

## Research Article

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# Pea crabs (*Pinnotheres pisum*) colonisation of five commercial bivalve species from southern Portugal

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## Abstract

This study reports the spatial and depth distributions, occurrence and prevalence, infestation rate and intensity of the pea crab *Pinnotheres pisum* colonising five commercial bivalve species (*Chamelea gallina*, *Donax semistriatus*, *Donax trunculus*, *Donax vittatus*, and *Spisula solida*) along the south and southwest coasts of Portugal. In addition, the study also analysed the colonisation pattern, morphometric measurements and relative growth of *P. pisum* inhabiting those bivalve species. Overall, 33,370 bivalves were analysed, collected in 371 sampling stations at depths ranging from 3 to 25 m. A total of 102 bivalves hosted 106 *P. pisum*, corresponding to an infestation rate of 0.31%. Besides 13 juveniles, were recorded 60 males and 33 females of *P. pisum*, corresponding to a male-biased sex ratio (1M: 0.6F). Pea crabs carapace width ranged from 1.3 to 13.5 mm and males were smaller than females (hard females were also smaller than ovigerous soft females). In general, larger and heavier bivalves hosted larger and heavier *P. pisum*. Pea crabs morphometric relationships presented negative allometries, reflecting slower growth rates in carapace length and total weight compared to carapace width throughout the species ontogeny. Overall, this study provides valuable insights into diverse descriptors of bivalves' colonisation by *P. pisum*, comparing the main results and trends with analogous information available throughout the species distributional range. Although the current fairly low infestation by *P. pisum* does not constitute a health concern for these bivalve species, its evolution under a climate change scenario should be periodically monitored in the mid- and long-term.

## Introduction

Pea crabs are small endosymbiotic crustaceans that colonise diverse marine invertebrates, being commonly found inside the mantle cavities of bivalves hosts such as cockles, clams, and mussels (e.g. Becker and Türkay, 2010, 2017; Drake *et al.*, 2014; Perez-Miguel *et al.*, 2019a; Cuesta *et al.*, 2020; Santos *et al.*, 2022). In European waters (Atlantic Ocean and Mediterranean Sea), five pea crabs species have been recorded, namely: *Afropinnotheres monodi* RB Manning, 1993; *Nepinnotheres pinnotheres* (Linnaeus, 1758); *Pinnotheres bicristatus* García Raso & Cuesta in Cuesta, García Raso, Abelló, Marco-Herrero, Silva & Drake 2019; *Pinnotheres pectunculi* Hesse, 1872; and *Pinnotheres pisum* (Linnaeus, 1767) (Becker, 2010; Becker and Türkay, 2010, 2017; Subida *et al.*, 2011; Marco-Herrero *et al.*, 2017, 2020; Cuesta *et al.*, 2019; Perez-Miguel *et al.*, 2019a). Among these pea crabs, *P. pisum* is the pinnotherid species with the widest latitudinal and longitudinal distribution (Perez-Miguel *et al.*, 2019a), being recorded along the Atlantic Ocean from the North Sea to the Gulf of Cádiz and the Canary Islands, and in the Mediterranean Sea from the Alboran Sea to the Marmara Sea (d'Udeken d'Acoz, 1999; Becker, 2010; Becker and Türkay, 2017; Triay-Portella *et al.*, 2018; González-Gordillo and Cuesta, 2020).

Throughout its distributional range, *P. pisum* inhabits the mantle cavity of diverse bivalve species, including non-commercial bivalves (*Acanthocardia echinata*, *Atrina pectinata*, *Clausinella fasciata*, *Dosinia lupinus*, *Gari fervensis*, *Mactra stultorum*, and *Pinna nobilis*) and commercial bivalves exploited by professional fisheries and/or recreational harvesting activities (*Cerastoderma edule*, *Cerastoderma glaucum*, *Chamelea gallina*, *Donax trunculus*, *Donax variegatus*, *Donax venustus*, *Donax vittatus*, *Ensis ensis*, *Ensis magnus*, *Modiolus modiolus*, *Mytilus edulis*, *Mytilus galloprovincialis*, *Ostrea edulis*, *Ruditapes decussatus*, *Spisula solida*, and *Venus verrucosa*) (e.g. Haines *et al.*, 1994; Delongueville and Scaillet, 2002; Becker, 2010; Becker and Türkay, 2010, 2017; Perez-Miguel *et al.*, 2019a; Cuesta *et al.*, 2020; de Gier and Becker, 2020; González-Gordillo and Cuesta, 2020). Just like other pea crabs (Becker and Türkay, 2010), *P. pisum* usually has a facultative free-living stage in both sexes (males and hard females), whereas reproductive females (soft females) have a last obligate symbiotic stage inside their bivalve hosts that allows them reaching larger sizes (Becker, 2010; Perez-Miguel *et al.*, 2018; González-Ortégón *et al.*, 2021).

Bivalves play a key role in marine ecosystems and constitute important shellfish resources for coastal communities worldwide (e.g. Newell, 2004), with several bivalve species being



targeted by commercial fisheries and recreational harvesting activities (e.g. Gaspar *et al.*, 2012). In Portugal, small-scale bivalve fisheries are traditional and relevant activities at cultural, social, and economic levels, due to fleet size, number of fishermen, and both volume and value of the catches (Gaspar *et al.*, 2002a; Oliveira *et al.*, 2013; Rufino *et al.*, 2017; Almeida *et al.*, 2021). In the last four decades, the Portuguese Institute for the Sea and Atmosphere (IPMA) performs annually bivalve dredge fishing surveys to assess the population status of the most important commercial bivalve species, including *C. gallina*, *Donax semistriatus*, *D. trunculus*, *D. vittatus*, and *S. solida* (with variable relevance depending on the fishing areas along mainland Portugal). These sampling campaigns cover an extensive coastal area and comprise hundreds of sampling stations within a substantial depth range. Besides gathering essential data for stock assessment and fishery management, these fishing surveys also provide an excellent opportunity to collect numerous samples of several species for diverse purposes (e.g. Gaspar *et al.*, 2002b; Cores *et al.*, 2017; Rufino *et al.*, 2018; Vasconcelos *et al.*, 2018).

Due to pinnotherids life strategy and colonisation of bivalve hosts, depending on the infestation degree, pea crabs might induce negative impacts on artisanal fisheries, recreational harvesting and aquaculture activities (e.g. Perez-Miguel *et al.*, 2018; Cuesta *et al.*, 2020; Marco-Herrero *et al.*, 2020; Santos *et al.*, 2022). Indeed, pea crabs can injure bivalve gills and affect filtering efficiency, leading to slower growth, maturation and condition index of their bivalve hosts (Christensen and McDermott, 1958; Sun *et al.*, 2006; Mena *et al.*, 2014; Yasuoka and Yusa, 2017; Cuesta *et al.*, 2020). In addition, pea crab infestation of edible bivalve species can prompt consumer complaint and rejection, which might constitute a concern for their commercial exploitation (Trottier *et al.*, 2012; Hutson and Cain, 2019). In this context, the present study reports the spatial and depth distribution, occurrence and prevalence, infestation rate and intensity of the pea crab *P. pisum* colonising *C. gallina*, *D. semistriatus*, *D. trunculus*, *D. vittatus*, and *S. solida* along the south and southwest coasts of Portugal.

## Materials and methods

### Fishing surveys

Fishing surveys were performed on-board IPMA's research vessel 'RV Diplodus' in two fishing areas (south and southwest coasts) along mainland Portugal (Figure 1). In the south coast, surveys were conducted from the 6<sup>th</sup> to the 16<sup>th</sup> of July 2017 and covered all bivalve fishing grounds between Vila Real de Santo António and Olhos d'Água (37°9'40"N, 7°23'55"W to 37°4'59"N, 8°11'13"W) (Figure 1). The coast was subdivided into transects perpendicular to the shoreline (separated ½ nautical mile between each other) and were surveyed 266 sampling stations ranging between 3 and 15 m depth. In the southwest coast, surveys were performed from the 22<sup>nd</sup> to the 31<sup>st</sup> July 2017 and covered two main zones between Costa da Caparica and Sines (38°39'00"N, 09°15'29"W to 37°59'00"N, 08°51'02"W) (Figure 1). The coast was also subdivided into transects perpendicular to the shoreline (distanced 1 nautical mile apart) and were surveyed 187 sampling stations ranging between 3 and 25 m depth. Overall, the two fishing surveys required 17 days at sea (south: 9 days; southwest: 8 days).

Samples were collected with mechanical bivalve dredges similar to those used in commercial fisheries, towed at a constant speed of 1.5 knots for 5 min. All bivalve samples were kept in identified plastic bags, which were preserved in ice until being transported to the laboratory for species sorting and biological sampling.

### Laboratory sampling

In the laboratory, bivalve species were sorted, identified and separated into commercial and bycatch species. Whenever available, were sampled 100 individuals of the main commercial bivalve species (*C. gallina*, *D. semistriatus*, *D. trunculus*, *D. vittatus*, and *S. solida*) per sampling station (otherwise if less than 100, all individuals of those species were sampled). Subsequently, shells of commercial bivalve species were opened using a scalpel and their mantle cavities were carefully examined for the existence of pea crabs, which were identified to the species level based on specialised literature (Becker and Türkay, 2010; Cuesta *et al.*, 2019; Marco-Herrero *et al.*, 2020).

Commercial bivalves (with and without pinnotherids) were counted and measured for shell length (SL, maximum distance along the anterior–posterior axis) using a digital caliper (precision = 0.01 mm) and weighed for total weight (TW) on a top loading balance (precision = 0.01 g). Pea crabs were measured for carapace width (CW, maximum distance along the cephalothorax) and carapace length (CL, maximum distance across the cephalothorax) using a digital caliper (precision = 0.01 mm) and weighed for total weight (TW) on a high precision balance (precision = 0.0001 g).

Pea crabs multiple infestations (more than one pinnotherid per bivalve host) were also quantified (e.g. Seed, 1969; Haines *et al.*, 1994; Perez-Miguel *et al.*, 2018; Cuesta *et al.*, 2020; Santos *et al.*, 2022). Smaller and sexually undifferentiated pinnotherids were classified as juveniles. Pea crabs were sexed depending on their external morphological features: males (with gonopods) or females (with pleopods). In addition, depending on their abdomen characteristics, developmental and maturity stages, females were further classified as hard or soft females (non-ovigerous and ovigerous). Illustrative photographs of *P. pisum* juveniles, males and females (hard, soft non-ovigerous and soft ovigerous females) are presented in Figure 2.

### Data treatment and statistical analyses

Bivalve's colonisation by pinnotherids was assessed through the infestation rate (IR – proportion of bivalves hosting pea crabs, either single or multiple occurrences), prevalence rate (PR – proportion of pea crabs within all sampled bivalves) and infestation intensity (II – proportion of pea crabs single and multiple occurrences). In the absence of multiple infestations (i.e. more than one pea crab inside a single bivalve host), the IR is equivalent to the PR.

Pea crabs sex ratio, expressed as the proportion of females per male (females including hard and soft, non-ovigerous and ovigerous), was compared with parity (1M: 1F) using the chi-square ( $\chi^2$ ) test. Pea crabs size (CW) was compared between locations (south and southwest coasts), sexes and female maturity stages through analysis of variance (ANOVA). Whenever ANOVA assumptions (data normality and homogeneity of variances) were not met, analyses were performed using the non-parametric Kruskal–Wallis test (K–W), followed by pairwise multiple comparisons to detect significant differences between groups.

In order to further analyse the relationships among diverse descriptors of pea crabs colonisation of commercial bivalve species (IR, PR, SL, CW, sex ratio, and depth) and detect eventual differences between fishing areas (south and southwest coasts), a principal component analysis (PCA) based on Pearson correlations was performed to improve the visualisation and interpretation of the dataset.

Relationships between bivalve host size (SL) and weight (TW), and pea crab size (CW) and weight (TW) were analysed through regression analysis, by fitting the linear function ( $Y = a + bX$ ) to

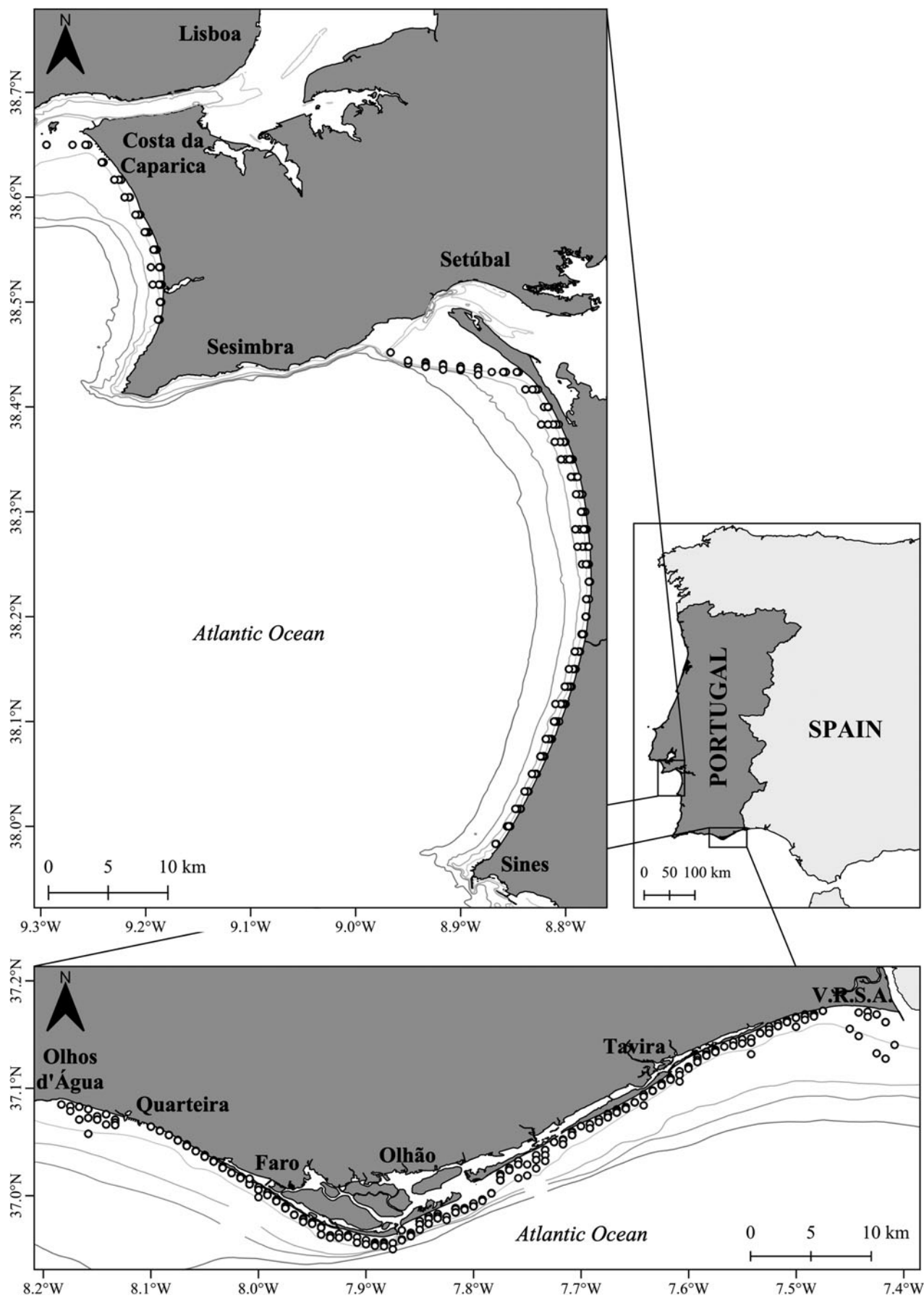
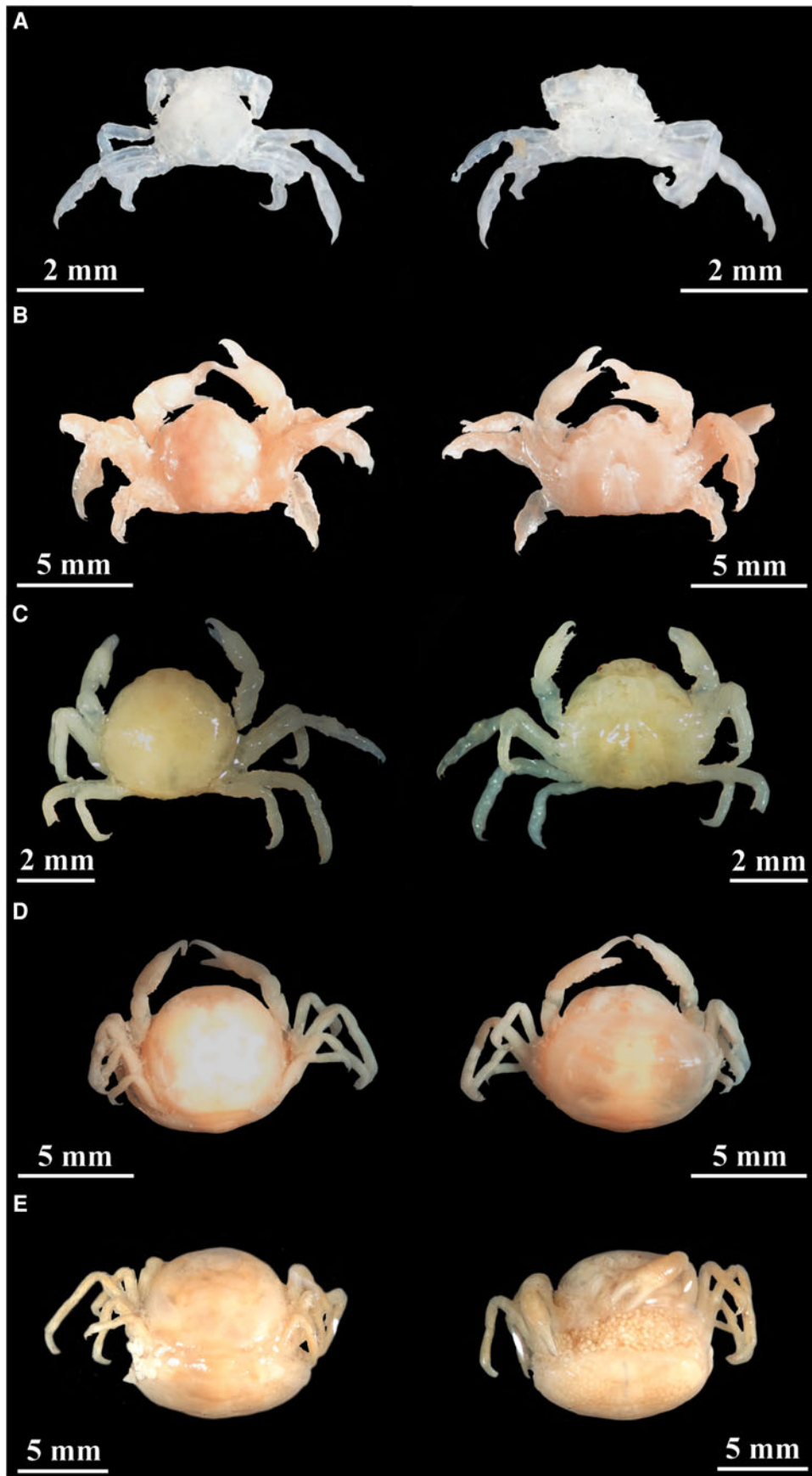


Figure 1. Map showing the sampling stations for collecting bivalve species during the fishing surveys with mechanical dredges along the south and southwest coasts of Portugal.



**Figure 2.** Illustrative photographs (dorsal and ventral views) of *Pinnotheres pisum*: (A) juvenile; (B) male; (C) hard female; (D) soft non-ovigerous female; (E) soft ovigerous female.

raw data and assessing the degree of association between measured morphometric variables with the correlation coefficient ( $r$ ). In addition, aiming to examine the relative growth of pea crabs

cephalothorax size and total weight during ontogeny, morphometric relationships were established through regression analysis between CW, CL and TW in juveniles, males and females

(hard and soft females), by fitting the power function ( $Y = aX^b$ ) to raw data and assessing relative growth (isometry vs allometry) through the allometry coefficient (regression slope –  $b$ ).

In relationships between linear variables (CW and CL), isometry growth occurs when  $b$  is not significantly different from 1, whereas in relationships between linear and ponderal variables (CW and TW) isometry occurs when  $b$  is not significantly different from 3, both reflecting similar growth rates between variables throughout ontogeny (Huxley and Teissier, 1936; Mayrat, 1970). Accordingly, a  $t$ -test ( $H_0$ ,  $b = 1$  or 3;  $H_A$ ,  $b \neq 1$  or 3) was performed to confirm the isometric ( $b = 1$  or 3) or allometric (negative allometry,  $b < 1$  or 3; positive allometry,  $b > 1$  or 3) relative growth between variables. All statistical analyses were performed following Sokal and Rohlf (1987) and Zar (1996), with statistical significance level set for  $P < 0.05$ .

## Results

### Commercial bivalves

Commercial bivalve species included in this study (*C. gallina*, *D. semistriatus*, *D. trunculus*, *D. vittatus*, and *S. solida*) were collected in 371 sampling stations (south: 207 stations; southwest: 164 stations) at variable fishing depths ranging from 3 to 25 m depth (south:  $6.2 \pm 2.8$  m; southwest:  $10.1 \pm 5.3$  m) (Table 1). Overall, 33,370 individuals of commercial bivalves were analysed (south: 25,348 inds.; southwest: 8,022 inds.) belonging to the following species: *S. solida* (south: 8,892 inds.; southwest: 4,666 inds.), *D. trunculus* (south: 6,990 inds.; southwest: 1,706 inds.), *C. gallina* (south: 8,360 inds.; southwest: 132 inds.), *D. vittatus* (south: 47 inds.; southwest: 1,518 inds.), and *D. semistriatus* (south: 1,059 inds.) (Table 1).

### Pea crabs

A total of 102 commercial bivalves hosted pea crabs, all belonging to the species *P. pisum* (south: 21 inds.; southwest: 81 inds.) (Table 1 and Figure 3). Most pea crabs colonised *S. solida* (south: 11 inds.; southwest: 67 inds.), distantly followed by *D. vittatus* (southwest: 11 inds.), *D. trunculus* (south: 5 inds.; southwest: 1 ind.), *C. gallina* (south: 3 inds.; southwest: 2 inds.), and *D. semistriatus* (south: 2 inds.) (Table 1 and Figure 3). Bivalve hosts comprised specimens with broad size and weight, ranging from 17.1 to 41.2 mm SL in *C. gallina* and from 1.1 g in *D. vittatus* to 16.5 g in *S. solida*. Overall, 106 *P. pisum* pea crabs (south: 23 inds.; southwest: 83 inds.) were detected inside bivalve shells, the vast majority as single occurrences (one pea crab per bivalve host) complemented by very few multiple occurrences (two pea crabs, one male and one female, per bivalve host) (Table 1).

Pea crabs size and weight ranged from 1.3 to 13.5 mm CW and from 0.001 to 0.350 g TW. Mean carapace width (K–W:  $H = 0.194$ ;  $P > 0.05$ ) did not display significant differences between the south coast ( $4.3 \pm 2.1$  mm CW) and the southwest coast ( $4.9 \pm 3.1$  mm CW) (Table 1). Besides 13 juveniles of *P. pisum* (undistinguishable sex), 60 males and 33 females were recorded, corresponding to a highly unbalanced ( $\chi^2 = 7.269$ ,  $P < 0.01$ ) and male-biased sex ratio (1M: 0.6F). Among females, were identified 5 hard and 28 soft females, of which 3 non-ovigerous and 25 ovigerous females (Table 1). On average, males ( $3.5 \pm 0.6$  mm CW) were significantly smaller (K–W:  $H = 57.222$ ,  $P < 0.001$ ) than females ( $8.3 \pm 2.8$  mm CW). In addition, mean carapace width was also significantly different among female maturity stages (ANOVA:  $F = 13.290$ ;  $P < 0.001$ ), with hard females ( $4.3 \pm 0.6$  mm CW) being significantly smaller (Tukey test:  $q = 6.957$ ;  $P < 0.001$ ) than ovigerous soft females ( $9.4 \pm 2.3$  mm CW).

### Pea crabs colonisation pattern

The colonisation pattern of commercial bivalves by *P. pisum* in terms of fishing depth and bivalve host size is presented in Figure 4. The number and relative frequency of pea crabs displayed a clear decreasing trend with increasing fishing depth ( $r = 0.974$ ;  $P < 0.05$ ), ranging from 39.6% at shallower depths (<10 m) to 14.2% in deeper sampling stations ( $\geq 20$  m) (Figure 4A). Such differences in the occurrence and proportion of pea crabs at each depth interval were mainly due to the sampling effort and number of commercial bivalves caught at each depth interval, since the vast majority ( $N = 28,712$ ; 86.0%) was collected at shallower depths (< 10 m) and only a minor fraction ( $N = 967$ ; 2.9%) was collected at greater depths ( $\geq 20$  m). The shallowest and deepest occurrences of *P. pisum* were recorded in *D. trunculus* at 3 m depth in the south coast and in *S. solida* at 25 m depth in the southwest coast, respectively. Pea crabs depth distribution was strongly influenced by the highly predominant bivalve host *S. solida* (comprising 81 *P. pisum*) mainly distributed at deeper bathymetrics ( $13.0 \pm 4.9$  m; range = 4.8–25.0 m). On the opposite, pea crabs colonising the hosts *C. gallina* (containing 6 *P. pisum*) and *Donax* spp. (hosting 19 *P. pisum*) occurred predominantly at intermediate ( $8.4 \pm 3.5$  m; range = 4.8–15.0 m) and shallower depths ( $6.9 \pm 3.1$  m; range = 3.0–12.0 m).

Although host bivalve species ranged from 17.1 to 41.2 mm SL, the vast majority of the pea crabs colonised intermediate-sized bivalve hosts belonging to the sizes classes 20–30 mm SL (52.8%) and 30–40 mm SL (41.5%), with only minor occurrence in smaller (<20 mm SL = 3.8%) and larger bivalves ( $\geq 40$  mm SL = 1.9%) that were also scarcer in the overall samples (Figure 4B). Overall, host bivalve species displayed the following decreasing trend in mean shell length: *S. solida* ( $30.2 \pm 6.4$  mm SL), *C. gallina* ( $28.9 \pm 5.9$  mm SL) and *D. semistriatus*, *D. trunculus*, and *D. vittatus* ( $27.4 \pm 4.6$  mm SL). In addition, while smaller *P. pisum* males clearly prevailed in 20–30 mm SL bivalve hosts (38 males and 8 females), larger *P. pisum* females predominated in 30–40 mm SL bivalve hosts (23 females and 18 males).

### Pea crabs infestation rate

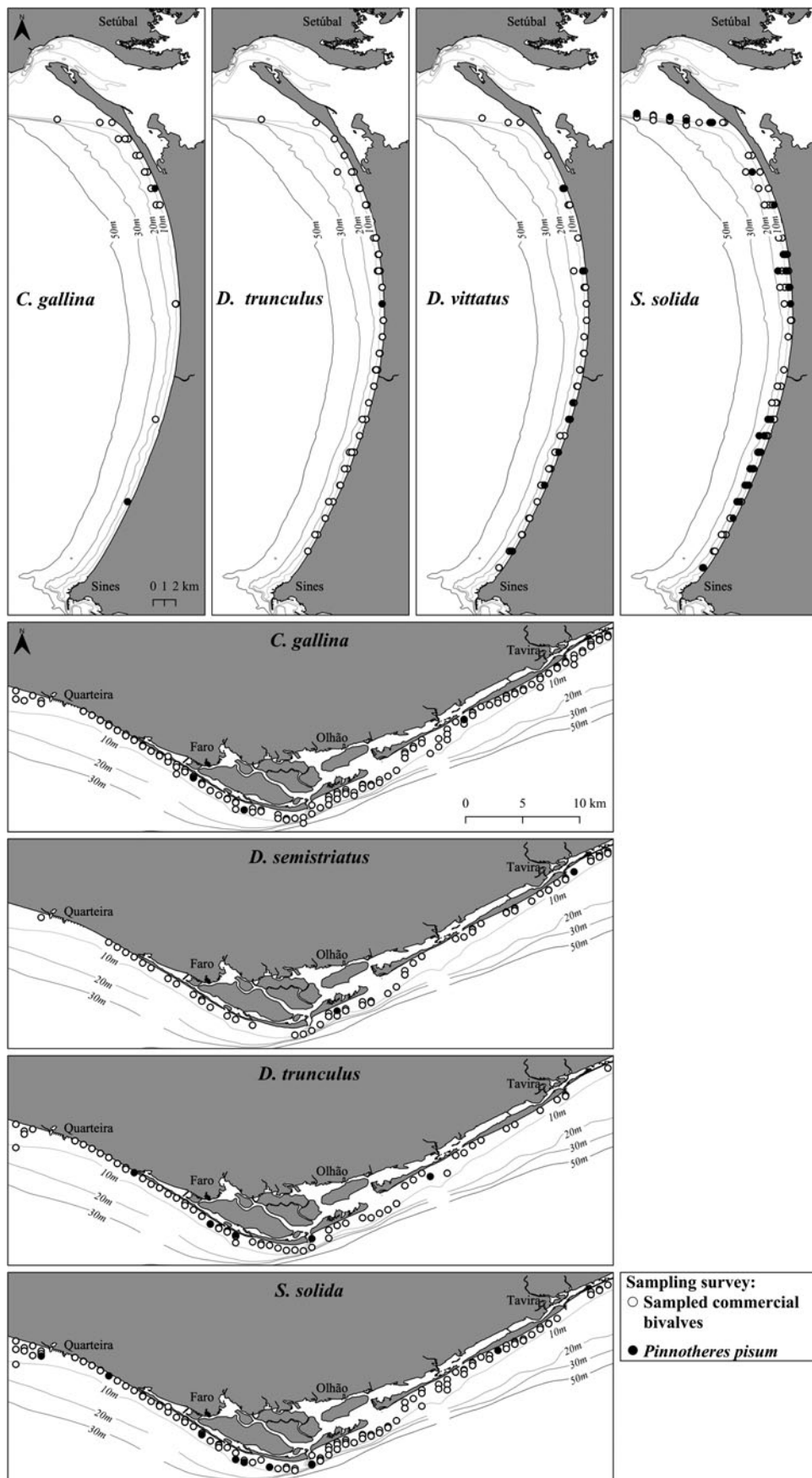
Overall, the 102 bivalves hosting 106 *P. pisum* corresponded to an infestation rate of 0.31% (prevalence rate of 0.32% due to the multiple infestation of four bivalves colonised by two pea crabs). Although invariably low, the infestation rate was clearly higher in the southwest coast (IR = 1.01%) than in the south coast (IR = 0.08%) (Table 1). The spatial and depth variation in the infestation rate of the five commercial bivalve species by pea crabs along the south and southwest coasts of Portugal is showed in Figure 5. In the south coast, the infestation rate ranged from 0% in *D. vittatus* to 0.19% in *D. semistriatus*. In the southwest coast, the lowest value occurred in *D. trunculus* (IR = 0.06%), with highest infestation rates recorded in *C. gallina* (IR = 1.52%) closely followed by *S. solida* (IR = 1.44%) (Figure 5A). Regarding the bathymetric variation, the infestation rate displayed a clear growing trend with increasing fishing depth ( $r = 0.954$ ;  $P < 0.05$ ), which was particularly evident between <10 and  $\geq 10$  m depth, ranging from 0.14% at shallower depths (<10 m) to 1.55% in the deeper sampling stations ( $\geq 20$  m) (Figure 5B).

The PCA provided further insights into the relationships between host bivalves and guest pea crabs, with the two principal axes of the PCA accounting for 84.0% of the total variance in the dataset (PC1 = 58.9%; PC2 = 25.1%) (Figure 6). The PCA depicted a clear relationship between IR and PR by *P. pisum* in the host bivalves *S. solida* and *C. gallina* in the southwest coast, which was strongly related to fishing depth. The opposite trend was

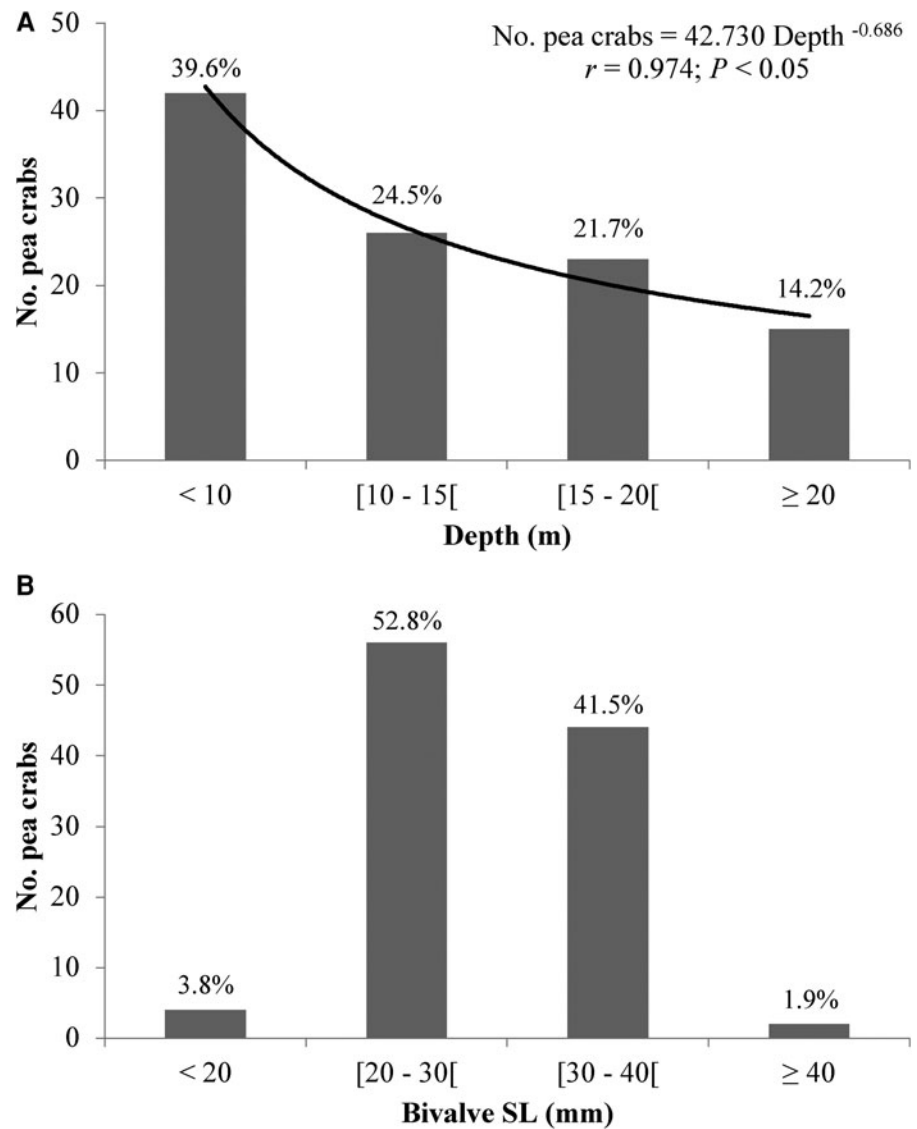
**Table 1.** Descriptive statistics of host commercial bivalves (*Chamelea gallina*, *Donax semistriatus*, *Donax trunculus*, *Donax vittatus*, and *Spisula solida*) and pea crabs (*Pinnotheres pisum*) collected during bivalve dredge fishing surveys along the south and southwest coasts of Portugal

Sampling stations		Bivalve hosts							<i>Pinnotheres pisum</i>												
Location	Ns	Depth (m) (min – max)	Species	Nb	SLs (mm) (min – max)	Ni	SLi (mm)	IR (%)	Np	CW (mm) (min – max)	PR (%)	M	F	M:F	HF	SFnov	SFov	J	M + HF	M + SFov	
South coast (Algarve)	179	3.0–13.8	<i>Chamelea gallina</i>	8,360	8.6–49.5	3	29.6 ± 4.6 (26.5–34.9)	0.04	4	6.2 ± 3.1 (3.3–10.3)	0.05	1	3	1M:3F	1	0	2	0	1	0	
	81	3.0–8.4	<i>Donax semistriatus</i>	1,059	14.1–31.7	2	27.9 ± 0.6 (27.5–28.3)	0.19	2	2.9 ± 0.7 (2.4–3.3)	0.19	2	0	–	0	0	0	0	0	0	
	74	3.0–12.0	<i>Donax trunculus</i>	6,990	11.9–41.7	5	27.5 ± 3.6 (24.7–33.8)	0.07	5	4.5 ± 1.6 (3.6–7.3)	0.07	4	1	1M:0.3F	0	0	1	0	0	0	
	7	3.0–8.4	<i>Donax vittatus</i>	47	21.1–34.4	0	–	0	0	–	0	–	–	–	–	–	–	–	–	–	–
	145	3.0–15.6	<i>Spisula solida</i>	8,892	12.3–42.0	11	22.6 ± 4.6 (17.1–30.8)	0.12	12	3.9 ± 2.0 (1.8–8.4)	0.13	9	2	1M:0.2F	0	0	2	1	0	1	
Total	207	3.0–15.6		25,348	8.6–49.5	21	25.3 ± 4.9 (17.1–34.9)	0.08	23	4.3 ± 2.1 (1.8–10.3)	0.09	16	6	1M:0.4F	1	0	5	1	1	1	
Southwest coast (Setúbal)	23	3.0–19.0	<i>Chamelea gallina</i>	132	14.4–38.9	2	28.6 ± 11.4 (20.6–36.7)	1.52	2	3.4 ± 1.1 (2.6–4.1)	1.52	2	0	–	0	0	0	0	0	0	
	70	3.0–20.0	<i>Donax trunculus</i>	1,706	15.6–46.3	1	39.0	0.06	1	3.5	0.06	1	0	–	0	0	0	0	0	0	
	75	3.0–20.0	<i>Donax vittatus</i>	1,518	15.5–35.6	11	26.2 ± 4.2 (21.4–34.9)	0.72	11	4.2 ± 3.2 (1.3–10.4)	0.72	5	2	1M:0.4F	0	0	2	4	0	0	
	105	3.0–25.0	<i>Spisula solida</i>	4,666	16.4–44.6	67	31.3 ± 5.8 (19.6–41.2)	1.44	69	5.1 ± 3.2 (1.4–13.5)	1.48	36	25	1M:0.7F	4	3	18	8	2	0	
Total	164	3.0–25.0		8,022	14.4–46.3	81	30.6 ± 6.0 (19.6–41.2)	1.01	83	4.9 ± 3.1 (1.3–13.5)	1.03	44	27	1M:0.6F	4	3	20	12	2	0	
TOTAL	371	3.0–25.0		33,370	8.6–49.5	102	29.5 ± 6.1 (17.1–41.2)	0.31	106	4.8 ± 2.9 (1.3–13.5)	0.32	60	33	1M:0.6F	5	3	25	13	3	1	

Ns, number of sampling stations where the bivalve species was collected; Nb, number of sampled bivalves; SLs, shell length of sampled bivalves; Ni, number of infested bivalves; SLi, mean shell length of infested bivalves; IR (%), infestation rate (%); Np, number of pea crabs; CW, mean carapace width of pea crabs; PR (%), prevalence rate (%); M, males; F, females; M:F, sex ratio; HF, hard females; SFnov, non-ovigerous soft females; SFov, ovigerous soft females; J, juveniles; M + HF, bivalve multiple infestation by one male and one hard female pea crab; M + SFov, bivalve multiple infestation by one male and one ovigerous soft female. Size data presented as mean ± SD and/or respective range (minimum and maximum).



**Figure 3.** Spatial and depth distribution of *Pinnotheres pisum* colonising commercial bivalve species (*Chamelea gallina*, *Donax semistriatus*, *Donax trunculus*, *Donax vittatus*, and *Spisula solida*) along the south and southwest coasts of Portugal.



**Figure 4.** Number and frequency of *Pinnotheres pisum* colonising commercial bivalve species as a function of (A) sampling station depth (5 m depth intervals); (B) bivalve host size (10 mm SL size classes).

displayed in the south coast, where pea crabs occurrence was influenced by host bivalves caught at shallower depths. Furthermore, the PCA also highlighted that IR and PR are mostly related to the most frequent host bivalves inhabiting deeper fishing grounds (*S. solida* and *C. gallina*), whereas pea crab descriptors (CW and sex ratio) are mainly determined by host bivalve size (SL) (Figure 6).

#### Host bivalves – guest pea crabs relationships

The relationships established between bivalve hosts size and weight and colonising pea crabs size and weight are illustrated in Figure 7. In general, larger bivalves hosted larger *P. pisum*, as revealed by the highly significant correlation ( $r = 0.403$ ;  $P < 0.001$ ) and positive slope ( $b = 0.194$ ) between bivalve shell length and pea crab carapace width (Figure 7A). Accordingly, following the same general trend, heavier bivalves also hosted heavier *P. pisum*, although with slightly lower correlation ( $r = 0.369$ ;  $P < 0.001$ ) and more gentle positive slope ( $b = 0.007$ ) (Figure 7B).

#### Pea crabs morphometric relationships and relative growth

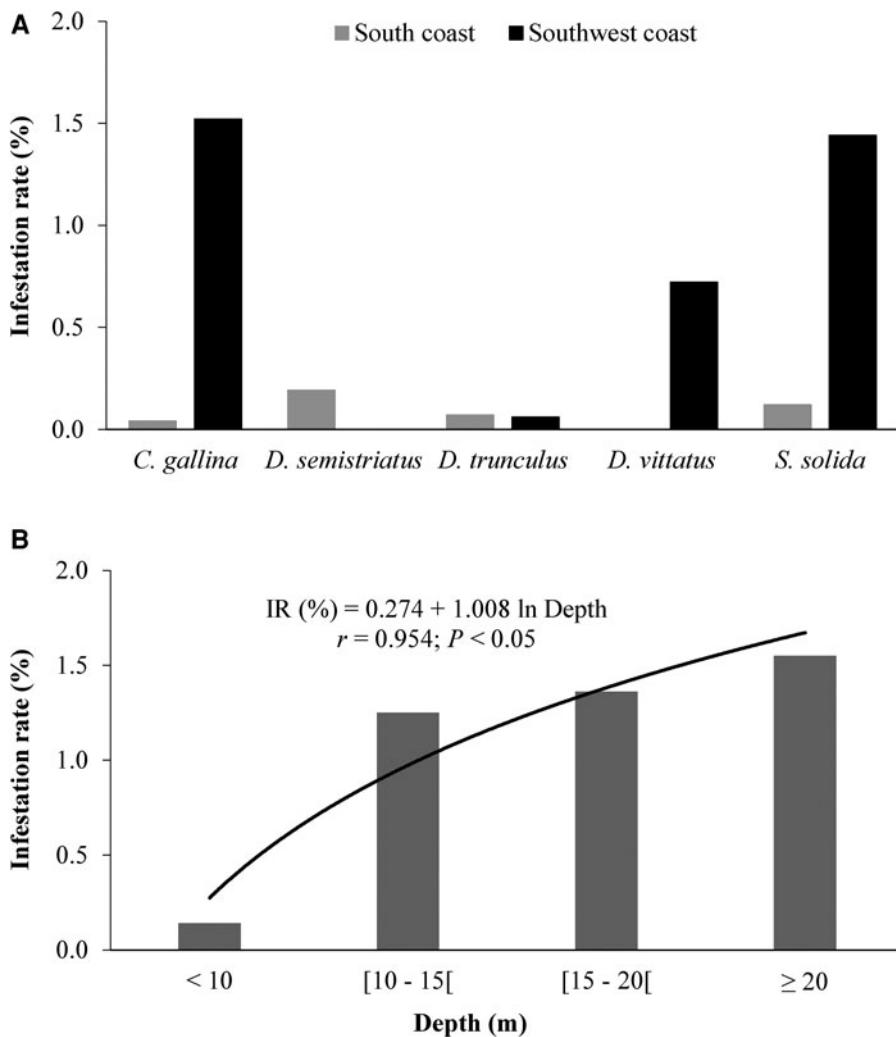
The morphometric relationships established between pea crabs carapace width, carapace length, and total weight are displayed in Figure 8. Both relationships were highly significant ( $P <$

0.001) and characterised by very high correlation coefficients between carapace width and length ( $r = 0.982$ ) (Figure 8A) and between carapace width and total weight ( $r = 0.964$ ) (Figure 8B). In both cases, negative allometric growth was recorded between variables, reflecting slower growth rates in both carapace length ( $b = 0.924$ ) and total weight ( $b = 2.374$ ) compared to carapace width throughout the ontogeny of *P. pisum* (Figures 8A and B).

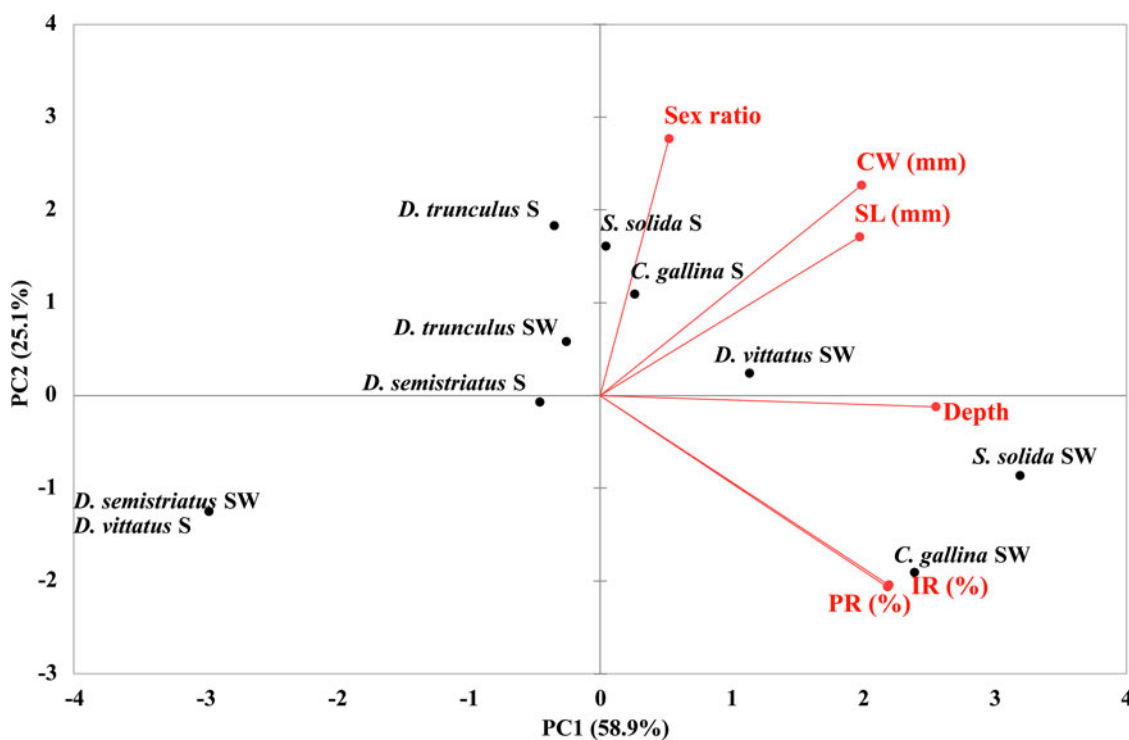
In addition, the relative growth between pea crabs carapace width, carapace length and total weight as function of their sex, developmental and female maturity stages is compiled in Table 2. Overall, only isometric ( $b = 1$  or 3) and negative allometric growth ( $b < 1$  or 3) were recorded between variables. The whole specimens recorded negative allometries in both morphometric relationships (CL vs CW and TW vs CW), revealing a comparatively faster growth rate in carapace width than in carapace length and total weight during growth (i.e. gradual widening of *P. pisum* carapace throughout the species ontogeny) (Table 2).

However, different trends in relative growth were detected depending on *P. pisum* development (juveniles = isometries), sex (males = isometry or negative allometry; females = negative allometries), and female maturity stages (hard females = isometry; soft females = negative allometries) (Table 2). Isometric growth in both morphometric relationships only occurred in juveniles, reflecting a similar growth rate between CW, CL, and TW in the smallest pea crabs. The few other isometries also occurred

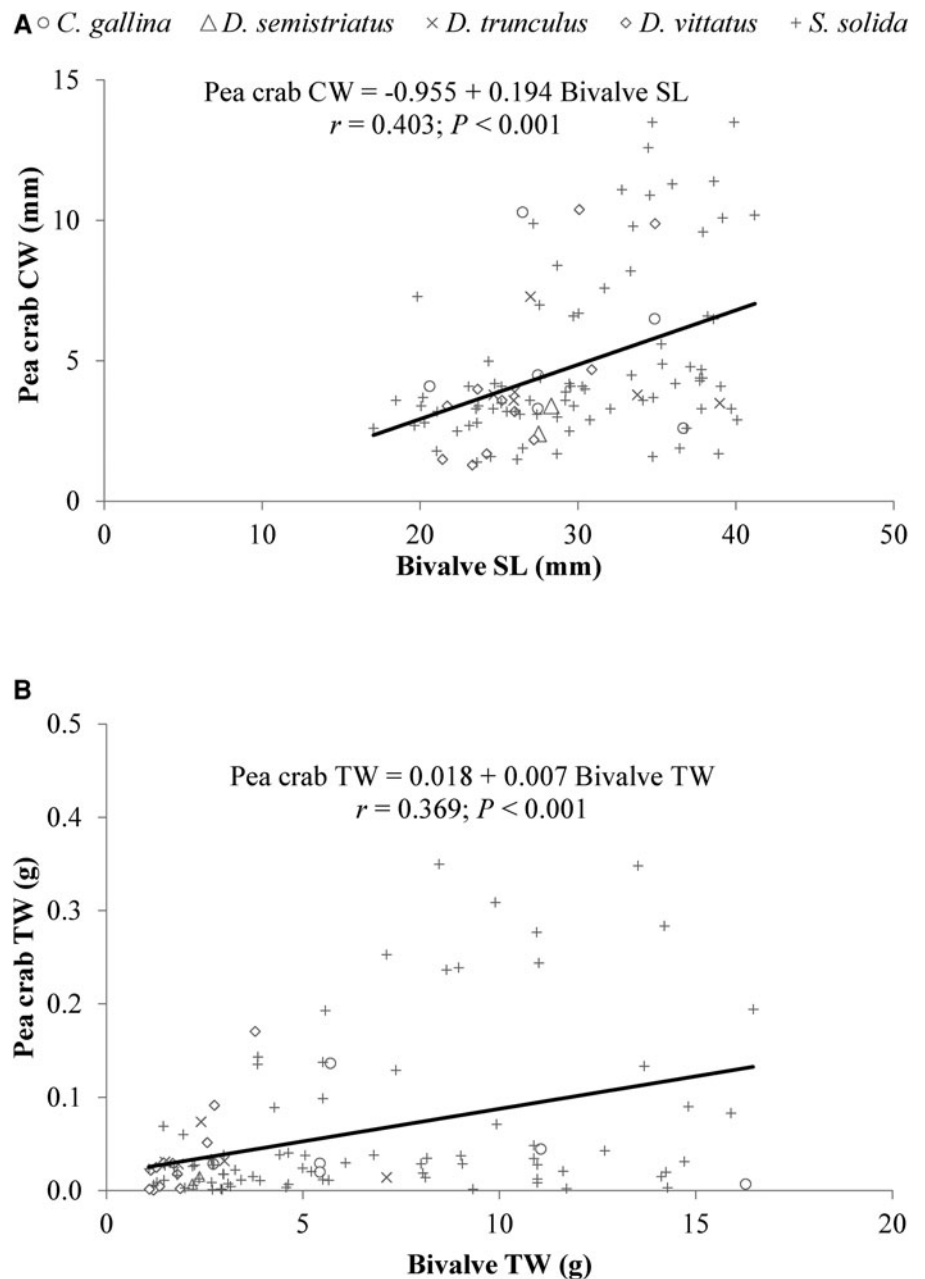




**Figure 5.** Variation in the infestation rate of commercial bivalve species by *Pinnotheres pisum* as a function of (A) sampling station location (south and southwest coasts of Portugal); (B) sampling station depth (5 m depth intervals).



**Figure 6.** Principal component analysis (PCA) biplot showing the relationship among diverse descriptors of *Pinnotheres pisum* colonisation of commercial bivalve species along the south and southwest coasts of Portugal. IR, infestation rate; PR, prevalence rate; SL, shell length; CW, carapace width.



**Figure 7.** Relationships between (A) bivalve host shell length and *Pinnotheres pisum* carapace width; (B) bivalve host total weight and *Pinnotheres pisum* total weight.

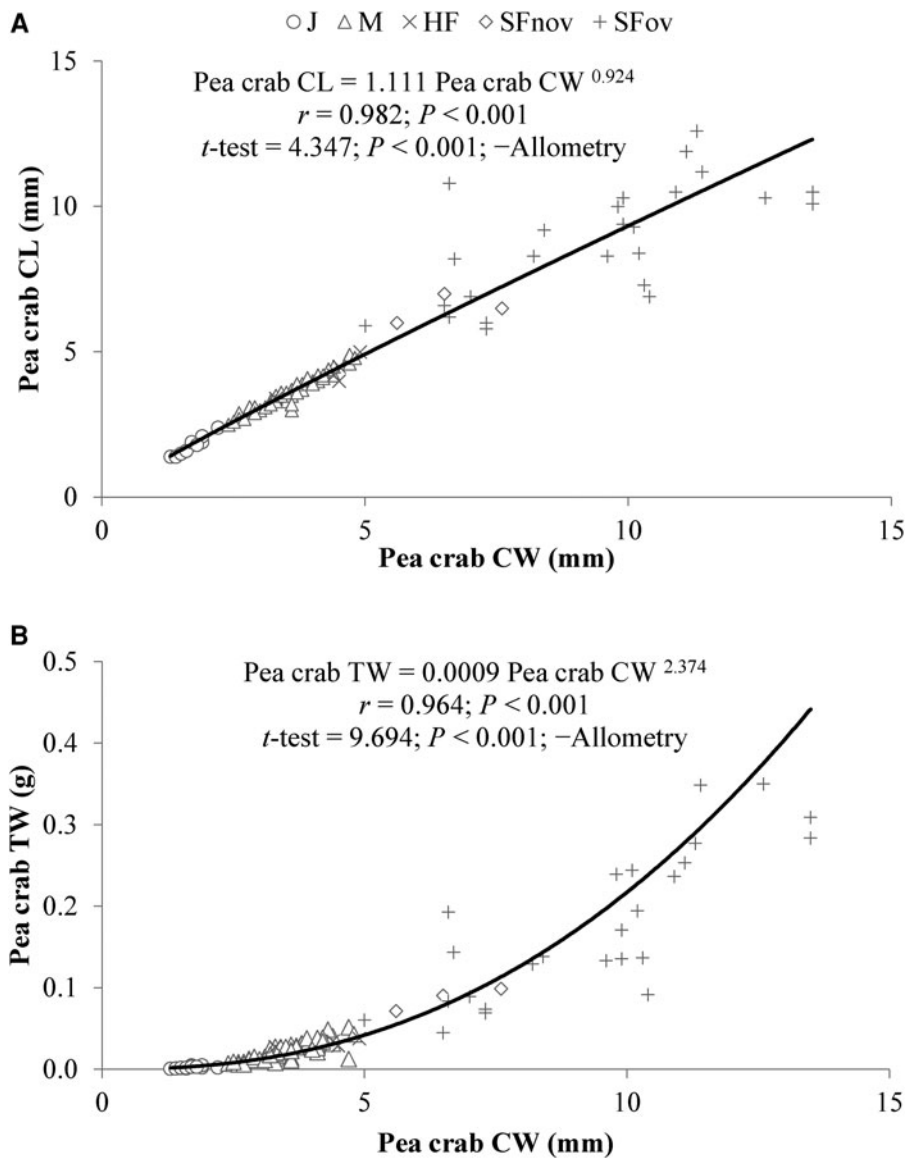
in the tending smaller males (TW vs CW) and hard females (CL vs CW). On the opposite, the clearly larger soft females (and consequently the whole females) exhibited invariably negative allometries in both morphometric relationships, denoting a continuous broadening of the carapace towards more developed and mature females of *P. pisum* (Table 2).

## Discussion

The pea crab *P. pisum* is among the five pinnotherid species previously reported for European waters (Becker, 2010; Becker and Türkay, 2010, 2017; Subida et al., 2011; Marco-Herrero et al., 2017, 2020; Cuesta et al., 2019; Perez-Miguel et al., 2019a). This pea crab has been recorded at several localities along the NE Atlantic Ocean and across the Mediterranean Sea (Table 3), being the European pinnotherid species with broadest latitudinal distribution (Perez-Miguel et al., 2019a) and with widest range of confirmed bivalve host species (Becker and Türkay, 2017; Perez-Miguel et al., 2019a; González-Gordillo and Cuesta, 2020). In the present study, *P. pisum* occurred in *C. gallina*, *D.*

*semistriatus*, *D. trunculus*, *D. vittatus*, and *S. solida*, corroborating previous colonisations of these commercial bivalve species in other geographical areas, such *D. vittatus* and *S. solida* in the North Sea (Becker and Türkay, 2017) and *C. gallina* and *D. trunculus* in the Mediterranean Sea (Perez-Miguel et al., 2019a) (Table 3). On the opposite, at least according to the present authors' best knowledge, this study reports the first record of *P. pisum* colonising the bivalve host *D. semistriatus*.

Recent studies in Iberian waters reported very low infestation rates by the European native *P. pisum* but high infestation rates of the African pea crab *A. monodi* (e.g. Drake et al., 2014; Perez-Miguel et al., 2019a, 2019b; Cuesta et al., 2020; Santos et al., 2022). During its northwards expansion along the Portuguese coast (Subida et al., 2011; Perez-Miguel et al., 2019a, 2019b; Santos et al., 2022), *A. monodi* colonised diverse bivalve species, including the commercially exploited *C. gallina*, *D. trunculus*, and *S. solida*. However, this pea crab was not recorded in the present study, absence that is probably related to inter-specific differences in preferential habitat, with *P. pisum* colonising preferentially subtidal bivalve hosts (Houghton, 1963;



**Figure 8.** Morphometric relationships of *Pinnotheres pisum* and relative growth between (A) carapace width and carapace length; (B) carapace width and total weight. J, juveniles; M, males; HF, hard females; SFnov, non-ovigerous soft females; SFov, ovigerous soft females.

Seed, 1969; Gam *et al.*, 2008; Becker, 2010; Perez-Miguel *et al.*, 2019a) and *A. monodi* preferring intertidal bivalve hosts (Drake *et al.*, 2014; Perez-Miguel, 2018; Perez-Miguel *et al.*, 2019a; Santos *et al.*, 2022).

High infestation rates by *P. pisum* have been recorded in *C. edule* from Spain (Bruzos *et al.*, 2020) and Morocco (Gam *et al.*, 2008), as well as in *M. edulis* from England (Seed, 1969; Haines *et al.*, 1994) and *M. modiolus* from the North Sea (Becker and Türkay, 2017) (Table 3). In the present study, infestation rates were much lower, ranging from 0.04% to 1.5% in *C. gallina* from the south and southwest coasts, respectively, more similar to values reported for *C. gallina* in the Mediterranean Sea (Perez-Miguel *et al.*, 2019a) and for *D. vittatus* and *S. solida* in the North Sea (Becker and Türkay, 2017). Overall, differences in infestation rates by *P. pisum* seem intrinsically related to diverse factors, including the host bivalve species (e.g. higher infestation rates in *C. edule*, *M. edulis*, and *M. modiolus* compared to other bivalve species), prevailing environmental conditions (sheltered vs exposed sites), habitat location and depth (intertidal vs subtidal areas) (e.g. Houghton, 1963; Seed, 1969, 1971; Haines *et al.*, 1994; Becker, 2010; Becker and Türkay, 2017; Perez-Miguel, 2018; Perez-Miguel *et al.*, 2019a; Santos *et al.*, 2022).

Regarding the prevalence rate (i.e. multiple pea crabs colonising a single bivalve), the vast majority of infested bivalves

contained a single *P. pisum* inside the shell (96.1%), with only four host bivalves (three *S. solida* and one *C. gallina*) presenting multiple infestations (3.9%). Higher prevalence rates of *P. pisum* were recorded in *M. edulis* from southwestern England, with 135 single infestations (87.7%), 17 double infestations (11.0%) and 2 triple infestations (1.3%) (Seed, 1969). Likewise, in *M. edulis* from southern England, where most mussels hosted a single pea crab (range: 74.9–88.0%) distantly followed by two (range: 12.0–24.9%) or three (range: 0.1–0.3%) pea crabs inside a host mussel (Haines *et al.*, 1994), and in *M. modiolus* from Helgoland Trench, with 191 single infestations (75.2%) and 63 double infestations (24.8%) (Becker and Türkay, 2017).

Likewise the present study in southern and southwestern Portugal, where multiple infestations comprised only pairs of males and hard females or soft ovigerous females, in southwestern England all double infestations of *M. edulis* involved one male and one female *P. pisum* (Seed, 1969), whereas in southern England the vast majority of double infestations also consisted of pea crabs couples (range: 85.2–90.2%) (Haines *et al.*, 1994). Indeed, although *P. pisum* tends to live alone inside bivalve hosts (just like other pea crab species), double infestations by male and female couples suggest mating encounters instead of casual co-occurrence by pure coincidence. Such recurrent and prevailing single infestations might be due to complementary

**Table 2.** Descriptive statistics, morphometric relationships and relative growth between carapace width, carapace length and total weight of the pea crabs (*Pinnotheres pisum*) depending on their sex, developmental and female maturity stages

Sex:	N	Size and weight			Morphometric relationship			Relative growth	
		CW	CL	TW	Equation	r	b ± SE (95% CI)	t-test (b)	Type
Juveniles (J)	13	1.7 ± 0.2 (1.3–2.2)	1.8 ± 0.3 (1.4–2.4)	0.003 ± 0.001 (0.001–0.005)	CL = 0.986 CW <sup>1.110</sup>	0.959***	1.110 ± 0.100 (0.891–1.329)	1.108 <sup>ns</sup>	I =
					TW = 0.0006 CW <sup>2.618</sup>	0.669*	2.618 ± 0.876 (0.690–4.547)	0.436 <sup>ns</sup>	I =
Males (M)	60	3.5 ± 0.6 (2.4–4.8)	3.6 ± 0.6 (2.5–4.9)	0.02 ± 0.01 (0.01–0.05)	CL = 1.139 CW <sup>0.905</sup>	0.966***	0.905 ± 0.032 (0.841–0.969)	2.975**	A–
					TW = 0.0007 CW <sup>2.690</sup>	0.837***	2.690 ± 0.231 (2.228–3.153)	1.341 <sup>ns</sup>	I =
Females (F)	33	8.3 ± 2.8 (3.3–13.5)	7.9 ± 2.5 (3.4–12.6)	0.15 ± 0.10 (0.03–0.35)	CL = 1.394 CW <sup>0.818</sup>	0.880***	0.818 ± 0.079 (0.656–0.980)	2.294*	A–
					TW = 0.002 CW <sup>1.924</sup>	0.910***	1.924 ± 0.158 (1.602–2.246)	6.820***	A–
Hard females (HF)	5	4.3 ± 0.6 (3.3–4.9)	4.2 ± 0.6 (3.4–5.0)	0.03 ± 0.004 (0.03–0.04)	CL = 1.185 CW <sup>0.868</sup>	0.917*	0.868 ± 0.218 (0.173–1.562)	0.607 <sup>ns</sup>	I =
					TW = 0.014 CW <sup>0.547</sup>	0.647 <sup>ns</sup>			
Soft females (SF)	28	9.1 ± 2.4 (5.0–13.5)	8.6 ± 2.0 (5.8–12.6)	0.17 ± 0.09 (0.04–0.35)	CL = 2.032 CW <sup>0.652</sup>	0.734***	0.652 ± 0.118 (0.409–0.894)	2.950**	A–
					TW = 0.003 CW <sup>1.770</sup>	0.824***	1.770 ± 0.239 (1.279–2.262)	5.143***	A–
Total	106	4.8 ± 2.9 (1.3–13.5)	4.7 ± 2.7 (1.4–12.6)	0.06 ± 0.08 (0.001–0.35)	CL = 1.111 CW <sup>0.924</sup>	0.982***	0.924 ± 0.017 (0.890–0.959)	4.347***	A–
					TW = 0.0009 CW <sup>2.374</sup>	0.964***	2.374 ± 0.065 (2.246–2.502)	9.694***	A–

N, number of individuals; CW, mean carapace width (mm); CL, mean carapace length (mm); TW, mean total weight (g); Size and weight data presented as mean ± SD and respective range (minimum – maximum); r, correlation coefficient; b, allometry coefficient; SE, standard error; 95% CI, 95% confidence interval.

Superscript letters and asterisks denote statistical significance level (P-value): <sup>ns</sup>, P > 0.05 (not significant); \*, P < 0.05 (significant); \*\*, P < 0.01 (highly significant); \*\*\*, P < 0.001 (very highly significant). I =, isometry; A–, negative allometry.

**Table 3.** Geographical distribution, bivalve host species and infestation rate of the pea crab *Pinnotheres pisum* throughout its distributional range in the Atlantic Ocean and Mediterranean Sea

Location	Study period	Host bivalve species	Infestation rate (%)	References
North Sea				
Loreley Bank	1985	<i>Spisula solida</i>	0.05	Becker and Türkay (2017)
Helgoland Trench	1983–1992	<i>Modiolus modiolus</i>	46.4	Becker and Türkay (2017)
	2003–2010	<i>Modiolus modiolus</i>	17.2	Becker and Türkay (2017)
Dogger Bank	2004–2010	<i>Donax vittatus</i>	2.1	Becker and Türkay (2017)
		<i>Gari fervensis</i>	8.1	Becker and Türkay (2017)
		<i>Mactra stultorum</i>	21.0	Becker and Türkay (2017)
		<i>Spisula elliptica</i>	0.9	Becker and Türkay (2017)
North Eastern Atlantic Ocean				
Westport, Ireland	2017–2019	<i>Cerastoderma edule</i>	1.0	Bruzos <i>et al.</i> (2020)
Southwest England	1968–1969	<i>Mytilus edulis</i>	30.3	Seed (1971)
		<i>Mytilus galloprovincialis</i>	1.4	Seed (1971)
Devon and Cornwall coasts, England	1966–1968	<i>Mytilus edulis</i>	0.7–46.4	Seed (1969)
Calshot, Solent, southern England	1972–1974	<i>Mytilus edulis</i>	54.2	Haines <i>et al.</i> (1994)
Elmore, Solent, southern England	1972–1974	<i>Mytilus edulis</i>	34.4	Haines <i>et al.</i> (1994)
Cotentin Peninsula, Normandy, France	1993	<i>Mytilus edulis</i>	20.0	Mizan (1993)
Arcachon, France	2017–2019	<i>Cerastoderma edule</i>	2.0	Bruzos <i>et al.</i> (2020)
Arcachon, France	2017–2019	<i>Cerastoderma edule</i>	19.0	Montaudouin <i>et al.</i> (2021)
Plentzia, Spain	2017–2019	<i>Cerastoderma edule</i>	82.0	Bruzos <i>et al.</i> (2020)
Setúbal, southwest coast, Portugal	2017	<i>Chamelea gallina</i>	1.5	Present study
		<i>Donax trunculus</i>	0.06	Present study
		<i>Donax vittatus</i>	0.7	Present study
		<i>Spisula solida</i>	1.4	Present study
Algarve, south coast, Portugal	2017	<i>Chamelea gallina</i>	0.04	Present study
		<i>Donax semistriatus</i>	0.2	Present study
		<i>Donax trunculus</i>	0.07	Present study
		<i>Spisula solida</i>	0.1	Present study
Ria Formosa lagoon, south coast, Portugal	2017	<i>Cerastoderma edule</i>	3.0	Montaudouin <i>et al.</i> (2021)
Ria Formosa lagoon, south coast, Portugal	2017	<i>Cerastoderma</i> sp.	1.5	Perez-Miguel <i>et al.</i> (2019a)
Isla Cristina, Andalusia, Spain	2017	<i>Mytilus galloprovincialis</i>	1.1	Perez-Miguel (2018)
Carreras estuary, Andalusia, Spain	2017	<i>Cerastoderma</i> sp.	1.0	Perez-Miguel <i>et al.</i> (2019a)
Piedras estuary, Andalusia, Spain		<i>Cerastoderma</i> sp.	0.2	Perez-Miguel <i>et al.</i> (2019a)
Merja Zerga lagoon, Morocco	2006	<i>Cerastoderma edule</i>	10–13 (inner area) 20–70 (outer area)	Gam <i>et al.</i> (2008)
Mediterranean Sea				
Rincon La Victoria, Andalusia, Spain	2017	<i>Chamelea gallina</i>	0.6	Perez-Miguel <i>et al.</i> (2019a)
Adra, Andalusia, Spain	2017	<i>Mytilus galloprovincialis</i>	0.7	Perez-Miguel (2018)
Garrucha, Andalusia, Spain	2017	<i>Mytilus galloprovincialis</i>	1.2	Perez-Miguel (2018)
Alboran Sea, Spain	2018	<i>Mytilus galloprovincialis</i>	25.0	Marco-Herrero <i>et al.</i> (2020)
Adriatic Sea				
Rovinj, Croatia	2003–2009	<i>Mytilus galloprovincialis</i>	3.6	Becker and Türkay (2017)
		<i>Ostrea edulis</i>	15.6	Becker and Türkay (2017)
		<i>Pinna nobilis</i>	46.2	Becker and Türkay (2017)

strategies: firstly, pea crabs releasing a chemical cue that discourages conspecifics from entering their bivalve hosts, coupled with resident pea crabs showing aggressive behaviour towards invading conspecifics (e.g. Bell, 1984; Haines et al., 1994; Soong, 1997; Takeda et al., 1997; Drake et al., 2014); secondly, mature males swimming ability enable them to survive for some time outside bivalve hosts, whereas mature free-living females have never been reported in nature, further suggesting that males prompt mating encounters while females usually remain inside bivalve hosts (Haines et al., 1994).

Although the number and relative frequency of pea crabs displayed an evident decreasing pattern with increasing depth of occurrence of commercial bivalve species, their infestation rate exhibited a clear growing trend with increasing fishing depth of host bivalves (graphically illustrated in Figure 6). In addition, the infestation rate was clearly higher in the southwest coast (IR = 0.31%) where fishing depth is higher ( $10.1 \pm 5.3$  m), than in the south coast (IR = 0.08%) where commercial bivalves occur and are exploited at lower bathymetrics ( $6.2 \pm 2.8$  m). These results corroborate previous studies that reported increasing *P. pisum* infestation rates increase with growing depth, namely from intertidal to subtidal sampling stations (Houghton, 1963; Seed, 1969; Gam et al., 2008; Becker, 2010; Perez-Miguel et al., 2019a). Moreover, higher infestations by *P. pisum* occurred in *S. solida* and *C. gallina* in the southwest coast and lower infestations occurred in *D. trunculus* in the south and southwest coasts, which is probably related to the preferential habitat of these bivalve species. In fact, while the former host species have a similar depth distribution (*C. gallina*: 3–10 m; *S. solida*: 3–14 m), *D. trunculus* occurs at much shallower depths (0–5 m, with higher densities at 3 m) (Gaspar et al., 2002c; Anjos et al., 2018), which in intertidal areas implies a continuous unburial and reburial that certainly makes more difficult the colonisation of *D. trunculus* by *P. pisum* (Perez-Miguel et al., 2019a).

In the present study, the size of *P. pisum* ranged from 1.3 to 13.5 mm CW, which is fairly similar to the ranges reported for this pea crab species inhabiting *M. edulis* in southern England (1.0–13.0 mm CW) (Haines et al., 1994) and southwestern England (2.1–18.0 mm CW) (Atkins, 1926), as well as colonising both *M. edulis* and *M. galloprovincialis* from southwest England (1.0–15.0 mm CW) (Seed, 1969). Moreover, a clear size dimorphism between sexes (males smaller than females) and female maturity stages (hard females smaller than ovigerous soft females) was detected in the present study, further corroborating previous studies of this pea crab species (Atkins, 1926; Seed, 1969; Haines et al., 1994). The space available inside the bivalve host shell apparently influences the distribution and size of colonising pea crabs (Haines et al., 1994; Becker and Türkay, 2017). Similar to other studies that recorded larger pea crabs inhabiting larger bivalve hosts (e.g. Houghton, 1963; Seed, 1969; Haines et al., 1994), the correlations established in the present study between bivalve shell length and pea crab carapace width confirmed a general trend for larger bivalves hosting larger *P. pisum*. Consequently, just like previously reported by Haines et al. (1994), due to *P. pisum* size dimorphism between sexes, smaller bivalves were mainly colonised by male pea crabs (smaller males prevailed in 20–30 mm bivalves), whereas larger bivalves were mostly inhabited by female pea crabs (larger females predominated in 30–40 mm bivalves).

The overall population of *P. pisum* exhibited negative allometries between morphometric variables, indicative of slower growth rate in both carapace length and total weight compared to carapace width ( $b = 0.924$  and  $b = 2.374$ , respectively). However, while the morphometric relationships between carapace width and total weight displayed an isometry in males ( $b = 0.905$ ) and a negative allometry in females ( $b = 1.924$ ), the relative growth between carapace width and carapace length was hypoallometric in both sexes ( $b = 0.905$

and  $b = 0.818$ , respectively), reflecting a progressive widening of pea crabs carapace throughout ontogeny. Just for comparison purposes, isometries in males ( $b = 2.979$ ) and hard females ( $b = 2.519$ ) and negative allometries in soft females ( $b = 2.597$ ) between carapace width and total weight, were also recorded in *P. bicristatus* colonising *Anomia ephippium* along the Atlantic and Mediterranean coasts of Andalusia (Spain) (Cuesta et al., 2019). Moreover, such morphometric relationships also detected differential growth between female maturity stages, with hard females being isometric ( $b = 0.868$ ) and soft females being hypoallometric ( $b = 0.652$ ), with this gradual enlargement of the carapace during maturation probably allowing soft females to carry more eggs at the subsequent ovigerous stage. Similarly to the present study, morphometric relationships between carapace width and carapace length of *A. monodi* colonising *M. galloprovincialis* in southern Portugal also revealed a negative allometry in males ( $b = 0.961$ ), isometric growth in hard females ( $b = 0.985$ ) and another hypoallometry in non-ovigerous soft females ( $b = 0.805$ ) (Santos et al., 2022).

Previous studies reported that bivalves infestation by pea crabs can injure their gills and impair filtering efficiency, which might lead to reduced growth, reproductive maturation and condition index (e.g. Stauber, 1945; Christensen and McDermott, 1958; Sun et al., 2006; Mena et al., 2014; Yasuoka and Yusa, 2017; Perez-Miguel et al., 2018, 2019a; Cuesta et al., 2020). Just for instance, populations of *M. edulis* with a high prevalence of *P. pisum* in southwest England exhibited gill damages, with infested mussels showing lower tissue weight than uninfested mussels of comparable size (Seed, 1969). In fact, more recently, *P. pisum* has been considered a constant pest in mussels and oysters in European coasts (Becker, 2010). The severity of pea crabs harmful effects on their hosts depends on diverse factors, such as the prevalence, intensity and duration of the infestation, as well as on the relative size of pea crabs and bivalve hosts (e.g. Sun et al., 2006; Trottier et al., 2012; Mena et al., 2014; Perez-Miguel et al., 2018; Cuesta et al., 2020). Although this study did not aim to assess host bivalves condition index, the fairly low infestation rates of *P. pisum* recorded in all commercial bivalves do not constitute a health concern for these highly valued species and fishery-exploited resources in the south and southwest coasts of Portugal. Overall, the present study provided further information and valuable insights on bivalves' colonisation by *P. pisum*, which should be periodically monitored in the mid- and long-term under a climate change scenario, in order to follow its evolution, eventual harmful effects on hosts' condition index and, therefore, on commercial bivalve populations.

**Data availability.** The data that support the findings of this study are available from the corresponding author, upon reasonable request.

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**Author's contributions.** L. Nicolau: Data curation, formal analysis, writing, review and editing. P. Vasconcelos: conceptualisation, data curation, formal analysis, writing, review and editing. F. Pereira: conceptualisation, fishing surveys and laboratory sampling, methodology, investigation, data curation, formal analysis, writing, review and editing. A.N. Carvalho: conceptualisation, fishing surveys and laboratory sampling, data curation, formal analysis, writing, review and editing. D. Piló: conceptualisation, fishing surveys and laboratory sampling, data curation, formal analysis, writing, review and editing. M.B. Gaspar: funding acquisition, supervision and coordination.

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**Competing interest.** None.

**Ethical standards.** Not applicable.

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