

## Research Article

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# Comparing responses of sensitive and resistant populations of Palmer amaranth (*Amaranthus palmeri*) and waterhemp (*Amaranthus tuberculatus* var. *rudis*) to PPO inhibitors

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**Abstract**

Resistance to protoporphyrinogen oxidase (PPO) inhibitors was first observed in waterhemp in 2001 and was conferred by the deletion of a glycine residue at the 210th position ( $\Delta$ Gly-210) of the PPO enzyme. PPO-inhibitor resistance in Palmer amaranth was first observed in 2011, 10 years later. The objectives of this study were to directly compare PPO inhibitor responses in plants of both species with or without the  $\Delta$ Gly-210 mutation. Using greenhouse experiments, early (EPOST) and late (LPOST) postemergence dose responses using lactofen and fomesafen, and preemergence (PRE) dose responses using fomesafen and flumioxazin, were obtained for a sensitive and resistant population each of waterhemp and Palmer amaranth. An additional spray study confirmed each sensitive population used in the dose responses was representative of its respective species, with regards to PPO-inhibitor sensitivity. When treated at either POST timing, Palmer amaranth was more tolerant than waterhemp, and the  $\Delta$ Gly-210 mutation provided greater resistance in Palmer amaranth (48-fold to >3,440-fold, depending on timing and herbicide) than in waterhemp (31-fold to 123-fold). The level of tolerance in Palmer amaranth was striking; the sensitive Palmer amaranth population treated LPOST survived as well or better than the resistant waterhemp population treated EPOST. With PRE applications, response differences both between species and between resistant and sensitive populations generally were less pronounced, relative to POST applications. Collectively, this research indicates Palmer amaranth tolerance to POST-applied PPO inhibitors could have initially slowed (relative to waterhemp) evolution of resistance to these herbicides, and resistant and sensitive populations of both species are more likely to be effectively controlled with PRE rather than POST applications.

**Introduction**

Protoporphyrinogen oxidase (PPO)-inhibiting herbicides have been used for more than 50 years to control weeds in agronomic crops. Although these herbicides were first used in the 1960s, resistance took a relatively long time to evolve, with the first case of resistance (in waterhemp) not observed until 2001 (Heap 2019). The mechanism of this first case of resistance involves the deletion of a glycine residue at the 210th position ( $\Delta$ Gly-210) of the PPO enzyme (PPO2) (Patzoldt et al. 2006). The region of the sequence spanning position 210 contains two overlapping trinucleotide repeats, and the loss of one of these repeats via a proposed DNA polymerase slippage-like mechanism results in the loss of a glycine codon (Gressel and Levy 2006). Riggins and Tranel (2012) found that Palmer amaranth has the same repeat motif in *PPX2* as waterhemp, suggesting that Palmer amaranth could also evolve resistance to PPO-inhibiting herbicides via the  $\Delta$ Gly-210 mutation.

Since waterhemp first evolved resistance to PPO inhibitors, 12 additional weed species have been found with resistance to PPO-inhibiting herbicides, including Palmer amaranth (Heap 2019). PPO-inhibitor resistance in Palmer amaranth was first observed in 2011, 10 years after resistance to PPO inhibitors in waterhemp was first observed. This Palmer amaranth population was also resistant via the  $\Delta$ Gly-210 mutation (Salas et al. 2016). After identification of the  $\Delta$ Gly-210 mutation in waterhemp and Palmer amaranth, additional mechanisms conferring PPO inhibitor resistance have been identified in these species. These additional mechanisms include mutations conferring other PPO2 amino acid changes (Arg128Gly, Arg128Met, Arg128Ile, and Gly399Ala) as well as enhanced herbicide detoxification (Giacomini et al. 2017; Nie et al. 2019; Rangani et al. 2019; Varanasi et al. 2018).

Given that Palmer amaranth and waterhemp are two very closely related species, it is notable that Palmer amaranth took a decade longer to evolve resistance compared to waterhemp.

Anecdotal evidence from growers indicates that Palmer amaranth is the more difficult species to control with PPO-inhibiting herbicides, and grower reports state that Palmer amaranth is unable to be controlled with PPO inhibitors after it reaches a height of 10 cm (Riar et al. 2013). These pieces of evidence suggest that Palmer amaranth is more tolerant to PPO inhibitors than is waterhemp, which may be why it evolved resistance 10 years later.

The objectives of this study were to characterize the relative levels of resistance to PPO inhibitors in Palmer amaranth and waterhemp conferred specifically by the  $\Delta$ Gly-210 mutation, and to determine the selective advantage of resistance to PPO inhibitors in Palmer amaranth relative to waterhemp.

## Materials and Methods

### Plant Materials

Palmer amaranth populations used in this research included one PPO-inhibitor-resistant (KLPA2-3) and one PPO inhibitor-sensitive (KLPAS) population from Kentucky, and waterhemp populations included one PPO-inhibitor-resistant (CHR14-7) and one PPO inhibitor-sensitive (WHWT) population from Illinois. KLPA2-3 resulted from crossing plants that were homozygous for the  $\Delta$ Gly-210 mutation from a Kentucky population (Lillie et al. 2019), whereas CHR14-7 resulted from crossing plants from the CHR population (Evans et al. 2019) that were homozygous for the  $\Delta$ Gly-210 mutation. Homozygosity for the  $\Delta$ Gly-210 mutation was determined by performing a TaqMan quantitative polymerase chain reaction (qPCR) assay using fluorescent probes designed by Wuerffel et al. (2015) to detect the  $\Delta$ Gly-210 mutation. DNA was extracted from new-leaf tissue using the hexadecyltrimethylammonium bromide method (Doyle and Doyle 1990). The same qPCR assay was performed on a subset of the progeny from the crosses to confirm homozygosity. KLPAS was collected in 2013 from a PPO-inhibitor-sensitive Palmer amaranth plant in Fulton County, Kentucky, and WHWT was made up of multiple wild-type waterhemp accessions collected from various counties in Illinois in 2003.

### Postemergence Dose Responses

Approximately 100 seeds from each population were broadcast into 12 cm by 12 cm plastic greenhouse flat inserts containing a mixture of Sunshine LC1 (Sun Gro Horticulture, Agawam, MA, USA), soil, peat, and torpedo sand (3:1:1:1 by weight) plus 13-13-13 Osmocote fertilizer (The Scotts Company, Marysville, OH, USA). The same soil mixture was used to cover the seeds at a thickness of approximately 2 mm. When they reached the 1- to 2-leaf stage, seedlings were transplanted into 8 cm by 8 cm square plastic pots containing the same mixture of Sunshine LC1, soil, peat, and torpedo sand plus fertilizer. Each pot contained one plant, and all plants were grown in a greenhouse maintained at 30 C/25 C day/night with artificial lighting from metal halide lamps programmed for a 16-hr photoperiod. To quantify the difference in resistance and sensitivity levels at different growth stages, each population was sprayed at a height of 8 to 10 cm (early postemergence [EPOST]) or 13 to 15 cm (late postemergence [LPOST]). Lactofen (Cobra, 240 g ai L<sup>-1</sup>, Valent USA, Walnut Creek, CA, USA) was applied at rates ranging from 0.219 to 6,920 g ha<sup>-1</sup> (1× field rate was 219 g ha<sup>-1</sup>) with a selection of seven rates evenly spaced along a log<sub>10</sub> scale. Similarly, fomesafen (Flexstar, 225 g ai L<sup>-1</sup>, Syngenta US, Wilmington, DE, USA) was

applied at rates ranging from 0.329 to 10,400 g ha<sup>-1</sup> (1× field rate was 329 g ha<sup>-1</sup>). The specific rate applied to each plant depended on whether it was sensitive or resistant to PPO inhibitors and whether the herbicide application timing was EPOST or LPOST. Sensitive plants treated EPOST were sprayed at 0.001× to 1× field rates, sensitive plants treated LPOST were sprayed at 0.00316× to 3.16× field rates, resistant plants treated EPOST were sprayed at 0.01× to 10× field rates, and resistant plants treated LPOST were sprayed at 0.0316× to 31.6× field rates. During the second repetition of the experiment, the range of rates used was shifted up by one rate for all populations and application timings except for the sensitive populations treated EPOST. This occurred to achieve higher levels of control. All herbicide treatments were applied with a compressed-air laboratory spray chamber (DeVries Manufacturing, Hollandale, MN, USA) equipped with an 80° even flat-fan spray nozzle (TeeJet Technologies, Wheaton, IL, USA) calibrated to deliver 187 L ha<sup>-1</sup>. All lactofen spray mixtures contained 1% v/v crop oil concentrate (COC), and all fomesafen spray mixtures contained 1% v/v COC and 1% v/v 32% urea ammonium nitrate (UAN). Adjuvant-only controls were included. Two weeks after treatment (WAT), survivorship ratings were taken on a scale from 0 to 10 where 0 represented plant mortality, and aboveground biomass of each plant was harvested, dried at 37 C for 5 days, and weighed. The dry weights were multiplied by the survivorship ratings to obtain an adjusted dry weight (Guo et al. 2015). To account for differences in plant growth between each population, these values were converted to a percentage of the untreated control using the adjuvant-only control plants from each respective population. Each treatment was replicated five or six times, and the experiment was conducted twice.

### Preemergence Dose Responses

Preemergence (PRE) dose responses were also carried out on these populations to test the differences in resistance and sensitivity to PPO inhibitors at seedling emergence. Fifty-seed weights of each population were planted in 6 cm by 6 cm greenhouse flat inserts filled with a mixture of soil, peat, and torpedo sand (1:1:1 by weight) plus 13-13-13 Osmocote. The same soil mixture was used to cover the seeds at a thickness of 1 to 2 mm, and herbicides were applied immediately after planting. Fomesafen was applied at rates ranging from 0.329 to 3,290 g ha<sup>-1</sup> with a selection of seven rates evenly spaced along a log<sub>10</sub> scale. Flumioxazin (Valor, 0.51 g ai g<sup>-1</sup>, Valent USA, Walnut Creek, CA, USA) was applied at rates ranging from 0.107 to 338 g ha<sup>-1</sup> (1× field rate was 107 g ha<sup>-1</sup>). The sensitive populations were sprayed from 0.001× to 1× field rates of both herbicides, and the resistant populations were sprayed from 0.01× to 10× field rates of fomesafen and 0.00316× to 3.16× field rates of flumioxazin. Water-only controls were included. Following herbicide application, overhead water was gently applied to leach the herbicide through the soil profile (simulating approximately 50 mm rainfall), and soil moisture was maintained with subirrigation for the remainder of the experiment. Ten days after treatment (DAT), emergence counts were taken. Seedlings were considered fully emerged when the adaxial leaf surfaces of the cotyledons were no longer touching each other (Wuerffel et al. 2015). Emergence counts were converted to a percentage of the untreated control using the water-only control counts from each respective population. Each treatment was replicated six times, and the experiment was conducted twice.

### Sensitive Spray Study

An additional spray study was carried out to determine whether the sensitive populations used in the dose-response experiments were representative of sensitive populations of each species. Six wild-type populations each of Palmer amaranth and waterhemp from various states were used. The wild-type waterhemp populations tested were from Illinois, Iowa, Kansas, Nebraska, Indiana, and Ohio. The Illinois population was the same as that used in the dose-response experiments, the Ohio population was collected during the 2016 growing season as described in Murphy (2018), and the remaining populations were obtained from the Germplasm Resources Information Network (GRIN). The sensitive Palmer amaranth populations tested were from Arizona, Arkansas, Illinois, Kentucky, Missouri, and Oklahoma. The population from Kentucky was the same population used in the dose-response experiments, the Arizona and Oklahoma populations were from an in-house collection, and the remaining populations were collected in 2009 from the margins of agricultural fields as explained in Davis et al. (2015).

Experimental procedures (e.g., growing plants, herbicide treatments, and plant evaluation) were identical to those described above for EPOST treatments, with the exception of the herbicide rates used. For this particular study, lactofen was applied at 0, 2.19, 11.0, or 110.0 g ha<sup>-1</sup> for Palmer amaranth and 0, 0.175, 2.19, or 11.0 g ha<sup>-1</sup> for waterhemp, whereas fomesafen was applied at 0, 1.04, 10.4, or 104 g ha<sup>-1</sup> for Palmer amaranth and 0, 0.330, 10.4, or 104 g ha<sup>-1</sup> for waterhemp. Each treatment was replicated five or six times, and the experiment was conducted twice. The study was conducted as a randomized complete block design (RCBD) in which the blocking factor was experimental run.

### Statistical Analysis

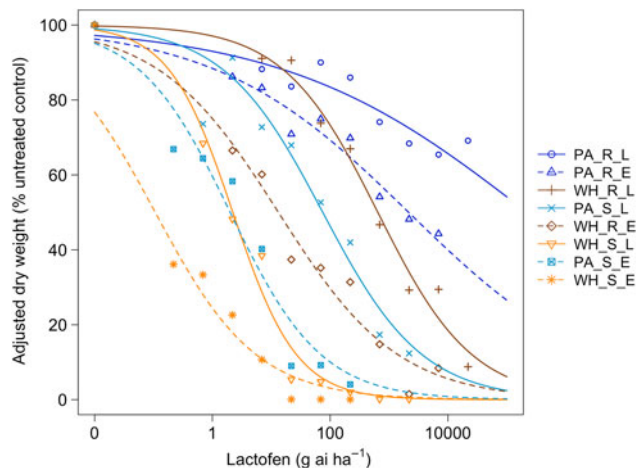
Adjusted dry weights (POST dose response) and emergence counts (PRE dose response) were analyzed in R software v. 1.0.143 using the dose response curve (drc) package (Ritz et al. 2015). The four-parameter log-logistic function used to generate dose-response curves is expressed as:

$$y = c + \frac{d - c}{1 + \exp(b(\log(x) - \log(ED_{50})))} \quad [1]$$

where  $y$  is the adjusted dry weight or emergence count,  $b$  is the slope of the curve,  $c$  is the lower asymptote,  $d$  is the upper asymptote,  $x$  is the herbicide rate, and  $ED_{50}$  is the herbicide rate required to reduce the response halfway between  $d$  and  $c$ . Homogeneity of variance was achieved through a square root transformation of the data. To determine if the fitted curves were different from one another,  $ED_{50}$  values were calculated and compared. The  $ED_{50}$  value and its standard error associated with each curve were obtained from the fit of the log-logistic model, and these values were used to calculate the 95% confidence intervals of the  $ED_{50}$  values from each fitted curve.

For the sensitive spray study, the data were separated into four groups and analyzed separately: waterhemp treated with lactofen, waterhemp treated with fomesafen, Palmer amaranth treated with lactofen, and Palmer amaranth treated with fomesafen. Adjusted dry weights were calculated as described above. For each set of data, an ANOVA was carried out using the following model:

$$y_{ijkl} = \mu + R_i + D_j + P_k + DP_{jk} + \varepsilon_{ijkl} \quad [2]$$



**Figure 1.** Dose responses of waterhemp (WH\_X\_X) and Palmer amaranth (PA\_X\_X) populations homozygous for (XX\_R\_X) or lacking (XX\_S\_X) the Gly210 PPO deletion treated with lactofen at early (XX\_X\_E) and late (XX\_X\_L) application stages. Dose-response curves were fitted using a four-parameter, log-logistic function in R software.

where  $y_{ijkl}$  is the adjusted dry weight of the plant under observation,  $\mu$  is the grand population mean,  $R_i$  is the random effect of the  $i^{\text{th}}$  experimental run,  $D_j$  is the random effect of the  $j^{\text{th}}$  herbicide dose,  $P_k$  is the random effect of the  $k^{\text{th}}$  population,  $DP_{jk}$  is the random interaction of the  $j^{\text{th}}$  herbicide dose and the  $k^{\text{th}}$  population, and  $\varepsilon_{ijkl}$  is the random error term, NID(0,  $\sigma_e^2$ ). Data were analyzed using the PROC MIXED procedure using the TYPE 3 model in SAS 9.4 (Statistical Analysis Software, Inc., Cary, NC, USA). The assumption of normality was analyzed by conducting a Shapiro-Wilks normality test on the residuals using PROC UNIVARIATE, and the assumption of homogeneous variances was analyzed using the Browne and Forsythe test in the MEANS option of PROC GLM. Because the residuals were not normally distributed, the Palmer amaranth data were transformed via the cube root, and the waterhemp data were transformed via the square root. Due to a lack of homogeneity of variance after transformation, separate variance groups were set for each data set.

## Results and Discussion

### Postemergence Dose Responses

Not surprisingly, the populations of both species sprayed at LPOST had significantly higher  $ED_{50}$  values than their respective populations sprayed at EPOST for both lactofen (Figure 1 and Table 1) and fomesafen (Figure 2 and Table 2). The sensitive waterhemp sprayed at EPOST had the lowest  $ED_{50}$  value for both lactofen (Table 1) and fomesafen (Table 2) dose responses. These results aligned with those reported by Hager et al. (2003), which found control of common waterhemp was greater after an EPOST timing than after a later POST timing. Results also coincided with those of the study performed by Wuerffel et al. (2015), which demonstrated that the response of resistant biotypes of waterhemp to PPO-inhibiting herbicides depends on the target plant growth stage.

When sprayed with lactofen, the resistant Palmer amaranth population was not sufficiently controlled at the highest dose for either application timing (Figure 1 and Table 1). Additionally, the  $ED_{50}$  value of the sensitive Palmer amaranth sprayed EPOST was not significantly different than the  $ED_{50}$  value of the sensitive waterhemp sprayed LPOST (Table 1). When sprayed LPOST, the sensitive Palmer amaranth population was

**Table 1.** Lactofen ED<sub>50</sub> values and relative sensitivities of each biotype sprayed early POST and late POST.

Biotype <sup>a</sup>	ED <sub>50</sub> (g ha <sup>-1</sup> ) <sup>b</sup>	ED <sub>50</sub> (x) <sup>b</sup>	Relative sensitivity <sup>c</sup>
PA_R_L	>21,900	>100	>205,000
PA_R_E	>6,920	>31.6	>64,800
WH_R_L	645 a	2.94 a	6,020
PA_S_L	74.9 b	0.342 b	701
WH_R_E	13.2 c	0.0604 c	124
WH_S_L	2.23 d	0.0102 d	20.9
PA_S_E	2.01 d	0.00916 d	18.8
WH_S_E	0.107 e	0.000488 e	1.00

<sup>a</sup>Abbreviations: PA\_X\_X, Palmer amaranth; WH\_X\_X, waterhemp; XX\_R\_X, homozygous for the Gly210 PPO deletion; XX\_S\_X, lacking the Gly210 PPO deletion; XX\_X\_E, early POST application; XX\_X\_L, late POST application.

<sup>b</sup>Difference in ED<sub>50</sub> values determined using 95% confidence intervals.

<sup>c</sup>Relative sensitivity values calculated relative to WH\_S\_E.

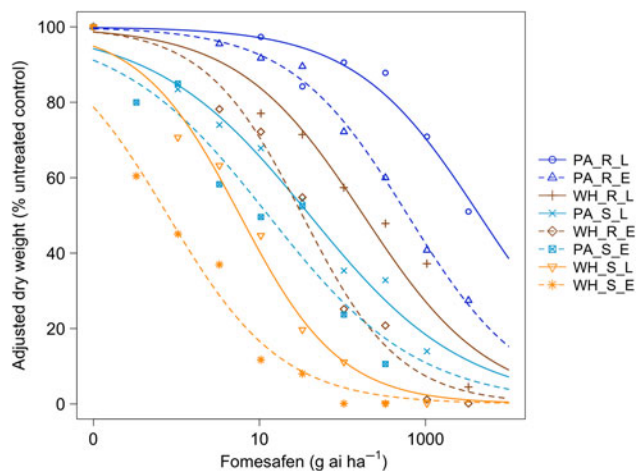
**Table 2.** Fomesafen ED<sub>50</sub> values and relative sensitivities of each biotype sprayed early POST and late POST.

Biotype <sup>a</sup>	ED <sub>50</sub> (g ha <sup>-1</sup> ) <sup>b</sup>	ED <sub>50</sub> (x) <sup>b</sup>	Relative sensitivity <sup>c</sup>
PA_R_L	4480 a	13.6 a	5670
PA_R_E	614 b	1.87 b	779
WH_R_L	180 c	0.546 c	228
PA_S_L	40.5 d	0.123 d	51.3
WH_R_E	32.9 d	0.100 d	41.7
PA_S_E	12.8 e	0.0389 e	16.2
WH_S_L	5.85 e	0.0178 e	7.42
WH_S_E	0.789 f	0.00240 f	1.00

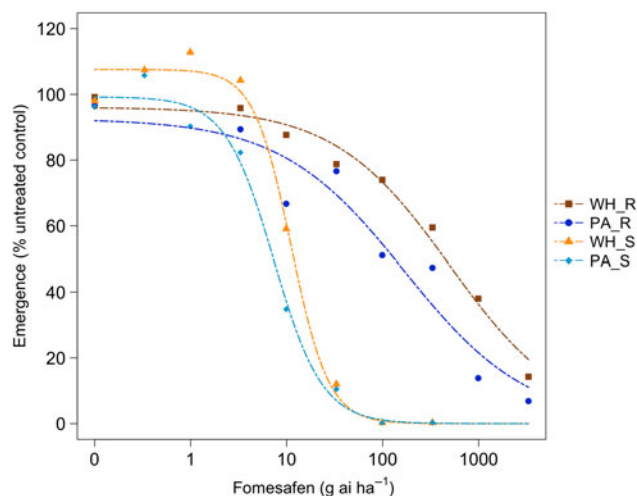
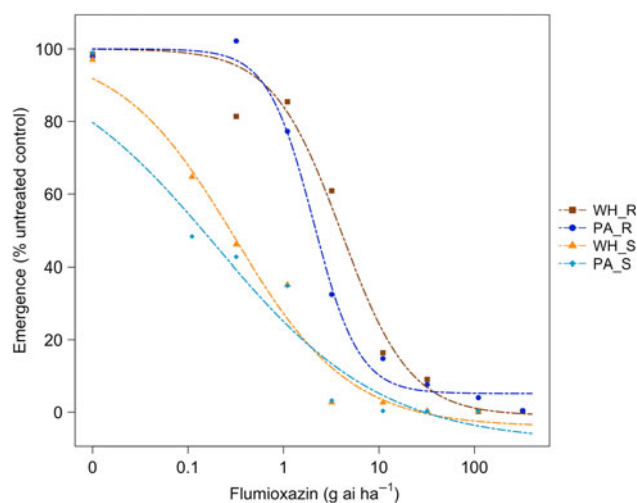
<sup>a</sup>Abbreviations: PA\_X\_X, Palmer amaranth; WH\_X\_X, waterhemp; XX\_R\_X, homozygous for the Gly210 PPO deletion; XX\_S\_X, lacking the Gly210 PPO deletion; XX\_X\_E, early POST application; XX\_X\_L, late POST application.

<sup>b</sup>Difference in ED<sub>50</sub> values determined using 95% confidence intervals.

<sup>c</sup>Relative sensitivity values calculated relative to WH\_S\_E.

**Figure 2.** Dose responses of waterhemp (WH\_X\_X) and Palmer amaranth (PA\_X\_X) populations homozygous for (XX\_R\_X) or lacking (XX\_S\_X) the Gly210 PPO deletion treated with fomesafen at early (XX\_X\_E) and late (XX\_X\_L) application stages. Dose-response curves were fitted using a four-parameter, log-logistic function in R software.

significantly less sensitive than the resistant waterhemp population when sprayed EPOST (Table 1). Furthermore, when sprayed LPOST with fomesafen, the ED<sub>50</sub> value of the sensitive Palmer amaranth was not significantly different than the ED<sub>50</sub> value of

**Figure 3.** Dose responses of waterhemp (WH\_X) and Palmer amaranth (PA\_X) populations homozygous for (XX\_R) or lacking (XX\_S) the Gly210 PPO deletion treated with fomesafen at the PRE application stage. Dose-response curves were fitted using a four-parameter, log-logistic function in R software.**Figure 4.** Dose responses of waterhemp (WH\_X) and Palmer amaranth (PA\_X) populations homozygous for (XX\_R) or lacking (XX\_S) the Gly210 PPO deletion treated with flumioxazin at the PRE application stage. Dose-response curves were fitted using a four-parameter, log-logistic function in R software.

the resistant waterhemp sprayed EPOST (Table 2). These results suggest that Palmer amaranth does not necessarily need a resistance mechanism against PPO inhibitors to survive a PPO-inhibitor application; rather, it is enough just to be taller and have more biomass.

The resistant to sensitive (R/S) ratios for waterhemp treated with fomesafen POST were similar to the R/S ratio that Wuerffel et al. (2015) reported. They calculated an R/S ratio of 38, which lies between the R/S ratios of 30.8 and 41.7 obtained in this study for waterhemp treated with fomesafen LPOST and EPOST, respectively (Table 5). The waterhemp population used in the Wuerffel et al. study was a field population that contained the ΔGly-210 mutation. For waterhemp treated with lactofen, Patzoldt et al. (2006) calculated an R/S ratio of 53 in a waterhemp population that was homozygous for ΔGly-210, which is similar to the R/S ratio of 48.9 obtained in this study for the LPOST

**Table 3.** Fomesafen ED<sub>50</sub> values and relative sensitivities of each biotype sprayed PRE.

Biotype <sup>a</sup>	ED <sub>50</sub>	ED <sub>50</sub>	Relative sensitivity <sup>c</sup>
	(g ha <sup>-1</sup> ) <sup>b</sup>	(x) <sup>b</sup>	
WH_R	500 a	1.52 a	67.9
PA_R	169 a	0.514 a	23.0
WH_S	11.2 b	0.0340 b	1.52
PA_S	7.36 b	0.0224 b	1.00

<sup>a</sup>Abbreviations: PA\_X, Palmer amaranth; WH\_X, waterhemp; XX\_R, homozygous for the Gly210 PPO deletion; XX\_X, lacking the Gly210 PPO deletion.

<sup>b</sup>Difference in ED<sub>50</sub> values determined using 95% confidence intervals.

<sup>c</sup>Relative sensitivity values calculated relative to PA\_S.

**Table 4.** Flumioxazin ED<sub>50</sub> values and relative sensitivities of each biotype sprayed PRE.

Biotype <sup>a</sup>	ED <sub>50</sub>	ED <sub>50</sub>	Relative sensitivity <sup>c</sup>
	(g ha <sup>-1</sup> ) <sup>b</sup>	(x) <sup>b</sup>	
WH_R	4.01 a	0.0375 a	20.9
PA_R	2.04 b	0.0191 b	10.6
WH_S	0.305 c	0.00285 c	1.59
PA_S	0.192 c	0.00179 c	1.00

<sup>a</sup>Abbreviations: PA\_X, Palmer amaranth; WH\_X, waterhemp; XX\_R, homozygous for the Gly210 PPO deletion; XX\_X, lacking the Gly210 PPO deletion.

<sup>b</sup>Difference in ED<sub>50</sub> values determined using 95% confidence intervals.

<sup>c</sup>Relative sensitivity values calculated relative to PA\_S.

application timing (Table 5). Both the Wuerffel et al. and Patzold et al. studies followed herbicide rate structures, COC rates, and application heights that were similar to this study. Salas et al. (2016) obtained R/S ratios ranging between 6 and 21 for Palmer amaranth treated with fomesafen POST. These values were much smaller than the R/S ratios of 48.0 and 111 calculated in the current study for Palmer amaranth treated with fomesafen EPOST and LPOST, respectively, but the study by Salas et al. followed a different herbicide rate structure, and their herbicide mixtures included 0.5% nonionic surfactant, which may account for this difference. Additionally, the Palmer amaranth populations used in that study were a mixture of plants both homozygous and heterozygous for the ΔGly-210 mutation.

### Preemergence Dose Responses

There were similarities and differences in the patterns of response in both species when sprayed PRE (Figure 3 and Figure 4) compared with when they were sprayed POST. When sprayed with fomesafen PRE, the resistant waterhemp demonstrated the same level of resistance as the resistant Palmer amaranth (Table 3). The sensitive waterhemp was also just as sensitive as the sensitive Palmer amaranth. The resistant populations of both species were significantly less sensitive than the sensitive populations for both herbicides (Table 3 and Table 4). When sprayed with flumioxazin PRE, the resistant waterhemp was more resistant than the resistant Palmer amaranth, but the sensitive waterhemp exhibited the same level of sensitivity as the sensitive Palmer amaranth (Table 4). These results were similar to those of the POST dose responses in that the resistant populations had significantly higher ED<sub>50</sub> values than the sensitive populations. However, the flumioxazin PRE results showed the resistant waterhemp population was more resistant than the resistant Palmer amaranth population, a finding that differed from the POST results. Overall, the PRE dose response study indicated that there were fewer differences in

**Table 5.** Resistant to sensitive ratios of Palmer amaranth and waterhemp sprayed EPOST, LPOST, or PRE with lactofen, fomesafen, or flumioxazin.

Species and application timing <sup>b</sup>	R/S <sup>a</sup>		
	Lactofen	Fomesafen	Flumioxazin
Waterhemp PRE	–	44.6	13.4
Waterhemp EPOST	123	41.7	–
Waterhemp LPOST	48.9	30.8	–
Palmer amaranth PRE	–	22.9	10.6
Palmer amaranth EPOST	>3,440	48.0	–
Palmer amaranth LPOST	>292	111	–

<sup>a</sup>R/S ratios not calculated are represented by (–).

<sup>b</sup>Abbreviations: EPOST, early postemergence; LPOST, late postemergence; PRE, preemergence; R/S, resistant to sensitive ratio.

resistance and sensitivity levels between waterhemp and Palmer amaranth when sprayed PRE than when sprayed EPOST or LPOST.

The R/S ratio of 44.8 for waterhemp treated with fomesafen PRE (Table 5) was similar to that reported by Wuerffel et al. (2015), who calculated an R/S ratio of 33 in a field population of waterhemp that was resistant via the ΔGly-210 mutation. Falk et al. (2006) reported an R/S ratio of 2.5 for fomesafen applied PRE to an uncharacterized population of waterhemp, which was much smaller than the R/S ratio of 44.8 obtained in this study. Umphres et al. (2018) reported an R/S ratio of 5.2 for a field population of resistant Palmer amaranth sprayed with fomesafen, which was much smaller than the R/S ratio of 22.9 we report in this study. However, the resistance mechanism in population of plants in the Umphres et al. study was not characterized. Both studies followed different herbicide rate structures than the current study did. For flumioxazin, Umphres et al. (2018) reported an R/S ratio of 4.7, which is not far from the R/S ratio of 10.6 calculated in this study.

A study by Vila-Aiub et al. (2018) found that an increase in size from the seedling stage of glyphosate-resistant johnsongrass [*Sorghum halepense* (L.) Pers.] resulted in increasing R/S ratios. The R/S ratios of the fomesafen LPOST, EPOST, and PRE dose responses (Table 5) reflected this same pattern occurring in Palmer amaranth but not in waterhemp. For waterhemp, the R/S ratios were more likely to decrease rather than increase across these timings. When treated with lactofen, the R/S ratios for waterhemp decreased from EPOST to LPOST. Why the R/S ratios exhibited different trends across application timings for the two species is not clear. Perhaps developmentally regulated differences exist in tolerance mechanisms (e.g., differential expression of a gene encoding an enzyme that contributes to herbicide metabolism) that influence the R/S ratio.

### Sensitive Spray Study

The ANOVA results from the sensitive spray study showed that the population main effect and the interaction between population and herbicide dose were insignificant for all experiments, indicating that the means of all populations were not statistically different, both overall and at each dose of herbicide applied (Table 6). This suggests that the populations used as the sensitive controls for each species in the dose responses were representative of wild-type populations for the corresponding species. The block effect, experimental run, was highly insignificant for the analysis of the waterhemp populations applied with lactofen. The block effect was significant in the remainder of analyses, which is likely due to different experimental conditions such as greenhouse

**Table 6.** One-way analysis of variance tables comparing the effect of dose of lactofen or fomesafen on six different wild-type populations of either Palmer amaranth or waterhemp, blocked by experimental run.

Experiment	Effect	Degrees of freedom	F-value	p value
Palmer amaranth treated with lactofen	Run	1	35.45	<0.05
	Dose	3	287.85	<0.05
	Population	5	0.81	0.5618
	Dose*Population	15	1.46	0.1225
Palmer amaranth treated with fomesafen	Run	1	23.44	<0.05
	Dose	3	343.51	<0.05
	Population	5	2.02	0.1342
	Dose*Population	15	1.48	0.1146
Waterhemp treated with lactofen	Run	1	0.26	0.6095
	Dose	3	239.18	<0.05
	Population	5	0.85	0.5196
	Dose*Population	15	1.33	0.1845
Waterhemp treated with fomesafen	Run	1	3.84	0.0513
	Dose	3	368.05	<0.05
	Population	5	2.76	0.0579
	Dose*Population	15	1.47	0.1184

temperature, because each run of the experiment was temporally separated. The effect of dose was significant for all analyses because significant differences in plant response existed between a 0X dose and all other doses. Because the sensitive populations used in the dose-response experiments are representative of wild-type populations, it can be assumed that the results of the dose-response experiments are indicative of each species as a whole.

Collectively, the results reported herein indicate that Palmer amaranth is more tolerant to PPO inhibitors than waterhemp when treated POST. This could be why Palmer amaranth took a decade longer to evolve resistance to PPO inhibitors compared with that of waterhemp (i.e., because Palmer amaranth is naturally more tolerant to POST application of these herbicides, there is less of a selective advantage for it to evolve resistance to PPO inhibitors). If a Palmer amaranth plant is sufficiently large at the time of application of a PPO inhibitor, it does not need a resistance mechanism to survive, and therefore, it is not under as strong of a selection pressure for resistance. This could be attributed to its high growth and photosynthetic rates (Ehleringer 1983; Horak and Loughin 2000) or various tolerance mechanisms (e.g., greater ability to metabolize the herbicide). Aside from herbicide-resistance evolution, understanding the mechanism underlying this natural tolerance could give insight into herbicide tolerance mechanisms that could be utilized in crops.

Another factor that could have contributed to waterhemp having evolved resistance to PPO inhibitors before Palmer amaranth did is that initially, these herbicides may have demonstrated a greater selection for waterhemp than there was for Palmer amaranth. In 1994, PPO-inhibitor use was highest primarily in midwestern states (Dayan et al. 2018), wherein waterhemp was likely being targeted, because this species was more prevalent in the Midwest than Palmer amaranth was at that time. The total usage of PPO inhibitors in the United States during the early 1990s was relatively high (Dayan et al. 2018), as this time period was just before the introduction of glyphosate-resistant crops. After these crops were introduced, PPO-inhibitor use declined, and did not rebound until after glyphosate-resistant weeds became common. It was not until this second wave of high PPO-inhibitor use that resistance to these herbicides was observed in Palmer amaranth. A study by Webster and Nichols (2012) evaluated

how weed species have changed over the 14-year period between 1994 and 2008 in the southern United States. In cotton and soybean systems, in which PPO inhibitors are typically used, Palmer amaranth went from being the 10th and 23rd most troublesome weed species, respectively, to the first and second most troublesome weed species, respectively. Perhaps the slower evolution of PPO-inhibitor resistance in Palmer amaranth relative to waterhemp is due simply to a higher number of individuals of the latter species being exposed to these herbicides before the era of glyphosate-resistant crops.

Regardless of the evolutionary history of resistance to PPO inhibitors in these two species, our results highlight the importance of being able to identify Palmer amaranth and waterhemp growing in the field to inform timely herbicide application. Based on our results, it is more important to make early applications of a PPO inhibitor when Palmer amaranth is present in the field. A PRE application would be the best way to control both species, because both have the same level of resistance or sensitivity to these herbicides at this application stage. Additionally, these results indicate that when making a POST application of a PPO inhibitor, it may be difficult to determine whether Palmer amaranth in the field is resistant or not based only on survival.

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## Reference

- Davis AS, Schutte BJ, Hager AG, Young BG (2015) Palmer amaranth (*Amaranthus palmeri*) damage niche in Illinois soybean is seed limited. *Weed Sci* 63:658–668
- Dayan FE, Barker A, Tranel PJ (2018) Origins and structure of chloroplastic and mitochondrial plant protoporphyrinogen oxidases: Implications for the evolution of herbicide resistance. *Pest Manag Sci* 74:2226–2234
- Doyle JJ, Doyle JL (1990) Isolation of plant DNA from fresh tissue. *Focus* 12: 13–15
- Ehleringer J (1983) Ecophysiology of *Amaranthus palmeri*, a sonoran desert summer annual. *Oecologia* 57:107–112
- Evans CM, Strom SA, Riechers DE, Davis AS, Tranel PJ, Hager AG (2019) Characterization of a waterhemp (*Amaranthus tuberculatus*) population from Illinois resistant to herbicides from five site-of-action groups. *Weed Technol* 33:400–410
- Falk JS, Shoup DE, Al-Khatib K, Peterson DE (2006) Protox-resistant common waterhemp (*Amaranthus rudis*) response to herbicides applied at different growth stages. *Weed Sci* 54:793–799
- Giacomini DA, Umphres AM, Nie H, Mueller TC, Steckel LE, Young BG, Scott RC, Tranel PJ (2017) Two new PPX2 mutations associated with resistance to PPO-inhibiting herbicides in *Amaranthus palmeri*. *Pest Manag Sci* 73: 1559–1563
- Gressel J, Levy AA (2006) Agriculture: The selector of improbable mutations. *Proc Natl Acad Sci USA* 103:12215–12216
- Guo J, Riggins CW, Hausman NE, Hager AG, Riechers DE, Davis AS, Tranel PJ (2015) Nontarget-site resistance to ALS inhibitors in waterhemp (*Amaranthus tuberculatus*). *Weed Sci* 63:399–407
- Hager AG, Wax LM, Bollero GA, Stoller EW (2003) Influence of diphenylether herbicide application rate and timing on common waterhemp (*Amaranthus rudis*) control in soybean (*Glycine max*). *Weed Technol* 17:14–20
- Heap IM (2019) International Survey of Herbicide Resistant Weeds. [www.weedscience.org](http://www.weedscience.org). Accessed March 21, 2019
- Horak MJ, Loughin TM (2000) Growth analysis of four *Amaranthus* species. *Weed Sci* 48:347–355

- Lillie KJ, Giacomini DA, Green JD, Tranel PJ (2019) Coevolution of resistance to PPO inhibitors in waterhemp (*Amaranthus tuberculatus*) and Palmer amaranth (*Amaranthus palmeri*). *Weed Sci* 67:521–526
- Murphy BP (2018) Discovery, surveillance, and management of herbicide-resistant *Amaranthus spp.* University of Illinois at Urbana-Champaign. 120 p
- Nie H, Mansfield BC, Harre NT, Young JM, Steppig NR, Young BG (2019) Investigating target-site resistance mechanism to the PPO-inhibiting herbicide fomesafen in waterhemp and interspecific hybridization of *Amaranthus* species using next generation sequencing. *Pest Manag Sci*, 10.1002/ps.5445
- Patzoldt WL, Hager AG, McCormick JS, Tranel PJ (2006) A codon deletion confers resistance to herbicides inhibiting protoporphyrinogen oxidase. *Proc Natl Acad Sci USA* 103:12329–12334
- Rangani G, Salas-Perez RA, Aponte RA, Knapp M, Craig IR, Mietzner T, Langaro AC, Noguera MM, Porri A, Roma-Burgos, N (2019) A novel single-site mutation in the catalytic domain of protoporphyrinogen oxidase IX (PPO) confers resistance to PPO-inhibiting herbicides. *Front Plant Sci* 10:568
- Riar DS, Norsworthy JK, Steckel LE, Stephenson DO, Eubank TW, Scott RC (2013) Assessment of weed management practices and problem weeds in the Midsouth United States—soybean: A consultant's perspective. *Weed Technol* 27:612–622
- Riggins CW, Tranel PJ (2012) Will the *Amaranthus tuberculatus* resistance mechanism to PPO-inhibiting herbicides evolve in other *Amaranthus species*? *Int J Agron* 2012:1–7
- Ritz C, Baty F, Streibig JC, Gerhard D (2015) Dose-response analysis using R. *PLOS ONE* 10:e0146021
- Salas RA, Burgos NR, Tranel PJ, Singh S, Glasgow L, Scott RC, Nichols RL (2016) Resistance to PPO-inhibiting herbicide in Palmer amaranth from Arkansas. *Pest Manag Sci* 72:864–869
- Umphres AM, Steckel LE, Mueller TC (2018) Control of protoporphyrinogen oxidase inhibiting herbicide resistant and susceptible Palmer amaranth (*Amaranthus palmeri*) with soil-applied protoporphyrinogen oxidase-inhibiting herbicides. *Weed Technol* 32:95–100
- Varanasi VK, Brabham C, Norsworthy JK (2018) Confirmation and characterization of non-target site resistance to fomesafen in Palmer amaranth (*Amaranthus palmeri*). *Weed Sci* 66:702–709
- Vila-Aiub M, Casas C, Gundel PE (2018) The role of plant size in the selection of glyphosate resistance in *Sorghum halepense*. *Pest Manag Sci* 74:2460–2467
- Webster TM, Nichols RL (2012) Changes in the prevalence of weed species in the major agronomic crops of the southern United States: 1994/1995 to 2008/2009. *Weed Sci* 60:145–157
- Wuerffel RJ, Young JM, Matthews JL, Young BG (2015) Characterization of PPO-inhibitor-resistant waterhemp (*Amaranthus tuberculatus*) response to soil-applied PPO-inhibiting herbicides. *Weed Sci* 63:511–521