

Research Article

Cite this article: Daversa DR, Baxter E, Rosa GM, Sergeant C and Garner TWJ (2024). Standard methods for marking caudate amphibians do not impair animal welfare over the short term: An experimental approach. *Animal Welfare*, **33**, e24, 1–7
<https://doi.org/10.1017/awf.2024.26>

Received: 02 October 2023
 Revised: 22 March 2024
 Accepted: 27 March 2024

Keywords:

Amphibians; animal behaviour; animal welfare science; experimental biology; marking methods; wildlife health

Corresponding author:

David R Daversa;
 Emails: ddaversa@gmail.com

Author contributions:

Conceptualisation: TG; Data curation: DRD; Investigation: EB, GMR, CS, TG; Formal analysis: DRD; Methodology: DRD, EB, GMR, CS, TG; Project administration: EB, GMR, CS; Supervision: GMR; Writing – original draft: TG; Writing – review & editing: DRD, EB, GMR, TG

*This article has been updated since original publication. A notice detailing the change has also been published

© The Author(s), 2024. Published by Cambridge University Press on behalf of The Universities Federation for Animal Welfare. This is an Open Access article, distributed under the terms of the Creative Commons Attribution-NonCommercial licence (<http://creativecommons.org/licenses/by-nc/4.0>), which permits non-commercial re-use, distribution, and reproduction in any medium, provided the original article is properly cited. The written permission of Cambridge University Press must be obtained prior to any commercial use.



Twitter: @UFAW_1926
 webpage: <https://www.ufaw.org.uk/>

Standard methods for marking caudate amphibians do not impair animal welfare over the short term: An experimental approach*

David R Daversa^{1,2} , Ella Baxter², Gonçalo M Rosa^{2,3,4}, Chris Sergeant² and Trenton WJ Garner^{2,5,6}

¹La Kretz Center for California Conservation Science, Institute of the Environment and Sustainability, University of California, Los Angeles, USA; ²Institute of Zoology, Zoological Society of London, Regents Park, London, UK; ³Biodiversity Research Institute (IMIB-CSIC, Universidad de Oviedo, Principality of Asturias), Mieres, Spain; ⁴Centre for Ecology, Evolution and Environmental Changes (CE3C) & Global Change and Sustainability Institute (CHANGE), Faculdade de Ciências da Universidade de Lisboa, Lisboa, Portugal; ⁵Department of Genetics, Evolution and Environment, UCL, London, UK and ⁶Unit for Environmental Sciences and Management, North-West University, Potchefstroom, South Africa

Abstract

Major advancements in ecology and biodiversity conservation have been made thanks to methods for marking and individually tracking animals. Marking animals is both widely used and controversial due to the potential consequences for animal welfare, which are often incompletely evaluated prior to implementation. Two outstanding knowledge gaps concerning the welfare consequences of individual marking are their short-term behavioural impacts and the relative impacts from marking versus the handling of animals while carrying out procedures. We addressed these knowledge gaps through an experimental study of alpine newts (*Ichthyosaura alpestris*) in which we varied handling and marking procedures. Examining individual responses to handling, toe clipping and visible implant elastomer (VIE) injection over 21 days showed that handling and marking elicited increased newt activity and hesitancy to feed compared to animals that did not get handled or marked. These effects were apparent even when animals were handled only (not marked), and marking did not further increase the magnitude of responses. Increases in newt activity and feeding hesitancy were transient; they were not observed in the weeks following handling and marking. While previous studies emphasise the welfare impacts of marking procedures themselves, these findings highlight that handling alone can elicit behavioural changes with possible costs to welfare. Yet, the transient nature of behavioural responses suggests that immediate costs of handling may be subsequently compensated for in the short term.

Introduction

Confronting the global biodiversity crisis requires a critical understanding of how threatening processes impact wildlife populations. Demographic studies are the most common approach for investigating impacts, studies which frequently require the capacity to discriminate between individuals (Major 2020). For many species, this requires handling and the application of an artificial mark. The minimising of pain and distress is a fundamental principle of wildlife marking and is also a legal requirement under numerous animal protection legislations; the mark ideally should not significantly impair the welfare of the marked individual (e.g. Locatelli *et al.* 2019). Despite the widespread acceptance of prioritising welfare (Dawkins 2006; Hecht 2021; Soryl *et al.* 2021), the impacts of capture and marking are not always tested, or at least not revisited regularly to reassess the consequence of marking methods under different circumstances (Soulsbury *et al.* 2020). As a result, some marking techniques have gained widespread application across taxonomic groups without explicit tests of impacts on each species, or reapplication in new populations, with examples including colour-coded bird banding, fish adipose fin clipping and amphibian and reptile toe clipping (Perry *et al.* 2011; Tinbergen *et al.* 2014; Uglem *et al.* 2020).

Amphibians are the most threatened vertebrate taxon (Luedtke *et al.* 2023), a conclusion largely justified through the outputs of numerous studies of population dynamics utilising individual marking strategies (e.g. Storfer 2003; Bucciarelli *et al.* 2020). Although less invasive methods are available, their use is frequently hampered by a range of limitations. For instance, individual skin patterns, while an option, sometimes lack distinctiveness or prove suitable solely for short-term studies due to their tendency to change over time (Arntzen *et al.* 2004; Ferner 2007; Aevansson *et al.* 2022; Kenyon *et al.* 2009); conversely, radio tracking, while a powerful tool, presents size restrictions, feasible only for relatively larger specimens, while bearing substantial economic costs (Ferner 2007; Andreone *et al.* 2013; Daversa *et al.* 2017). As a result, researchers

have traditionally resorted to established methods like toe clipping (the complete or partial removal of digits; e.g. Perry *et al.* 2011) and, more recently, visible implant elastomer (VIE) injection (subcutaneous injection of silicone-based polymer that hardens after injection), which are attractive due to the inexpensive costs and relatively fast execution that permits large sample sizes.

Toe clipping and VIE injection, while instrumental in assessing how threatening processes impact demography, have sparked controversies concerning welfare implications (Narayan *et al.* 2011; Perry *et al.* 2011; Palmer *et al.* 2023). Critical studies of their impacts have revealed real and potential impacts on individual performance and survival (Bloch & Irschick 2005; Narayan *et al.* 2011). Marking may directly reduce survival through physical impairment (Bloch & Irschick 2005), for example, how improperly cured elastomer can migrate to organs where it can presumably impair organ function (McCarthy & Parris 2004; Cabot *et al.* 2021). Even when no physical impairment occurs, marking may elicit behavioural responses immediately post-marking that have the potential to mediate downstream welfare and survival. Short-term behavioural responses to marking have been largely unexplored (but see Sapsford *et al.* 2014), a knowledge gap which may overlook opportunities to improve welfare without abandoning methods, particularly for cases where the effects are transitory.

While most extant amphibians are anurans, caudates are disproportionately more threatened (57.3% caudates vs 33.2% anurans; IUCN 2023). Despite this, the majority of assessments of marking techniques have focused on anurans (but see Ott & Scott 1999; Davis & Ovaska 2001; Kinkead *et al.* 2006). Here, we examined the behavioural impacts of two invasive marking techniques on the European alpine newt (*Ichthyosaura alpestris*), a caudate species that is often the subject of numerous demographic and behavioural studies (for recent examples, see Diego-Rasilla & Phillips 2021; Gvoždík 2022; Bernabò *et al.* 2023). The overriding aim of the study was to systematically evaluate the short-term effects of handling versus toe clipping and VIE marking on newt behaviour. We achieved this aim through an experiment designed to discriminate between the effects of handling from the impacts of marking on newt activity, shelter use, and feeding.

Materials and methods

The following procedures were approved by the Zoological Society of London's Ethics Committee prior to commencement and licensed by the Home Office (PPL 80/2466 to TG). Alpine newts were collected from invasive populations in the UK and treated prophylactically to eliminate infections with *Batrachochytrium dendrobatidis*: individuals were placed in tanks and treated with an itraconazole solution bath (1 mg L⁻¹; Sporanox, Janssen-Cilag, Titusville, NJ, USA) for 5 min, followed by 10 min in ringer solution once a day for seven days (Garner *et al.* 2009). Our previous work showed that post-treatment newt behaviour, including their activity levels and habitat use, was still informative for comparative studies examining health and welfare (Daversa *et al.* 2018).

Newts (n = 40; 32 female, 8 male) were weighed to the nearest 0.1 g, measured snout-to-vent (SVL) to the nearest mm and then individually housed in 5-L plastic tubs (Really Useful Boxes, Castleford, UK; 340 × 200 × 125 mm; length × width × height), where they were given five days to acclimate. Each tub was divided in half, one half containing 1.5 L aged tap water while the other was filled with autoclaved gravel (5–20-mm diameter). Cover objects (small PVC shelters) were embedded on gravel substrate and submerged

in water. During acclimation and throughout the experiment, 1/3 of the tub water was replaced twice per week and debris removed from the aquatic side using sterile turkey basters to maintain sanitary aquatic conditions. We fed newts earthworms (*Lumbricus terrestris*) after water changes (twice per week; total mass at each feed 0.4–0.5 g). The housing and husbandry protocols followed our previous experimental studies (Daversa *et al.* 2018) and pilot work (unpublished) with alpine newts in the same facilities, which indicated no adverse welfare impacts to newts. To further ensure the minimising of adverse welfare impacts, all work was overseen by licenced veterinarians and trained herpetologists.

Experimental treatments were designed to discriminate between the behavioural effects of handling versus the effects of either marking protocol. Treatment #1 (placebo) involved capturing and holding newts in the hand before returning to the housing unit. Treatments #2 (toe clipping) and #3 (VIE) comprised hand-capture followed by distinct procedures: complete removal of the middle digit of the rear right foot and return to the housing unit, and hand-capture followed by injection of approximately 0.015 mL of elastomer (Northwest Marine Technology, Anacortes, WA, USA) in the ventral side of the thigh of the right rear leg and return to the housing unit. These first three treatments were standardised to each last approximately 4 min, while newts in the #4 treatment (control) were left undisturbed during the marking period. We used a sex ratio of eight females for every two males across the four treatments, but within sex, newts were assigned randomly to treatments.

Newt behaviour was sampled for 10 min immediately following the return to the aquatic part of the housing unit (Winandy & Denoël 2011; Bateson & Martin 2021). One observer recorded the position (aquatic versus terrestrial, hidden with head and greater than half the body under cover versus in view with head and greater than half the body exposed) and activity (active versus stationary) for 30 s at 60-s intervals (Winandy & Denoël 2011; Bateson & Martin 2021). The same observer repeated focal sampling 48 h and seven days after the initial sampling. To eliminate treatment effects of time of day on sampling, we assigned one individual per treatment to one of ten sampling groups and collected behavioural data simultaneously across all four treatments in a sampling group. We recorded feeding behaviour immediately after handling/marketing and once weekly after that for three weeks, as a yes/no event and as latency (time taken to show interest in food up to 10 min after the food was added to the enclosure). We also recorded total consumption. The experiment was concluded 21 days after marking, with euthanasia performed by immersing the animals in buffered MS-222 (American Veterinary Medical Association [AVMA] Panel on Euthanasia 2020).

For data analyses, we characterised three behavioural responses: activity level (the proportion of time active), visibility level (proportion of time visible), and habitat use (proportion of time aquatic). We chose these responses for their functional role in newt avoidance of threats; activity and visibility level are commonly measured responses to predation risk (Winandy & Denoël 2013; Daversa *et al.* 2021), and habitat use relates to reproductive opportunities, predation and parasitism risk (Winandy *et al.* 2015; Daversa *et al.* 2018). We ran Generalised Linear Mixed Models (GLMMs) in R (R Core Team 2023; lme4 package), one each for each response, to examine: (1) how newt behaviour changed over the course of the experiment; and (2) how handling and marking influenced newt behaviour. We used a binomial error structure for GLMMs and included a unique newt identification as a random effect to account for repeated sampling. We also included experimental treatment, observation number, and their interaction as

fixed effects. We assessed the influence of the fixed effects on newt responses by performing likelihood ratio tests with a Chi-squared distribution, using the `dropterm()` function in R (MASS package). In cases where newt behaviour depended upon the timing of observation, we also assessed the effect of experimental treatment on newt behaviour separately for each observation. To do so, we ran GLMMs with a binomial error structure and an observation-level random effect to account for overdispersion of the data (Harrison 2014). These models included experimental treatment as a single, fixed effect. We compared model coefficients for the three treatment groups (placebo, toe clipping, VIE) against coefficients for the control groups to determine whether newts that were handled and/or marked behaved differently than controls.

To test for feeding latency, we first performed a ‘time-to-event’ analysis (i.e. survival analysis). The response variables used were: (1) the time within the 10-min observation period when newts were first observed feeding on worms (0–10 min); and (2) an event status indicating whether newts fed within the 10-min observation period (0 = no observed feeding, 1 = observed feeding). For example, newts that were not observed feeding were assigned values of ten for the observation period and 0 for the event status. Second, we ran Generalised Linear Models (GLMs) to detect treatment effects on total consumption as a proportion of the total food provided, including experimental treatment as the fixed effect and again using a binomial error structure. We again used likelihood ratio tests with a Chi-squared distribution and the `dropterm()` function in R to examine the overall influence of experimental treatment on total worm consumption. We also compared model coefficients for the three treatment groups against coefficients for the control group to

determine whether newts that were handled and/or marked consumed more or less than controls. We explored feeding trends over time by running GLMMs with a binomial error structure and including individual ID as a random effect to account for repeated sampling and interactive effects of treatment and sampling week. We tested for interactive and additive effects by performing likelihood ratio tests with a Chi-squared distribution and the `dropterm()` function.

Results

The proportion of time that newts were active varied over observations (fixed effect of sampling event, $\chi^2 = 37.28$; $P < 0.001$; Figure 1[a]), though handling/marking did not generally influence the proportion of time that newts were active (treatment: sample interaction, $\chi^2 = 3.89$; $P = 0.273$; main effect of treatment, $\chi^2 = 4.98$; $P = 0.174$). However, examining activity patterns on a sample-by-sample basis revealed the effects of all experimental treatments at a specific time-point. Newts in all three handling/marking treatments were more active than newts in the control group immediately following handling/marking ($t \geq 2.06$; $P \leq 0.039$), an effect not detected in subsequent samples (Figure 1[a]). Newt visibility varied across different observations (fixed effect of sampling event, $\chi^2 = 43.36$; $P < 0.001$); yet again, handling/marking newts did not generally influence the proportion of time that newts were visible versus hidden in shelters (treatment: sample interaction, $\chi^2 = 4.96$; $P = 0.175$; main effect of treatment, $\chi^2 = 4.26$; $P = 0.236$). Examining visibility on a sample-by-sample basis did not reveal any event-specific effects (Figure 1[b]). Newt

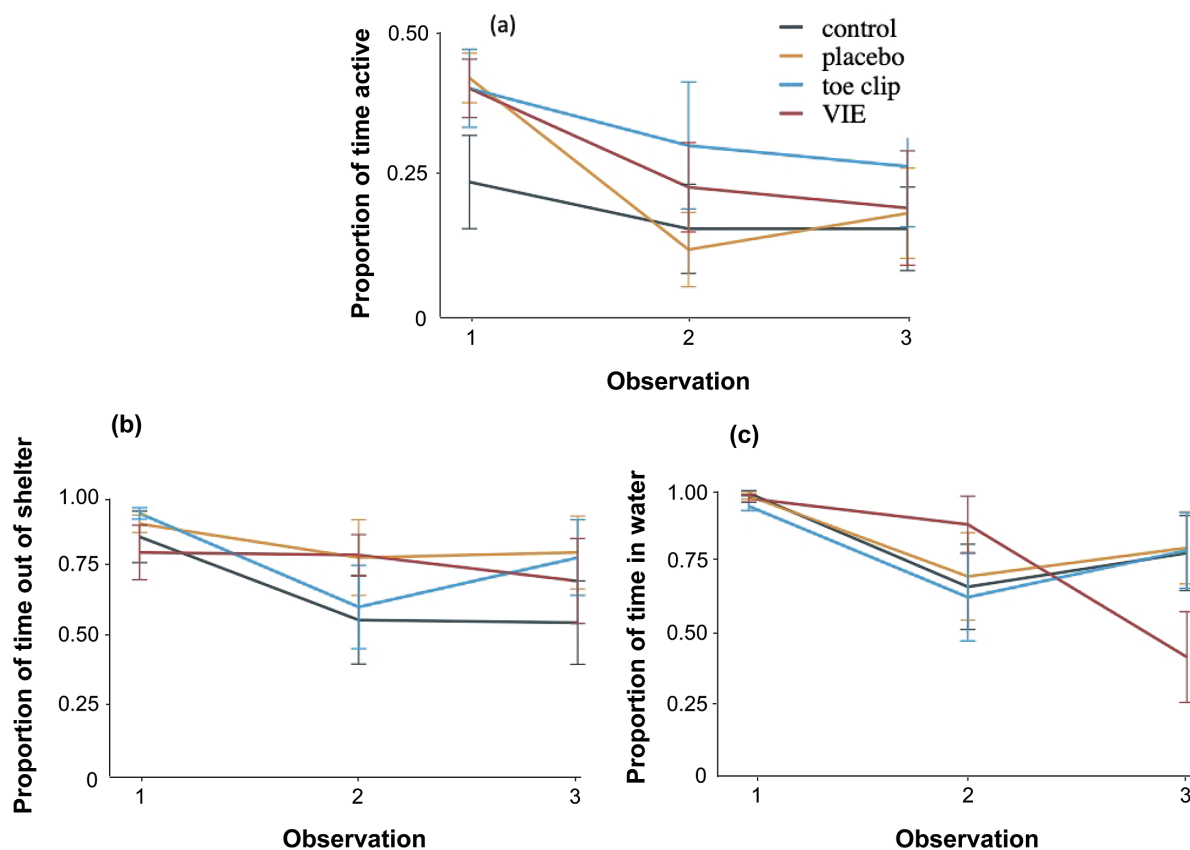


Figure 1. The proportion of observations that newts were (a) active (i.e. moving), (b) outside of their shelter and (c) in water as opposed to on land. Newts were sampled on three occasions: (1) immediately after treatment; (2) 48 h after treatment; and (3) one week after treatment. VIE: visible implant elastomer.

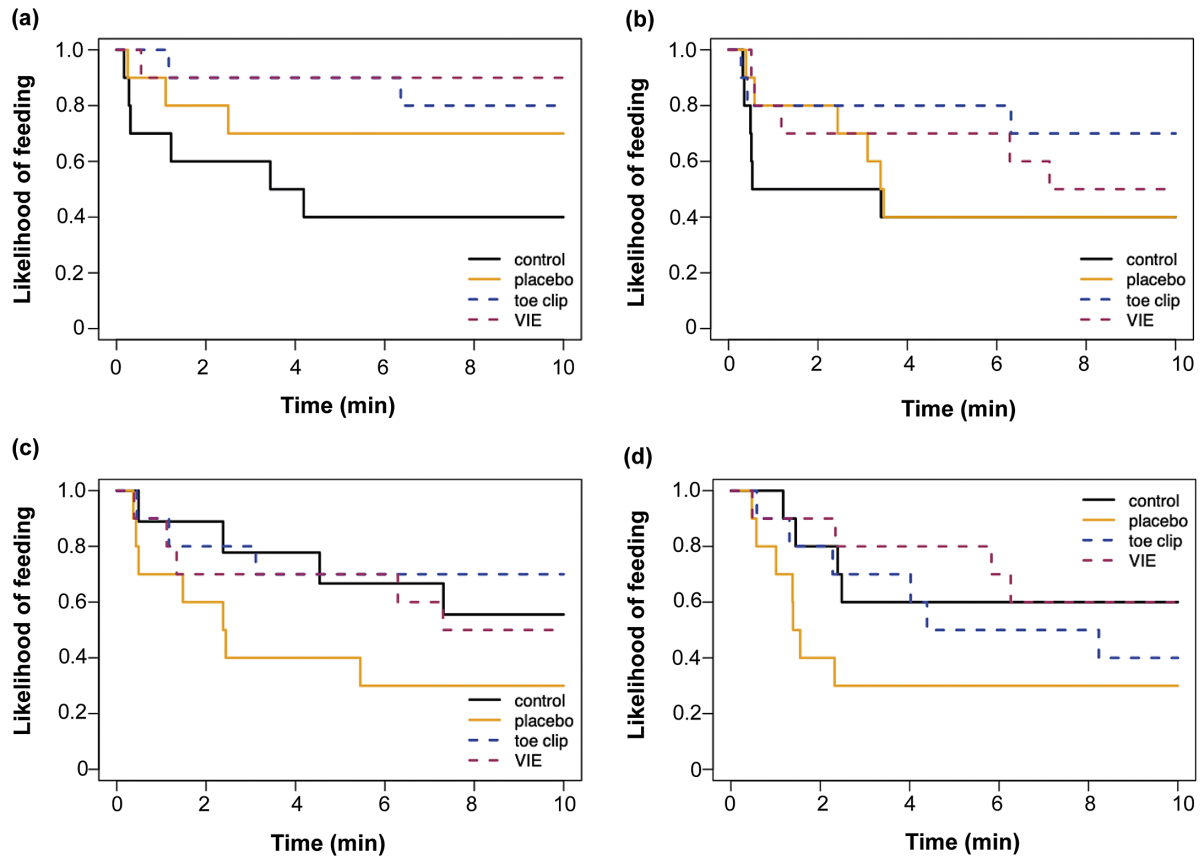


Figure 2. Probability that newts were feeding at a given time during each ([a]–[d]) of the 10-min observation periods (Feeding 1–4, respectively) performed weekly for four weeks (Kaplan-Meier plots for the time-to-event) for each of the four handling/tagging treatments: control, placebo, toe clip, and visible implant elastomer (VIE). Observations began immediately after food was administered into new tanks.

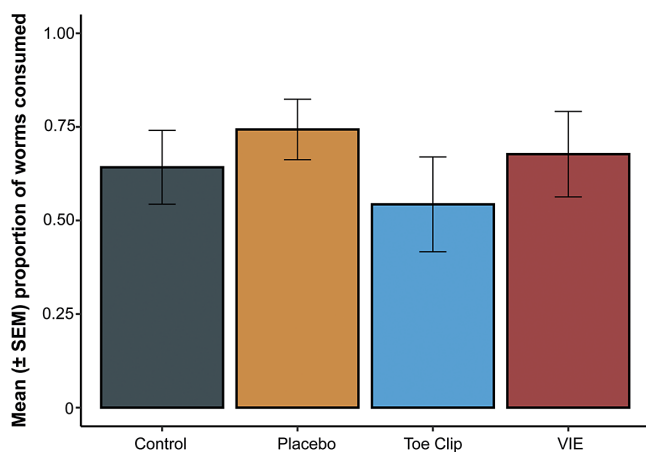


Figure 3. The mean proportion of worms consumed by newts for each of the four handling/tagging treatments. Newts were fed a total of 12 worms over the course of the experiment (four feedings of three worms). Error bars show the standard error of the mean. VIE = visible implant elastomer.

habitat use was influenced by handling/marking, depending on the sampling period and marking protocol (treatment:sample interaction, $\chi^2 = 72.05$; $P < 0.001$). Newts that were VIE-tagged spent proportionally less time in the water than control and other treatment newts at the last sampling ($t = -4.28$; $P < 0.001$; Figure 1[c]).

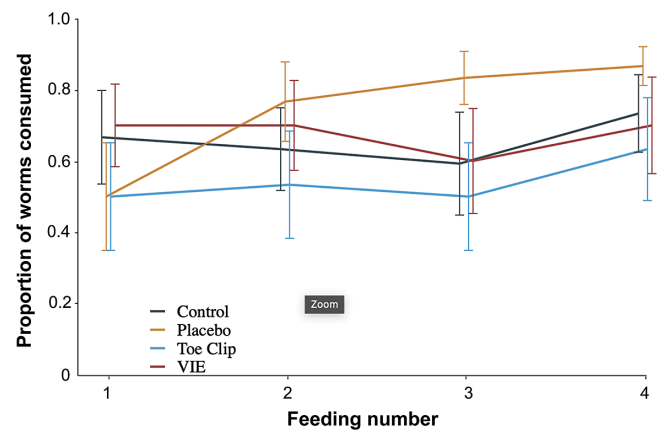


Figure 4. The mean proportion of worms consumed by newts on a weekly basis, with different lines distinguishing the specific marking/handling treatment received. Newts were fed three worms per feeding across four feedings ($n = 12$). The lines denote mean values within the treatment groups and error bars denote the standard error of the mean. VIE = visible implant elastomer.

Although there was no statistically significant treatment effect on feeding latency (including experimental treatment in Cox Proportional Hazards model only marginally improved fit, $\chi^2 = 6.60$; $P = 0.086$), in qualitative terms, newts in the control group were, on average, quicker to start feeding after handling/marking than newts in treatment groups (Figure 2[a]). The above trends in feeding

latency were not apparent after the first week (Figure 2[b]–[d]). There was a general treatment effect on the proportion of worms that newts consumed across the experiment (dropping treatment from the model reduced goodness of fit, $\chi^2 = 10.99$; $P = 0.012$), but this effect was due to differences between newts receiving a placebo and newts that were toe clipped ($z = 3.20$; $P = 0.001$; Figure 3). The proportion of worms that newts consumed in the three treatment groups generally did not differ from observed consumption in the control group ($P > 0.05$ in all cases; Figure 3). Examining worm consumption on a weekly basis revealed interactive effects between marking/handling and the time of sampling ($\chi^2 = 10.61$; $P = 0.014$), with effects largely driven by temporal changes in feeding by newts in the placebo group (Figure 4). Newts in the placebo group (handled only) initially consumed fewer worms on average compared to subsequent observations (Figure 4).

Discussion

Assessments of the welfare impacts involved in the process of marking amphibians do not always discriminate between handling and marking (but see Oropeza Sánchez *et al.* 2020). This overlooks the directionality of the welfare assessment process, embedded in Soulsbury *et al.*'s (2020) decision tree for marking wildlife. Here, the determination of both the necessity and welfare implications of capture precedes that of the impact of marking (Soulsbury *et al.* 2020; Figure 1). For a more meaningful assessment of the marking technique itself, then, the impacts of animal capture and restraint first need to be ascertained. Our study illustrates this, as we showed that the initial changes in activity by alpine newts elicited by handling and marking are largely attributable to the handling itself, and transient. Nevertheless, the initial increase in activity immediately after marking may not be in the best interests of the marked animal. Typically, reduced activity and immobility are amphibian responses to predator risk, and so increased activity possibly exposes animals to predators (Winandy *et al.* 2015; Chapman *et al.* 2017; Passos *et al.* 2017; Daversa *et al.* 2021). Yet, animals employ diverse anti-predator behaviours depending on the context (Polčák & Gvoždík 2014; Daversa *et al.* 2021). In the context of handling by humans, the increased activity we observed is likely an anti-predator escape behaviour initiated in response to what newts perceive as a predation attempt (Polčák & Gvoždík 2014). This hypothesis is supported by studies of other wildlife that mount anti-predator responses to human stimuli (Clinchy *et al.* 2016; Palmer *et al.* 2022). Still, increased activity and feeding reductions were not associated with an increased propensity to seek refuge under cover objects or to move onto land. The reasons for these behavioural patterns are unclear, but together they may be indicative of human interventions causing aquatic habitats to be preferential for newts, perhaps because adult newts can flee faster in water than on land (Gvoždík & Van Damme 2008).

The escape-like behaviours that we observed immediately after handling are characteristic of responses made in 'fear' and permit the hypothesis that human handling acts as a fear stimulus for newts (Zanette & Clinchy 2019; Daversa *et al.* 2021). Fear-like responses in wild animals are well-studied and have widespread ecological consequences (Zanette & Clinchy 2019), and frameworks for understanding fear in wildlife hold value to welfare science. For example, fear-like escape behaviours involve physiological stress responses that cause temporary distress, heightened energetic demands, and internal damage (i.e. 'wear and tear'; McEwen & Wingfield 2003; Wingfield 2005). Stress responses involve a

recovery period and may decrease investment in reproduction and feeding in caudates (Moore 1984; Bliley & Woodley 2012). In addition to relatively low initial feeding rates by newts in the placebo group, we observed increasing hesitancy to forage in two of the three handling treatments immediately after handling/mark-ing, a trend that was strongest in the two treatments where marking was involved. Stress-associated inappetence may explain this hesitancy, an argument supported by evidence in badgers (*Meles meles*) that human noise causes delayed feeding (Clinchy *et al.* 2016). Alternatively, given that newts increased activity immediately after the marking/handling, they may have simply been too distracted to eat. In either case, the responses that we observed provide evidence that human handling induces fear-like responses in caudate amphibians, as they do in other animal groups (Clinchy *et al.* 2016; Palmer *et al.* 2022).

There have long been calls for longitudinal studies in research of the ecology of fear (Daversa *et al.* 2021). Longitudinal data are essential for measuring the duration of pain and/or distress and the how either might be behaviourally manifested. This study marks a step toward addressing these calls by tracking individual responses over time. Doing so, we found that behavioural responses to handling and marking were transient, providing hope that distress caused to animals is only temporary. The transient nature of the responses also suggests that newts may be able to develop tolerance to human handling that mediates fear and associated impacts to welfare. Nevertheless, repeated human handling can have covert physiological costs that accumulate over individual lifespans (Wingfield 2005) and should be explored more deeply to understand the extent of welfare impacts of handling and marking. Extending our experimental design to longer time-periods, integrating measures of physiological rates (metabolic rate, hormone levels, etc) and assessing physical damage from fear responses would mark a further step toward understanding how human handling and marking impact welfare in caudates and other wild animals.

Longitudinal studies of welfare could help to resolve open questions concerning the ethics of amphibian research. The ethical landscape of animal use in research has a long history, with procedures like amphibian toe clipping undergoing particular scrutiny (for a review, see Perry *et al.* 2011). Our study establishes amphibian handling as an additional facet in need of ethical consideration independently of any procedures. The ethics of handling animals is a function of its impact on an animal's ability to carry out adapted behaviours, its impact on the affective states of animals, and the duration of those impacts (Fraser *et al.* 1997). Our findings underscore the relevance of considering the duration of these impacts. Procedures yielding transient impacts, such as the heightened activity and reduced feeding observed in newts post-handling and marking, present unresolved ethical questions. Therefore, prior to drawing conclusions about the ethical treatment of transient welfare impacts, progressive long-term costs to animal function and affective states should be evaluated (Fraser *et al.* 1997). Longitudinal studies play a crucial role in this objective of future research into animal welfare.

Animal welfare implications

Animal welfare is defined in large part by how individuals cope with their environment (Broom 1991). While marking methods have received considerable attention in terms of the pain they inflict on animals (Palmer *et al.* 2023), marking may also affect functional behaviours in ways that compromise the capacity of animals to cope

with the environment and maintain sound health. The behavioural effects of marking methods used in field research are particularly understudied in amphibians despite the common usage of the methods and the high degree of concern this group of animals receives from biodiversity conservationists. We show in alpine newts that behavioural effects arise largely from the handling process, irrespective of the specific marking method used. From a behavioural perspective, handling seems to be as consequential to animal welfare as does the actual marking of animals.

The transient nature of the behavioural changes should be factored into cost-benefit analyses of marking animals and provide promise that methods need not be abandoned to uphold strong welfare standards concerning animal behaviour (not factoring in pain and suffering of the procedures). This is especially relevant with studies posing a high conservation benefit, for which certain levels of impact on individuals may be warranted. However, we only considered a single handling and marking event, which overlooks possible costs that arise distinctly from repeated interventions. In terms of welfare science, the transience of altered behaviour permits the hypothesis that immediate behavioural impacts of handling do not compromise long-term welfare of newts, at least when handling is not repeated over time. Testing this hypothesis was beyond the scope of this study, but the hypothesis could be tested in future studies via monitoring of welfare across age classes of handled and marked newts. We see such studies as imperative to progress in our understanding of the long-term effects of human interventions on the ability of wild animals to cope with their environment and, in turn, maintain good health and welfare.

Acknowledgements. We thank the Biodiversity and Conservation MRes programme at the University College of London and the Zoological Society of London (ZSL) for sponsoring this study. We also thank the students and staff in the Institute of Zoology, ZSL, for their kindness and moral support as we carried out the experiments. TG acknowledges the contribution of Research England to all research undertaken at the Institute of Zoology.

Competing interest. None.

References

- Aearsson U, Graves A, Carter KC, Doherty-Bone TM, Kane D, Servini F, Tapley B and Michaels CJ 2022 Individual identification of the lake oku clawed frog (*Xenopus longipes*) using a photographic identification technique. *Herpetological Conservation and Biology* 17: 67–75.
- Andreone F, Bergò PE, Mercurio V and Rosa GM 2013 Spatial ecology of *Scaphiophryne gottlebei* in the canyons of the Isalo Massif, Madagascar. *Herpetologica* 69: 11–21. <https://doi.org/10.1655/HERPETOLOGICA-D-12-00005>
- Arntzen JW, Goudie IBJ, Halley J and Jehle R 2004 Cost comparison of marking techniques in long-term population studies: PIT-tags versus pattern maps. *Amphibia-Reptilia* 25: 305–315.
- American Veterinary Medical Association (AVMA) Panel on Euthanasia 2020 AVMA Guidelines for the Euthanasia of Animals: 2020 Edition. American Veterinary Medical Association: Schaumburg, IL, USA.
- Bateson M and Martin P 2021 *Measuring Behaviour: An Introductory Guide*. Cambridge University Press: Cambridge, UK.
- Bernabò I, Iannella M, Cittadino V, Corapi A, Romano A, Andreone F, Biondi M, Gallo Splendore M and Tripepi S 2023 Survived the glaciations, will they survive the fish? Allochthonous ichthyofauna and alpine endemic newts: A road map for a conservation strategy. *Animals* 13: 871. <https://doi.org/10.3390/ani13050871>
- Bliley JM and Woodley SK 2012 The effects of repeated handling and corticosterone treatment on behavior in an amphibian (Ocoee salamander: *Desmognathus ocoee*). *Physiology & Behavior* 105: 1132–1139. <https://doi.org/10.1016/j.physbeh.2011.12.009>
- Bloch N and Irschick DJ 2005 Toe-clipping dramatically reduces clinging performance in a pad bearing lizard (*Anolis carolinensis*). *Journal of Herpetology* 39: 288–293. <https://doi.org/10.1670/97-04N>
- Broom DM 1991 Animal welfare: concepts and measurement. *Journal of Animal Science* 69: 4167–4175. <https://doi.org/10.2527/1991.69104167x>
- Bucciarelli GM, Clark MA, Delaney KS, Riley SPD, Shaffer HB, Fisher RN, Honeycutt RL and Kats LB 2020 Amphibian responses in the aftermath of extreme climate events. *Scientific Reports* 10: 3409. <https://doi.org/10.1038/s41598-020-60122-2>
- Cabot ML, Troan BV, Ange-van Heugten K, Schnellbacher RW, Smith D, Ridgley F and Minter LJ 2021 Migration and histologic effects of Visible Implant Elastomer (VIE) and Passive Integrated Transponder (PIT) tags in the marine toad (*Rhinella marina*). *Animals* 11: 3255. <https://doi.org/10.3390/ani11113255>
- Chapman TL, Spivey KL, Lundergan JM, Schmitz AL, Bast DL, Sehr EK and Gall BG 2017 Only fear the fatal foe: predation risk assessment by eastern newts (*Notophthalmus viridescens*) in response to common snapping turtles and other potential predators. *Ethology Ecology & Evolution* 29: 218–228. <https://doi.org/10.1080/03949370.2015.1137358>
- Clinchy M, Zanette LY, Roberts D, Suraci JP, Buesching CD, Newman C and Macdonald DW 2016 Fear of the human “super predator” far exceeds the fear of large carnivores in a model mesocarnivore. *Behavioral Ecology*: arw117. <https://doi.org/10.1093/beheco/arw117>
- Daversa DR, Fenton A, Dell AI, Garner TWJ and Manica A 2017 Infections on the move: how transient phases of host movement influence disease spread. *Proceedings of the Royal Society B: Biological Sciences* 284: 20171807. <https://doi.org/10.1098/rspb.2017.1807>
- Daversa DR, Hechinger RF, Madin E, Fenton A, Dell AI, Ritchie EG, Rohr J, Rudolf VHW and Lafferty KD 2021 Broadening the ecology of fear: non-lethal effects arise from diverse responses to predation and parasitism. *Proceedings of the Royal Society B* 288: 20202966.
- Daversa DR, Manica A, Bosch J, Jolles JW and Garner TWJ 2018 Routine habitat switching alters the likelihood and persistence of infection with a pathogenic parasite. *Functional Ecology* 32: 1262–1270. <https://doi.org/10.1111/1365-2435.13038>
- Davis TM and Ovaska K 2001 Individual recognition of amphibians: Effects of toe clipping and fluorescent tagging on the salamander *Plethodon vehiculum*. *Journal of Herpetology* 35: 217. <https://doi.org/10.2307/1566111>
- Dawkins M 2006 A user’s guide to animal welfare science. *Trends in Ecology & Evolution* 21: 77–82. <https://doi.org/10.1016/j.tree.2005.10.017>
- Diego-Rasilla FJ and Phillips JB 2021 Evidence for the use of a high-resolution magnetic map by a short-distance migrant, the Alpine newt (*Ichthyosaura alpestris*). *Journal of Experimental Biology* 224: jeb238345. <https://doi.org/10.1242/jeb.238345>
- Ferner JW 2007 *A review of marking and individual recognition techniques for amphibians and reptiles*. Society for the Study of Amphibians and Reptiles: Salt Lake City, UT, USA.
- Fraser D, Weary DM, Pajor EA and Milligan BN 1997 A scientific conception of animal welfare that reflects ethical concerns. *Animal Welfare* 6: 187–205. <https://doi.org/10.1017/S0962728600019795>
- Garner TWJ, Garcia G, Carroll B and Fisher MC 2009 Using itraconazole to clear *Batrachochytrium dendrobatidis* infection, and subsequent depigmentation of *Alytes muletensis* tadpoles. *Diseases of Aquatic Organisms* 83: 257–260. <https://doi.org/10.3354/dao02008>
- Gvoždík L 2022 Thermoregulatory opportunity and competition act independently on life-history traits in aquatic ectotherms. *Functional Ecology* 36: 2520–2530. <https://doi.org/10.1111/1365-2435.14134>
- Gvoždík L and Van Damme R 2008 The evolution of thermal performance curves in semi-aquatic newts: Thermal specialists on land and thermal generalists in water? *Journal of Thermal Biology* 33: 395–403. <https://doi.org/10.1016/j.jtherbio.2008.06.004>
- Harrison XA 2014 Using observation-level random effects to model overdispersion in count data in ecology and evolution. *PeerJ* 2: e616. <https://doi.org/10.7717/peerj.616>
- Hecht L 2021 The importance of considering age when quantifying wild animals’ welfare. *Biological Reviews* 96: 2602–2616. <https://doi.org/10.1111/brv.12769>

- IUCN 2023 *The IUCN Red List of Threatened Species. Version 2022-2*. <https://www.iucnredlist.org> (accessed 7 September 2023).
- Kenyon N, Phillott AD and Alford RA 2009 Evaluation of the photographic identification method (PIM) as a tool to identify adult *Litora genimaculata* (Anura: Hylidae). *Herpetological Conservation and Biology* 4(3): 403–410.
- Kinkead KE, Lanham JD and Montanucci RR 2006 Comparison of anesthesia and marking techniques on stress and behavioral responses in two desmognathus salamanders. *Journal of Herpetology* 40: 323–328. [https://doi.org/10.1670/0022-1511\(2006\)40\[323:COAAMT\]2.0.CO;2](https://doi.org/10.1670/0022-1511(2006)40[323:COAAMT]2.0.CO;2)
- Locatelli AG, Ciuti S, Presetnik P, Toffoli R and Teeling E 2019 Long-term monitoring of the effects of weather and marking techniques on body condition in the Kuhl's pipistrelle bat, *Pipistrellus Kuhlii*. *Acta Chiropterologica* 21: 87. <https://doi.org/10.3161/15081109ACC2019.21.1.007>
- Luedtke JA, Chanson J, Neam K, Hobin L, Maciel AO, Catenazzi A, Borzée A, Hamidy A, Aowphol A, Jean A, Sosa-Bartuano Á, Fong GA, De Silva A, Fouquet A, Angulo A, Kidov AA, Muñoz Saravia A, Diesmos AC, Tomi-naga A, Shrestha B, Gratwicke B, Tjaturadi B, Martínez Rivera CC, Vásquez Almazán CR, Señaris C, Chandramouli SR, Striussmann C, Cortez Fernández CF, Azat C, Hoskin CJ, Hilton-Taylor C, Whyte DL, Gower DJ, Olson DH, Cisneros-Heredia DF, Santana DJ, Nagombi E, Najafi-Majd E, Quah ESH, Bolaños F, Xie F, Brusquetti F, Álvarez FS, Andreone F, Glaw F, Castañeda FE, Kraus F, Parra-Olea G, Chaves G, Medina-Rangel GF, González-Durán G, Ortega-Andrade HM, Machado IF, Das I, Dias IR, Urbina-Cardona JN, Crnobrnja-Isailović J, Yang J-H, Jianping J, Wangyal JT, Rowley JJJ, Measey J, Vasudevan K, Chan KO, Gururaja KV, Ovaska K, Warr LC, Canseco-Márquez L, Toledo LF, Díaz LM, Khan MMH, Mee-gaskumbura M, Acevedo ME, Napoli MF, Ponce MA, Vaira M, Lampo M, Yáñez-Muñoz MH, Scherz MD, Rödel M-O, Matsui M, Fildor M, Kusirini MD, Ahmed MF, Rais M, Kouamé NG, García N, Gonwouo NL, Burrowes PA, Imbun PY, Wagner P, Kok PJR, Joglar RL, Auguste RJ, Brandão RA, Ibáñez R, Von May R, Hedges SB, Biju SD, Ganesh SR, Wren S, Das S, Flechas SV, Ashpole SL, Robleto-Hernández SJ, Loader SP, Inchaústegui SJ, Garg S, Phimmachak S, Richards SJ, Slimani T, Osborne-Naikatini T, Abreu-Jardim TPF, Condez TH, De Carvalho TR, Cutajar TP, Pierson TW, Nguyen TQ, Kaya U, Yuan Z, Long B, Langhammer P and Stuart SN 2023 Ongoing declines for the world's amphibians in the face of emerging threats. *Nature*. <https://doi.org/10.1038/s41586-023-06578-4>
- Major T 2020 Marking the un-markable: visible implant elastomer in wild juvenile snakes. *Herpetological Journal*: 173–176. <https://doi.org/10.33256/hj30.3.173176>
- McCarthy MA and Parris KM 2004 Clarifying the effect of toe clipping on frogs with Bayesian statistics. *Journal of Applied Ecology* 41: 780–786. <https://doi.org/10.1111/j.0021-8901.2004.00919.x>
- McEwen BS and Wingfield JC 2003 The concept of allostasis in biology and biomedicine. *Hormones and Behavior* 43: 2–15. [https://doi.org/10.1016/S0018-506X\(02\)00024-7](https://doi.org/10.1016/S0018-506X(02)00024-7)
- Moore F 1984 Stress-induced inhibition of sexual behavior: Corticosterone inhibits courtship behaviors of a male amphibian (*Taricha granulosa*). *Hormones and Behavior* 18: 400–410. [https://doi.org/10.1016/0018-506X\(84\)90026-6](https://doi.org/10.1016/0018-506X(84)90026-6)
- Narayan EJ, Molinia FC, Kindermann C, Cockrem JF and Hero J-M 2011 Urinary corticosterone responses to capture and toe-clipping in the cane toad (*Rhinella marina*) indicate that toe-clipping is a stressor for amphibians. *General and Comparative Endocrinology* 174: 238–245. <https://doi.org/10.1016/j.ygcen.2011.09.004>
- Oropeza Sánchez MT, Sandoval Comte A, García Bañuelos P, Hernández López P and Pineda E 2020 Use of visible implant elastomer and its effect on the survival of an endangered minute salamander. *Animal Biodiversity and Conservation* 43: 187–190.
- Ott JA and Scott DE 1999 Effects of toe-clipping and PIT-tagging on growth and survival in metamorphic *Ambystoma opacum*. *Journal of Herpetology* 33: 344. <https://doi.org/10.2307/1565740>
- Palmer C, Fischer B, Gambourg C, Hampton J and Sandøe P 2023 Wildlife research: Toe clipping. *Wildlife Ethics: The Ethics of Wildlife Management and Conservation* pp 216–229. John Wiley & Sons, Ltd: London, UK.
- Palmer MS, Gaynor KM, Abraham JO and Pringle RM 2022 The role of humans in dynamic landscapes of fear. *Trends in Ecology & Evolution*: S0169534722003330. <https://doi.org/10.1016/j.tree.2022.12.007>
- Passos LF, Garcia G and Young RJ 2017 The tonic immobility test: Do wild and captive golden mantella frogs (*Mantella aurantiaca*) have the same response? *PLOS One* 12: e0181972. <https://doi.org/10.1371/journal.pone.0181972>
- Perry G, Wallace MC, Perry D, Curzer H and Muhlberger P 2011 Toe clipping of amphibians and reptiles: science, ethics, and the law. *Journal of Herpetology* 45: 547–555. <https://doi.org/10.1670/11-037.1>
- Polčák D and Gvoždík L 2014 Should I stay or should I go? The influence of temperature and sex on predator-induced responses in newts. *Animal Behaviour* 89: 79–84. <https://doi.org/10.1016/j.anbehav.2013.12.024>
- R Core Team 2023 R: A Language and Environment for Statistical Computing. R Foundation for Statistical Computing: Vienna, Austria. <https://www.R-project.org/>
- Sapsford SJ, Roznik EA, Alford RA and Schwarzkopf L 2014 Visible implant elastomer marking does not affect short-term movements or survival rates of the treefrog *Litoria rheocola*. *Herpetologica* 70: 23–33. <https://doi.org/10.1655/HERPETOLOGICA-D-13-0004>
- Soryl AA, Moore AJ, Seddon PJ and King MR 2021 The case for welfare biology. *Journal of Agricultural and Environmental Ethics* 34: 7. <https://doi.org/10.1007/s10806-021-09855-2>
- Soulsbury CD, Gray HE, Smith LM, Braithwaite V, Cotter SC, Elwood RW, Wilkinson A and Collins LM 2020 The welfare and ethics of research involving wild animals: A primer. *Methods in Ecology and Evolution* 11: 1164–1181. <https://doi.org/10.1111/2041-210X.13435>
- Storfer A 2003 Amphibian declines: future directions. *Diversity and Distributions* 9: 151–163. <https://doi.org/10.1046/j.1472-4642.2003.00014.x>
- Tinbergen JM, Tinbergen J and Ubels R 2014 Is fitness affected by ring colour? *Ardea* 101: 153–163. <https://doi.org/10.5253/078.101.0210>
- Uglem I, Kristiansen TS, Mejdell CM, Basic D and Mortensen S 2020 Evaluation of large-scale marking methods in farmed salmonids for tracing purposes: Impact on fish welfare. *Reviews in Aquaculture* 12: 600–625. <https://doi.org/10.1111/raq.12342>
- Winandy L, Darnet E and Denoël M 2015 Amphibians forgo aquatic life in response to alien fish introduction. *Animal Behaviour* 109: 209–216. <https://doi.org/10.1016/j.anbehav.2015.08.018>
- Winandy L and Denoël M 2011 The use of visual and automatized behavioral markers to assess methodologies: a study case on PIT-tagging in the Alpine newt. *Behavior Research Methods* 43: 568–576. <https://doi.org/10.3758/s13428-011-0058-z>
- Winandy L and Denoël M 2013 Cues from introduced fish alter shelter use and feeding behaviour in adult alpine newts. *Ethology* 119: 121–129. <https://doi.org/10.1111/eth.12043>
- Wingfield JC 2005 The concept of allostasis: coping with a capricious environment. *Journal of Mammalogy* 86: 248–254. <https://doi.org/10.1644/BHE-004.1>
- Zanette LY and Clinchy M 2019 Ecology of fear. *Current Biology* 29: R309–R313. <https://doi.org/10.1016/j.cub.2019.02.042>