viola elatior*, *V. pumila* and *V. stagnina*. *Perspectives in Plant Ecology, Evolution and Systematics* **8**, 45–66.
- Eggers Ware DM** (1983) Genetic fruit polymorphism in North American *Valerianella* (Valerianaceae) and its taxonomic implications. *Systematic Botany* **8**, 33–44.
- Ellner S and Shmida A** (1981) Why are adaptations for long-range seed dispersal rare in desert plants? *Oecologia* **51**, 133–144.
- Ernst WHO** (1981) Ecological implication of fruit variability in *Phleum arenarium* L., an annual dune grass. *Flora* **171**, 387–398.
- Esashi Y and Leopold AC** (1968) Physical forces in dormancy and germination of *Xanthium* seeds. *Plant Physiology* **43**, 871–876.
- Fandrich L and Mallory-Smith CA** (2006) Factors affecting germination of jointed goatgrass (*Aegilops cylindrica*) seed. *Weed Science* **54**, 677–684.
- Fenner M** (1992) Environmental influences on seed size and composition. *Horticultural Reviews* **13**, 183–213.
- Fernández IA, Aguilar JF, Panero JL and Feliner GN** (2001) A phylogenetic analysis of *Doronicum* (Asteraceae, Senecioneae) based on morphological, nuclear ribosomal (ITS), and chloroplast (*trnL-F*) evidence. *Molecular Phylogenetics and Evolution* **20**, 41–64.
- Flores-Olvera H, Vrijdaghs A, Ochoterena H and Smets E** (2011) The need to re-investigate the nature of homoplastic characters: an ontogenetic case study of the ‘bracteoles’ in Atripliceae (Chenopodiaceae). *Annals of Botany* **108**, 847–865.
- Freudenstein JV** (1999) A new species of *Corallorhiza* (Orchidaceae) from West Virginia, U.S.A. *Novon* **9**, 511–513.
- Gan L, Lu J, Baskin JM, Baskin CC and Tan D** (2020) Phenotypic plasticity in diaspore production of a amphibasicarpic cold desert annual that produces polymorphic diaspores. *Scientific Reports* **10**, 11142.
- Ganders FR, Carey K and Griffiths AJF** (1977a) Natural selection for a fruit dimorphism in *Plectritis congesta* (Valerianaceae). *Evolution* **31**, 873–881.
- Ganders FR, Carey K and Griffiths AJF** (1977b) Outcrossing rates in natural populations of *Plectritis brachystemon* (Valerianaceae). *Canadian Journal of Botany* **55**, 2070–2074.
- Gao R and Wei Y** (2007) Amphicarp of *Ceratocarpus arenarius* (Chenopodiaceae) in Junggar Desert. *Acta Botanica Yunnanica* **29**, 300–302. (in Chinese with English abstract)
- Gao R, Wei Y and Yan C** (2008) Amphicarp and seed germination behavior of *Ceratocarpus arenarius* L. (Chenopodiaceae). *Chinese Journal of Ecology* **27**, 23–27. (in Chinese with English abstract)
- Gianella M, Balestrazzi A, Pagano A, Müller JV, Kyratzis AC, Kikodze D, Canella M, Mondoni A, Rossi G and Guzzon F** (2020) Heteromorphic seeds of wheat wild relatives show germination niche differentiation. *Plant Biology* **22**, 191–202.
- Gianella M, Balestrazzi A, Ravasio A, Mondoni A, Börner A and Guzzon F** (2022) Comparative seed longevity under genebank storage and artificial ageing: a case study in heteromorphic wheat wild relatives. *Plant Biology* **24**, 836–845.
- Gianella M, Bradford KJ and Guzzon F** (2021) Ecological, (epi)genetic and physiological aspects of bet-hedging in angiosperms. *Plant Reproduction* **34**, 21–36.
- Gomez-Cabellos A, Toorop PE, Cañal MJ, Iannetta PPM, Fernández-Pascual E, Pritchard HW and Visscher AM** (2022) Global DNA methylation and cellular 5-methylcytosine and H4 acetylated patterns in primary and secondary dormant seeds of *Capsella bursa-pastoris* (L.) Medik. (shepherd’s purse). *Protoplasma* **259**, 595–614.
- Gutterman Y** (1993) *Seed Germination in Desert Plants*. Berlin, Springer-Verlag.
- Gutterman Y** (1994a) In memoriam – Michael Evenari and his desert. Seed dispersal and germination strategies of *Spergularia diandra* compared with some other desert annual plants inhabiting the Negev Desert of Israel. *Israel Journal of Plant Sciences* **42**, 261–274.
- Gutterman Y** (1994b) Strategies of seed dispersal and germination in plants inhabiting deserts. *The Botanical Review* **60**, 373–425.
- Gutterman Y** (1996) Environmental influences during seed maturation and storage affecting germinability in *Spergularia diandra* genotypes inhabiting the Negev Desert, Israel. *Journal of Arid Environments* **34**, 313–323.

- Gutterman Y** (1997a) Effect of daylength on flowering and seed morphology of *Spergularia diandra* occurring in the Negev Desert, Israel. *Journal of Arid Environments* **36**, 611–622.
- Gutterman Y** (1997b) Genotypic, phenotypic and opportunistic germination strategies of some common desert annuals compared with plants with other seed dispersal and germination strategies, pp. 611–622 in Ellis RH; Black M; Murdoch AJ; Hong TD (Eds) *Basic and Applied Aspects of Seed Biology*, Dordrecht, Kluwer Academic Publishers.
- Gutterman Y** (1998) Ecological strategies of desert annual plants, pp. 203–231 in Ambasht RS (Ed) *Modern Trends in Ecology and Environment*, Leiden, Backhuys Publishers.
- Gutterman Y** (2000) Environmental factors and survival strategies of annual plant species in the Negev Desert, Israel. *Plant Species Biology* **15**, 113–125.
- Guzzon F, Orsenigo S, Gianella M, Müller JV, Vagge I, Rossi G and Mondoni A** (2018) Seed heteromorphy influences seed longevity in *Aegilops*. *Seed Science Research* **28**, 277–285.
- Haines R** (1971) Amphicarp in east African Cyperaceae. *Mitteilungen der Botanischen Staatssammlung München* **10**, 534–538.
- Haines RW and Lye KA** (1977) Studies in African Cyperaceae. XV. Amphicarp and spikelet structure in *Trianoptiles solitaria*. *Botaniska Notiser* **130**, 235–240.
- Hall JC, Tisdale TE, Donohue K, Wheeler A, Al-Yahya MA and Kramer EM** (2011) Convergent evolution of a complex fruit structure in tribe Brassiceae (Brassicaceae). *American Journal of Botany* **98**, 1989–2003.
- Harper JL** (1977) *Population biology of plants*. London, Academic Press.
- Harper JL, Lovell PH and Moore KG** (1970) The shapes and sizes of seeds. *Annual Review of Ecology and Systematics* **1**, 327–356.
- Hayamizu M, Hosokawa I, Kimura O and Ohara M** (2014) Intraspecific variation in life history traits of *Viola brevistipulata* (Violaceae) in Hokkaido. *Plant Species Biology* **29**, 85–91.
- Hiratsuka A and Inoue O** (1988) Dispersability of dimorphic seeds in *Impatiens noli-tangere* and *I. textori* (Balsaminaceae). *Ecological Review* **21**, 157–161.
- Hocking PJ** (1982) Salt and mineral nutrient levels in fruits in two strand species, *Cakile maritima* and *Arctotheca populifolia*, with special reference to the effect of salt on the germination of *Cakile*. *Annals of Botany* **50**, 335–343.
- Hocking PJ and Liddle MJ** (1986) The biology of Australian weeds: 15. *Xanthium occidentale* Bertol. complex and *Xanthium spinosum* L. *Journal of the Australian Institute of Agricultural Science* **52**, 191–221.
- Horowitz A, Ezrati S and Anikster Y** (2013) Are soil seed banks relevant for agriculture in our day? *Crop Wild Relatives* **9**, 27–30.
- Huebner CD** (2011) Seed mass, viability, and germination of Japanese stilt-grass (*Microstegium vimineum*) under variable light and moisture conditions. *Invasive Plant Science and Management* **4**, 274–283.
- Imbert E** (2002) Ecological consequences and ontogeny of seed heteromorphism. *Perspectives in Plant Ecology, Evolution and Systematics* **5**, 13–36.
- Imbert E and Ronce O** (2001) Phenotypic plasticity for dispersal ability in the seed heteromorphic *Crepis sancta* (Asteraceae). *Oikos* **93**, 126–134.
- Ingram R, Weir J and Abbott RJ** (1980) New evidence concerning the origin of inland radiate groundsel, *S. vulgaris* L. var. *hibernicus* Syme. *New Phytologist* **84**, 543–546.
- Janzen DH** (1977a) Variation in seed weight in Costa Rican *Cassia grandis* (Leguminosae). *Tropical Ecology* **18**, 177–186.
- Janzen DH** (1977b) Variation in seed size within a crop of a Costa Rican *Mucuna andreana* (Leguminosae). *American Journal of Botany* **64**, 347–349.
- Jones TA and Nielson DC** (1999) Intrapopulation genetic variation for seed dormancy in Indian ricegrass. *Journal of Range Management* **52**, 646–650.
- Jones A, Redinbaugh MG, Larson SR, Zhang Y and Dow BD** (2007) Polymorphic Indian ricegrass populations result from overlapping waves of migration. *Western North American Naturalist* **67**, 338–346.
- Katznelson J and Morley FHW** (1965) A taxonomic revision of sect. *Calycomorphum* of the genus *Trifolium*. I. The geocarpic species. *Israel Journal of Botany* **14**, 112–134.
- Keddy PA** (1980) Population ecology in an environmental mosaic: *Cakile edentula* on a gravel bar. *Canadian Journal of Botany* **58**, 1095–1100.
- Keddy PA** (1982) Population ecology on an environmental gradient: *Cakile edentula* on a sand dune. *Oecologia* **52**, 348–355.
- Koller D and Roth N** (1964) Studies on the ecological and physiological significance of amphicarp in *Gymnarrhena micrantha* (Compositae). *American Journal of Botany* **51**, 26–35.
- Kucewicz M and Gojlo E** (2013) Ecological importance of seed dimorphism in corn spurry (*Spergula arvensis* L.) 1. The effects of day length on seed germinability. *Polish Journal of Ecology* **61**, 665–673.
- Kuo J and Kirkman H** (1992) Fruits, seeds and germination in the seagrass *Halophila ovalis* (Hydrocharitaceae). *Botanica Marina* **35**, 197–204.
- Lerner PD, Bai Y and Morici EFA** (2008) Does seed heteromorphism have different roles in the fitness of species with contrasting life history strategies? *Botany* **86**, 1404–1415.
- Liyanage GS, Ayre DJ and Ooi MKJ** (2016) Seedling performance covaries with dormancy thresholds: maintaining cryptic seed heteromorphism in a fire-prone system. *Ecology* **97**, 3009–3018.
- Lloyd DG and Schoen DJ** (1992) Self- and cross-fertilization in plants. I. Functional dimensions. *International Journal of Plant Sciences* **153**, 358–369.
- Lord EM** (1981) Cleistogamy: a tool for the study of floral morphogenesis, function and evolution. *The Botanical Review* **47**, 421–449.
- Loria M and Noy-Meir I** (1979/80) Dynamics of some annual populations in a desert loess plain. *Israel Journal of Botany* **28**, 211–225.
- Lu JJ, Tan DY, Baskin JM and Baskin CC** (2010) Fruit and seed heteromorphism in the cold desert annual ephemeral *Diptychocarpus strictus* (Brassicaceae) and possible adaptive significance. *Annals of Botany* **105**, 999–1014.
- Lu JJ, Tan DY, Baskin JM and Baskin CC** (2012) Phenotypic plasticity and bet-hedging in a heterocarpic winter annual/spring ephemeral cold desert species of Brassicaceae. *Oikos* **121**, 357–366.
- Lu JJ, Tan DY, Baskin JM and Baskin CC** (2013) Tradeoffs between seed dispersal and dormancy in an amphibasicarpic cold desert annual. *Annals of Botany* **112**, 1815–1827.
- Lu JJ, Gan L, Tan DY, Baskin JM and Baskin CC** (2021) Effects of the habitat-soil factor on transgenerational plasticity in a diaspore-polymorphic cold desert annual. *Plant and Soil* **461**, 355–367.
- Mandák B** (1997) Seed heteromorphism and the life cycle of plants: a review. *Preslia* **69**, 129–159.
- Mandák B and Pyšek P** (1999) Effects of plant density and nutrient levels on fruit polymorphism in *Atriplex sagittata*. *Oecologia* **119**, 63–72.
- Marañón T** (1989) Variations in seed size and germination in three *Aegilops* species. *Seed Science & Technology* **17**, 583–588.
- Martin RJ and Carnahan JA** (1983) The effect of field storage and laboratory conditions on germination of five *Xanthium* species. *Australian Journal of Agricultural Research* **34**, 249–260.
- Martorell C and Martínez-López M** (2014) Informed dispersal in plants: *Heterosperma pinnatum* (Asteraceae) adjusts its dispersal mode to escape from competition and water stress. *Oikos* **123**, 225–231.
- Mathez J and Xena de Enrich N** (1985a) Heterocarpy, fruit polymorphism and discriminating dissemination in the genus *Fedia* (Valerianaceae), pp. 431–441 in Jacquar P; Heim G; Antonovics J (Eds) *Genetic Differentiation and Dispersal in Plants*, Berlin, Springer-Verlag.
- Mathez J and Xena de Enrich N** (1985b) Le polymorphisme genetique de la morphologie des fruits du genre *Fedia* Gaertn. (Valerianaceae). I. Determination du mecanisme de controle genetique chez les especes *F. cornucopiae* (L.) Gaertn. et *F. graciliflora* Fisch. & Meyer. *Candollea* **40**, 425–434.
- Mattatia J** (1977) Amphicarp and variability in *Pisum fulvum*. *Botaniska Notiser* **130**, 27–34.
- Maun MA and Payne AM** (1989) Fruit and seed polymorphism and its relation to seedling growth in the genus *Cakile*. *Canadian Journal of Botany* **67**, 2743–2750.
- Maun MA, Boyd RS and Olson L** (1990) The biological flora of coastal dunes and wetlands. 1. *Cakile edentula* (Bigel.) Hook. *Journal of Coastal Research* **6**, 137–156.
- Mazer SJ and Lowrey DE** (2003) Environmental, genetic, and seed mass effects on winged seed production in the heteromorphic *Spergularia marina* (Caryophyllaceae). *Functional Ecology* **17**, 637–650.
- McGinley MA** (1989) Within and among plant variation in seed mass and pappus size in *Tragopogon dubious*. *Canadian Journal of Botany* **67**, 1298–1304.

- Memon RS, Bhatti GR, Khalid S, Arshad M, Mirbahar AA and Qureshi R** (2010) Microstructural features of seeds of *Spergularia marina* (L.) Griseb. (Caryophyllaceae). *Pakistan Journal of Botany* **42**, 1423–1427.
- Meney KA, Pate JS and Dixon KW** (1990) Comparative morphology, anatomy, phenology and reproductive biology of *Alexgeorgea* spp. (Restionaceae) from south-western Western Australia. *Australian Journal of Botany* **38**, 523–541.
- Michaels HJ, Benner B, Hartgerink AP, Lee TD and Rice S** (1988) Seed size variation: magnitude, distribution, and ecological correlates. *Evolutionary Ecology* **2**, 157–166.
- Minter TC and Lord EM** (1983) Effects of water stress, abscisic acid, and gibberellic acid on flower production and differentiation in the cleistogamous species *Collomia grandiflora* Dougl. ex Lindl. (Polemoniaceae). *American Journal of Botany* **70**, 618–624.
- Munguía-Rosas MA, Parra-Tabla V, Ollerton J and Cervera JC** (2012) Environmental control of reproductive phenology and the effect of pollen supplementation on resource allocation in the cleistogamous weed, *Ruellia nudiflora* (Acanthaceae). *Annals of Botany* **109**, 343–350.
- Musselman LJ** (1982) The Orobanchaceae of Virginia. *Castanea* **47**, 266–275.
- Nave M, Avni R, Ben-Zvi B, Hale I and Distelfeld A** (2016) QTLs for uniform grain dimensions and germination selected during wheat domestication are co-located on chromosome 4B. *Theoretical and Applied Genetics* **129**, 1303–1315.
- New JK** (1958) A population study of *Spergula arvensis*. I. Two clines and their significance. *Annals of Botany* **22**, 457–477.
- New JK** (1959) A population study of *Spergula arvensis*. II. Genetics and breeding behaviour. *Annals of Botany* **23**, 23–33.
- New JK** (1961) Biological flora of the British Isles. *Spergula arvensis* L. *Journal of Ecology* **49**, 205–215.
- New JK** (1978) Change and stability of clines in *Spergula arvensis* L. (corn spurry) after 20 years. *Watsonia* **12**, 137–143.
- New JK and Herriott JC** (1981) Moisture for germination as a factor affecting the distribution of the seed coat morphs of *Spergula arvensis* L. *Watsonia* **13**, 323–324.
- Oakley CG, Moriuchi KS and Winn AA** (2007) The maintenance of outcrossing in predominantly selfing species: ideas and evidence from cleistogamous species. *Annual Review of Ecology, Evolution and Systematics* **38**, 437–457.
- Orth RJ, Harwell MC and Inglis GJ** (2006) Ecology of seagrass seeds and seagrass dispersal processes, pp. 111–133 in Larkum AWD, Orth RJ, Duarte CM (Eds) *Seagrasses: Biology, Ecology and Conservation*, Dordrecht, Springer.
- Ortiz PL, Berjano R, Talavera M and Arista M** (2009) The role of resources and architecture in modeling floral variability for the monoecious amphicarpic *Emex spinosa* (Polygonaceae). *American Journal of Botany* **96**, 2062–2073.
- Payne AM and Maun MA** (1981) Dispersal and floating ability of dimorphic fruit segments of *Cakile edentula* var. *lacustris*. *Canadian Journal of Botany* **59**, 2595–2602.
- Pitelka LF, Thayer ME and Hansen SB** (1983) Variation in achene weight in *Aster acuminatus*. *Canadian Journal of Botany* **61**, 1415–1420.
- Redbo-Torstensson P** (1994) Variation in plastic response to a salinity gradient within a population of the halophytic plant *Spergularia marina*. *Oikos* **70**, 349–358.
- Redbo-Torstensson P and Telenius A** (1995) Primary and secondary seed dispersal by wind and water in *Spergularia salina*. *Ecography* **18**, 230–237.
- Richards AJ** (1975) The inheritance and behavior of the rayed gene complex in *Senecio vulgaris*. *Heredity* **34**, 95–104.
- Robinson D, Bogdanova T, Cohen O and Gruntman M** (2023) Trade-off between dispersal traits in a heterocarpic plant across its invasive route. *American Journal of Botany* **110**, e16192.
- Rodman JE** (1974) Systematics and evolution of the genus *Cakile* (Cruciferae). *Contributions from the Gray Herbarium* **205**, 3–146.
- Ruban AS and Badaeva ED** (2018) Evolution of the S-genomes in *Triticum-Aegilops* alliance: evidences from chromosome analysis. *Frontiers in Plant Science* **9**, 1756.
- Ruiz de Clavijo E** (1994) Heterocarpy and seed polymorphism in *Ceratocarpus heterocarpa* (Fumariaceae). *International Journal of Plant Sciences* **155**, 196–202.
- Ruiz de Clavijo E** (1995) The ecological significance of fruit heteromorphism in the amphicarpic species *Catananche lutea* (Asteraceae). *International Journal of Plant Sciences* **156**, 824–833.
- Ruiz de Clavijo E and Jiménez MJ** (1998) The influence of achene type and plant density on growth and biomass allocation in the heterocarpic annual *Catananche lutea* (Asteraceae). *International Journal of Plant Sciences* **159**, 637–647.
- Salisbury EJ** (1958) *Spergularia salina* and *Spergularia marginata* and their heteromorphic seeds. *Kew Bulletin* **1**, 41–51.
- Schat H** (1981) Seed polymorphism and germination ecology of *Plantago coronopus* L. *Acta Oecologica* **2**, 367–380.
- Scholl JP, Calle L, Miller N and Venable DL** (2020) Offspring polymorphism and bet hedging: a large-scale, phylogenetic analysis. *Ecology Letters* **23**, 1223–1231.
- Schrenk H** (1894) Parasitism of *Epiphegus* [*Epifagus*] *virginiana*. *Proceedings of the American Microscopical Society* **15**, 91–127. Sixteenth Annual Meeting. Part II (January 1894), pp. 91–127 + plates I–X.
- Smith BW** (1950) *Arachis hypogaea*. Aerial flower and subterranean fruit. *American Journal of Botany* **37**, 802–815.
- Smith M, Ammann S, Parker N and Mettler-Cherry P** (2006) A quantitative study of styles and achenes of terminal and basal flowers of *Schoenoplectus hallii* (Cyperaceae), a rare plant species of transient wetland habitats. *Sida* **22**, 1159–1173.
- Stanton ML** (1984) Developmental and genetic sources of seed weight variation in *Raphanus raphanistrum* L. (Brassicaceae). *American Journal of Botany* **71**, 1090–1098.
- Sterk AA** (1969a) Biosystematic studies of *Spergularia media* and *S. marina* in The Netherlands I. The morphological variability of *S. media*. *Acta Botanica Neerlandica* **18**, 325–338.
- Sterk AA** (1969b) Biosystematic studies of *Spergularia media* and *S. marina* in The Netherlands II. The morphological variation of *S. marina*. *Acta Botanica Neerlandica* **18**, 467–476.
- Sterk AA** (1969c) Biosystematic studies of *Spergularia media* and *S. marina* in The Netherlands III. The variability of *S. media* and *S. marina* in relation to the environment. *Acta Botanica Neerlandica* **18**, 561–577.
- Sterk AA** (1969d) Biosystematic studies of *Spergularia media* and *S. marina* in The Netherlands IV. Reproduction, dissemination, karyogenetics and taxonomy. *Acta Botanica Neerlandica* **18**, 639–650.
- Sterk AA and Dijkhuizen L** (1972) The relation between the genetic determination and the ecological significance of the seed wing in *Spergularia media* and *S. marina*. *Acta Botanica Neerlandica* **21**, 481–490.
- Suetsuga K** (2013) *Gastrodia takeshimensis* (Orchidaceae), a new mycoheterotrophic species from Japan. *Annales Botanici Fennici* **50**, 375–378.
- Suetsuga K** (2014) *Gastrodia flexistyloides* (Orchidaceae), a new mycoheterotrophic plant with complete cleistogamy from Japan. *Phytotaxa* **175**, 270–274.
- Suetsuga K** (2016) *Gastrodia kuroshimensis* (Orchidaceae: Epidendroideae: Gastrodieae), a new mycoheterotrophic and complete cleistogamous plant from Japan. *Phytotaxa* **278**, 265–272.
- Sun M** (1999) Cleistogamy in *Scutellaria indica* (Labiatae): effective mating system and population genetic structure. *Molecular Ecology* **8**, 1285–1295.
- Sun HZ, Lu JJ, Tan DY, Baskin JM and Baskin CC** (2009) Dormancy and germination characteristics of the trimorphic achenes of *Garhadiolus papposus* (Asteraceae), an annual ephemeral from the Junggar Desert, China. *South African Journal of Botany* **75**, 537–545.
- Susko DJ and Lovett-Doust L** (2000) Patterns of seed mass variation and their effects on seedling traits in *Alliaria petiolata* (Brassicaceae). *American Journal of Botany* **87**, 56–66.
- Talavera M, Arista M and Ortiz PL** (2012) Evolution of dispersal traits in a biogeographical context: a study using the heterocarpic *Rumex bucephalophorus* as a model. *Journal of Ecology* **100**, 1194–1203.
- Talavera M, Ortiz PL, Arista M, Berjano R and Imbert E** (2010) Disentangling sources of maternal effects in the heterocarpic species *Rumex bucephalophorus*. *Perspectives in Plant Ecology, Evolution and Systematics* **12**, 295–304.
- Talavera M, Balao F, Casimiro-Soriguer R, Ortiz MÁ, Terrab A, Arista M, Ortiz PL, Stuessy TF and Talavera S** (2011) Molecular phylogeny and systematics of the highly polymorphic *Rumex bucephalophorus* complex (Polygonaceae). *Molecular Phylogenetics and Evolution* **61**, 659–670.

- Telenius A** (1992) Seed heteromorphism in a population of *Spergularia media* in relation to the ambient vegetation density. *Acta Botanica Neerlandica* **41**, 305–318.
- Telenius A and Torstensson P** (1989) The seed dimorphism of *Spergularia marina* in relation to dispersal by wind and water. *Oecologia* **80**, 206–210.
- Telenius A and Torstensson P** (1991) Seed wings in relation to seed size in the genus *Spergularia*. *Oikos* **61**, 216–222.
- Telenius A and Torstensson P** (1999) Seed type and seed size variation in the heteromorphic saltmarsh annual *Spergularia salina* along the coast of Sweden. *Plant Biology* **1**, 585–593.
- Thieret JW** (1969) Notes on *Epifagus*. *Castanea* **34**, 397–402.
- Thieret JW** (1971) The genera of Orobanchaceae in the southeastern United States. *Journal of the Arnold Arboretum* **52**, 404–434.
- Thompson PA** (1981) Variations in seed size within populations of *Silene dioica* (L.) Clairv. in relation to habitat. *Annals of Botany* **47**, 623–634.
- Thompson JN** (1984) Variation among individual seed masses in *Lomatium grayi* (Umbelliferae) under controlled conditions: magnitude and partitioning of the variance. *Ecology* **65**, 626–631.
- Toorop PE, Cuerva RC, Begg GS, Locardi B, Squire GR and Iannetta PPM** (2012) Co-adaptation of seed dormancy and flowering time in the arable weed *Capsella bursa-pastoris* (shepherd's purse). *Annals of Botany* **109**, 481–489.
- Trow AH** (1912) On the inheritance of certain characters in the common groundsel – *Senecio vulgaris*, Linn. – and its segregates. *Journal of Genetics* **2**, 239–276.
- Uphof JCT** (1938) Cleistogamic flowers. *The Botanical Review* **4**, 21–49.
- van der Pijl L** (1982) *Principles of dispersal in higher plants. Third revised and expanded edition*. Berlin, Springer.
- Venable DL** (1985) The evolutionary ecology of seed heteromorphism. *The American Naturalist* **126**, 577–595.
- Venable DL and Búrquez A** (1989) Quantitative genetics of size, shape, life-history, and fruit characteristics of the seed-heteromorphic composite *Heterosperma pinnatum*. I. Variation within and among populations. *Evolution* **43**, 113–124.
- Venable DL, Dyreson E and Morales E** (1995) Population dynamic consequences and evolution of seed traits of *Heterosperma pinnatum* (Asteraceae). *American Journal of Botany* **82**, 410–420.
- Venable DL, Búrquez A, Corral G, Morales E and Espinosa F** (1987) The ecology of seed heteromorphism in *Heterosperma pinnatum* in central Mexico. *Ecology* **68**, 65–76.
- Volis S** (2016) Seed heteromorphism in *Triticum dicoccoides*: association between seed positions within a dispersal unit and dormancy. *Oecologia* **181**, 401–412.
- Wagner LK** (1986) Variation in seed-coat morph ratios in *Spergula arvensis* L. *Bulletin of the Torrey Botanical Club* **113**, 28–35.
- Wagner LK** (1988) Germination and seedling emergence in *Spergula arvensis*. *American Journal of Botany* **75**, 465–475.
- Waisel Y and Adler Y** (1959) Germination behavior of *Aegilops kotschy* Boiss. *Canadian Journal of Botany* **37**, 741–742.
- Walck JL and Hidayati SN** (2007) Ombrohydrochory and its relationship to seed dispersal and germination strategies in two temperate North American *Oenothera* species (Onagraceae). *International Journal of Plant Sciences* **168**, 1279–1290.
- Wang H and Wei Y** (2007) Seed polymorphism and fruit-set of *Salsola affinis*. *Biodiversity Science* **15**, 419–424. (in Chinese with English abstract)
- Wang L, Huang Z, Baskin CC, Baskin JM and Dong M** (2008) Germination of dimorphic seeds of the desert annual halophyte *Suaeda aralocaspica* (Chenopodiaceae), a C₄ plant without Kranz anatomy. *Annals of Botany* **102**, 757–769.
- Wang AB, Tan DY, Baskin CC and Baskin JM** (2010) Effect of seed position in spikelet on life history of *Eremopyrum distans* (Poaceae) from the cold desert of northwest China. *Annals of Botany* **106**, 95–105.
- Wang Z, Baskin JM, Baskin CC, Yang X, Liu G, Ye X, Huang Z and Cornelissen JHC** (2021) Great granny still ruling from the grave: phenotypic response of plant performance and seed functional traits to salt stress affects multiple generations of a halophyte. *Journal of Ecology* **110**, 117–128.
- Wang AB, Baskin CC, Baskin JM and Ding J** (2022) Seed position in spikelet as a contributing factor to the success of the winter annual invasive grass *Aegilops tauschii*. *Frontiers in Plant Science* **13**, 91651.
- Wang AB, Baskin CC, Baskin JM and Ding J** (2023) Trade-offs between diaspore dispersal and dormancy within a spike of the invasive annual grass *Aegilops tauschii*. *Planta* **257**, 121.
- Warcup JH** (1985) *Rhizanthella gardneri* (Orchidaceae), its *Rhizoctonia* endophyte and close association with *Melaleuca uncinata* (Myrtaceae) in western Australia. *New Phytologist* **99**, 273–280.
- Wareing PF and Foda HA** (1957) Growth inhibition and dormancy in *Xanthium* seed. *Physiologia Plantarum* **10**, 266–280.
- Wilken DH** (1982) The balance between chasmogamy and cleistogamy in *Collomia grandiflora* (Polemoniaceae). *American Journal of Botany* **69**, 1326–1333.
- Winn AA** (1991) Proximate and ultimate sources of within-individual variation in seed mass in *Prunella vulgaris* (Lamiaceae). *American Journal of Botany* **78**, 838–844.
- Wolf LL, Hainsworth FR, Mercier T and Benjamin R** (1986) Seed size variation and pollinator uncertainty in *Ipomopsis aggregata* (Polemoniaceae). *Journal of Ecology* **74**, 361–371.
- Wulff RD** (1986) Seed size variation in *Desmodium paniculatum*. I. Factors affecting seed size. *Journal of Ecology* **74**, 87–97.
- Wurzburger J and Leshem Y** (1967) Gibberellin and hull controlled inhibition of germination in *Aegilops kotschy* Boiss. *Israel Journal of Botany* **16**, 181–186.
- Wurzburger J and Koller D** (1976) Differential effects of the parental photo-thermal environment on development of dormancy in caryopses of *Aegilops kotschy*. *Journal of Experimental Botany* **27**, 43–48.
- Wurzburger J, Leshem Y and Koller D** (1974) The role of gibberellin and the hulls in the control of germination in *Aegilops kotschy* caryopses. *Canadian Journal of Botany* **52**, 1597–1601.
- Yang F, Baskin JM, Baskin CC, Yang X, Cao D and Huang Z** (2015a) Effects of germination time on seed morph ratio in a seed-dimorphic species and possible ecological significance. *Annals of Botany* **115**, 137–145.
- Yang F, Yang X, Baskin JM, Baskin CC, Cao D and Huang Z** (2015b) Transgenerational plasticity provides diversity for a seed heteromorphic species in response to environmental heterogeneity. *Perspectives in Plant Ecology, Evolution and Systematics* **17**, 201–208.
- Zeide B** (1978) Reproductive behavior of plants in time. *The American Naturalist* **112**, 636–639.
- Zhang J** (1994) Early seedling development in relation to seed mass and morph in *Cakile edentula*. *Canadian Journal of Botany* **72**, 402–406.
- Zhang K, Baskin JM, Baskin CC, Yang X and Huang Z** (2017) Effect of seed morph and light level on growth and reproduction of the amphicarpic plant *Amphicarpha edgeworthii* (Fabaceae). *Scientific Reports* **7**, 39886.
- Zhang K, Baskin JM, Baskin CC, Cheplick G, Yang X and Huang Z** (2020) Amphicarpic plants: definition, ecology, geographic distribution, systematics, life history, evolution and use in agriculture. *Biological Reviews* **95**, 1442–1466.
- Zohary M** (1937) Die verbreitungsökologischen Verhältnisse der Pflanzen Palästinas. I. Die antitelochorischen Erscheinungen. *Beihefte zum Botanischen Zentralblatt* **56**, 1–55.
- Zohary M** (1962) *Plant life of Palestine: Israel and Jordan*. New York, Ronald Press.
- Zohary M and Fahh A** (1950) On the heterocarpy of *Aethionema*. *Palestine Journal of Botany* **5**, 28–31.
- Zohary M and Imber D** (1963) Genetic dimorphism in fruit types in *Aegilops speltoides*. *Heredity* **18**, 223–231.

Appendix A Genetic diaspore polymorphism in angiosperms

Genetic diaspore polymorphism is best known in the family Valerianaceae, including *Plectritis* (Ganders et al., 1977a,b; Carey and Ganders, 1980) and *Valerianella* (Eggers Ware, 1983). In *Plectritis congesta*, experimental crosses showed that the kind of fruit morph is monogenically inherited with the allele for winged fruits dominant over wingless fruits (Ganders et al., 1977a). In *Plectritis brachystemon*, homozygous winged plants (i.e. selfed plants produced only winged fruits) × homozygous wingless fruits (i.e. selfed plants produced only wingless fruits) → F₁ hybrids, all of which produced winged fruits. The F₂ (F₁ selfed) consisted of 27 plants that produced winged fruits and nine that

produced wingless fruits (a 3:1 ratio). Thus, fruit dimorphism in *P. brachystemon* is controlled by a single locus with the allele for winged fruit dominant (Ganders et al., 1977b). In *Valerianella ozarkana*, crosses between forma *ozarkana* (winged) and forma *bushii* (fusiform) indicated that there was a simple monogenic relationship in which the *ozarkana* (winged) allele is dominant over the *bushii* (fusiform) allele (Eggers-Ware, 1983). It should be pointed out that not all species that produce two or more kinds of diaspores fit either somatic polymorphism or genetic polymorphism, i.e. they neither fit one nor the other. *Fedia cornucopiae* and *F. grandiflora* (Valerianaceae) have been shown to exhibit both somatic and genetic polymorphism (Mathez and Xena de Enrich, 1985a, 1985b).

Genetic diaspore heteromorphism (only one morph per plant) has been reported in the two grass species *Aegilops speltoides* (e.g. Zohary and Imber, 1963; Belyayev and Raskina, 2013; Ruban and Badaeva, 2018) and *Achnatherum hymenoides* (Jones and Nielson, 1999; Jones et al., 2007). In one of the two morphotypes of *A. speltoides* (i.e. form *aucheri*), the spike is the dispersal unit, and in the other morphotype (form *ligustica*) the spikelet is the dispersal unit. In *A. hymenoides*, seed size, i.e. jumbo > globose > elongate) is under genetic control, and degree of seed dormancy decreases from jumbo to globose to elongate.

A kind of genetic polymorphism for capitulum type occurs in British populations of *Senecio vulgaris* (Asteraceae). Plants of this species have radiate and non-radiate capitulum morphs, which are under genetic control. The radiate morph originated via introgressive hybridization between the native non-radiate allotetraploid *S. vulgaris* and the introduced diploid radiate *S. squalidus*. Two tightly-linked genes of *S. squalidus* were introgressed into *S. vulgaris*. A short-rayed form (a heterozygote) is produced from crosses between radiate and non-radiate forms of *S. vulgaris*. Fresh cypselae from

radiate and non-radiate morphs differ in germination phenology. There also were differences in germination percentages after cypselae from radiate and non-radiate morphs were stored in the laboratory from October to June, during which time after-ripening may have occurred (e.g. Trow, 1912; Richards, 1975; Ingram et al., 1980; Abbott, 1986, 1992; Abbott et al., 1988, 1992, 1998; Abbott and Horrill, 1991; Chapman and Abbott, 2010).

Spergula and *Spergularia* are two genera in the Caryophyllaceae containing species that produce heteromorphic seeds with a genetic component. *Spergula arvensis* produces papillate (P) and smooth, i.e. non-papillate (NP), seeds and a hybrid (P × NP) intermediate between the two morphs; the hybrid is produced in a low frequency. Inheritance of seed coat character is controlled by one gene, one locus and two alleles. The intermediate morph is heterozygous with incomplete dominance (New, 1958, 1959, 1961, 1978; New and Herriott, 1981; Wagner, 1986, 1988; Kucwicz and Gojlo, 2013).

Several species of *Spergularia* (e.g. *S. marina* and *S. media*) produce winged and non-winged seeds and a heterozygous intermediate with a narrow wing produced in a low frequency (Salisbury, 1958; Sterk, 1969a,b,c,d; Sterk and Dijkhuizen, 1972; Telenius and Torstensson, 1989, 1991; Telenius, 1992; Ceynowa-Gieldon, 1993; Redbo-Torstensson, 1994; Redbo-Torstensso and Telenius, 1995; 1999; Mazer and Lowrey, 2003; Memon et al., 2010). Seeds of *Spergularia diandra* collected near Sede Boker in the Negev Desert of Israel consisted of three genotypes (hairy, partly-hairy and smooth). Each genotype had three seed-color phenotypes (black, brown and yellow), and there were differences in germination of the phenotypes. Thus, there were nine seed morphs, i.e. 3 phenotypes × 3 genotypes = 9 types of seed morphs (Gutterman, 1994a, 1996, 1997a, b, 1998, 2000).