

## Research Article

**Cite this article:** Baruah S, Bharti T, Rajasekhar D, Tatiparthi H, Lyngdoh Nongbri E, Sen D (2024). Genetic characterization and evaluation of pigmented maize (*Zea mays* L.) landraces of the North East Hill Region of India—an established centre of maize diversity. *Plant Genetic Resources: Characterization and Utilization* **22**, 223–233. <https://doi.org/10.1017/S1479262124000182>

Received: 22 February 2023

Revised: 4 March 2024

Accepted: 5 March 2024

First published online: 30 April 2024

### Keywords:


coloured maize; genetic characterization; genetic diversity; landraces; population structuring

### Corresponding author:

Devyani Sen;

Email: [devyani.sen@gmail.com](mailto:devyani.sen@gmail.com)

# Genetic characterization and evaluation of pigmented maize (*Zea mays* L.) landraces of the North East Hill Region of India—an established centre of maize diversity

Sristishila Baruah, Thounaojam Bharti, Duddukur Rajasekhar, Harshavardhan Tatiparthi, Ernieca Lyngdoh Nongbri and Devyani Sen 

School of Crop Improvement, College of Post Graduate Studies in Agricultural Sciences, Central Agricultural University (Imphal), Umiam, Meghalaya, India

## Abstract

The North East Hill Region (NEHR) of India is home to diverse maize landraces including pigmented accessions rich in antioxidants and nutritional properties. The present study attempted to characterize a representative collection of this mostly unexplored diversity. Altogether eighty-three local maize landraces from the seven hill states of the NEHR were studied with special emphasis on pigmentation diversity. For the morphological traits, a significant ANOVA indicated the presence of substantial genetic variability for which selection would be fruitful. A number of these accessions were found to have traits that help cope with moisture stress, improve stalk strength and optimize photosynthesis. Principal component analysis studies for the yield attributing traits indicated that ear weight was most variable. The bleaching/histological studies confirmed that anthocyanin pigment when present was always restricted to the aleurone layer of the kernels, typical of blue maize. Quantitative analysis for kernel anthocyanin/phlobaphene content also revealed genetic differences among the accessions. Genetic analysis using the model-based STRUCTURE indicated significant population structuring among the accessions. Specifically, for the pigmentation diversity studies both principal coordinate analysis and neighbour joining methods revealed near concurrent population structuring due in part to the high differentiation of seven of the twenty-one pigmentation specific loci studied. The results obtained provide comprehensive evidence of a significant amount of genetic differentiation among the landraces under study. Landraces are valuable reservoirs of favourable alleles for which selection can be made and, as in this study, identify accessions for breeding maize with enhanced levels of beneficial secondary metabolites.

## Introduction

Maize (*Zea mays* L.) which is currently the leading cereal crop in terms of production (FAO, 2019) originated from teosinte approximately 9000 years ago in the Balsas region of south-western Mexico (Bennetzen *et al.*, 2001; Stitzer and Ross-Ibarra, 2018; Bruggeman *et al.*, 2020). Following its diversification through combined interventions of man and nature in the Mesoamerican region, maize spread to the other continents chiefly via trade routes. Today abundant genetic diversity exists for maize – the primary reason being its unique reproductive structure which allows extensive outcrossing in natural settings (Hallauer and Carena, 2009). Thus, secondary centres of crop diversity have emerged globally in different agro-climatic zones generating even higher levels of variability especially in the form of maize landraces (Matsuoka *et al.*, 2002; Flint-Garcia, 2013; Andorf *et al.*, 2019).

In India, the North Eastern Hill Region (NEHR) a recognized Asiatic centre of maize diversity (Sharma and Brahmi, 2011) is home to various races of maize (Singh, 1977), including the distinct primitive popcorn ‘Sikkim Primitive’. Based on morphological, cytogenetic and isozyme comparisons between the Sikkim Primitive and the ancient indigenous maize variety ‘Nal-tel’ of Mexico (Dhawan, 1964; Singh, 1977; Sachan and Sarkar, 1982; Bhat *et al.*, 1998), it has been suggested that corn cultivation in the NEHR could have pre-dated the Columbian era. Variants of the Sikkim Primitive having evolved over time due to both natural and human-driven selection are presently distributed across the foothills of North Eastern Himalayan region (Sachan and Sarkar, 1986; Prasanna, 2012). The hill regions of these foothills dominated by different indigenous tribal population have for generations ensured the conservation of these maize landraces as part of their cultural tradition (Singh *et al.*, 2016).

This extensive collection of diverse maize finds recognition as a valuable source of desirable genes (Sharma *et al.*, 2010; Prasanna, 2012; Sanjenbam *et al.*, 2018; Rathod *et al.*, 2019) and the pigmented landraces are particularly important from a nutritional perspective.



Pigmented kernels get their colour from synthesis of secondary metabolites such as anthocyanin and phlobaphene. Kernels with anthocyanin deposition in the pericarp and/or aleurone layers have high antioxidant and antiradical properties and the health benefits include anti-mutagenic, anti-carcinogenic and oestrogenic effects (Adom and Liu, 2002; de Pascual-Teresa and Sanchez-Ballesta, 2008; Zilic *et al.*, 2012) to name a few. Phlobaphene deposition has also been reported to prevent mycotoxin contamination of maize kernels post kernel development (Pilu *et al.*, 2011). Additionally, since majority of anthocyanin found in maize kernels are acylated, they are more stable as natural food colourants (Chatham *et al.*, 2019; Paulsmeyer and Juvik, 2021).

A growing public focus on healthy eating reshaping the purchasing behaviour of the general consumer (Petrescu *et al.*, 2020) therefore aligns well with the nutritional potential of pigmented maize cultivars (hereafter also referred to as coloured maize). However, previous studies (Ryu *et al.*, 2013; Lago *et al.*, 2015; Paauw *et al.*, 2019) have shown that majority of the coloured maize are landraces that have remained underutilized confined to their areas of origin. If we were to exploit this resource and benefit from its diversity, it is essential to have first a preliminary understanding of it. This is because, direct evaluation of maize landraces is a more effective method of incorporating favourable alleles into commercial breeding programmes (Gorjanc *et al.*, 2016) and organizing this information along scientific lines leads to better planning and execution (Andorf *et al.*, 2019).

When characterizing local germplasm besides phenotyping of visible morphological differences, molecular studies provide insights into the underlying genetic variation and associations of the material under study. With maize, the expectation is that landraces are out-crossers and therefore in Hardy Weinberg equilibrium. However often, deviation from random mating is observed and therefore an understanding of the population dynamics becomes necessary (Lia *et al.*, 2009). In plant breeding, because of their codominant behaviour, SSR markers have been commonly used for both model as well as distance-based population structuring studies (Kalia *et al.*, 2011; Ilyas *et al.*, 2023). The current study was therefore planned to characterize the genetic diversity in a collection of coloured maize landraces adapted to the NEHR at the phenotypic and genotypic level to develop nutritionally rich maize.

## Materials and methods

### Plant material

The maize landraces used in this study comprised of eighty-three maize accessions of which seventy-three bore pigmented ears while four and six were white and yellow respectively (online Supplementary Fig. S1). Fifty-two of these landraces (online Supplementary Table S1) were collected from different districts of the states of Arunachal Pradesh, Nagaland, Manipur, Mizoram, Tripura, Meghalaya and Sikkim in India during 2019 and 2020 and thirty-one landraces were obtained from ICAR-NBPGR Regional Station, Umiam -793103, Meghalaya (online Supplementary Table S2). The accessions were assigned alphanumeric numbers.

### Methodology

#### Field design and agronomy

The experimental trial was carried out in spring 2021 at the College of Post Graduate Studies in Agricultural Sciences,

Umiam located at an elevation of 950 m above mean sea level and at geographical coordinates of 25°34'32"N latitude and 91° 52'23"E longitude. The experiment was set in an alpha lattice design over two replications and four blocks. The rows were spaced 0.60 m apart and the spacing between plants within rows was maintained at 0.30 m. Routine inter-culture operations were taken up to ensure a healthy crop stand. An additional trial was done to confirm the observations of the previous year in spring of 2022.

### Morphological observations

In all fourteen quantitative, eight colour related and nine qualitative traits were studied. The quantitative traits included days to tasselling (DT, days), days to silking (DS, days), Anthesis Silking Interval (ASI, days), plant height (PH, cm), tassel length (TL, cm), ear height (EH, cm), total number of cobs (TNC, number), ear weight (EW, g), seed index (SI, g), total grain weight (TGW, g), total number of kernels (TNK, number), ear length (EL, cm), ear diameter (ED, cm) and number of grain kernel rows (NGKR, number). From the data of plant and ear height, the EH/PH ratio was worked out for principal component analysis (PCA) analysis. In 2021, observations with respect to presence of pigmentation in leaf sheath, pigmentation in brace root, pigmentation in base of glume of tassel, pigmentation in glume excluding base, pigmentation in anther on the day of emergence, pigmentation in silk on the day of emergence and pigmentation in glume of the cob were also taken. The qualitative traits included observation for angle between leaf and stem, attitude of leaf blade, density of tassel spikelets, angle between main axis of tassel and lateral branches, attitude of lateral branches of tassel, cob shape, kernel row arrangement, kernel shape and type of kernels. The data were recorded on five random plants for each of the two replications using standard descriptors outlined by the Protection of Plant Varieties and Farmers' Rights authority, New Delhi (Anonymous, 2007).

### Biochemical studies

Pigment characterization studies included spectrophotometric determination of anthocyanin (Pilu *et al.*, 2011) and phlobaphene (Das *et al.*, 1994; Pilu *et al.*, 2011) content, bleaching test and histological analysis of kernel tissues (Cassani *et al.*, 2017). For variegated maize, the pigmented kernels were used.

For anthocyanin estimation, 1 g of maize flour was added to 25 ml of acidified ethanol (96% ethanol and 1 N HCl 85:15 v/v) and incubated for 30 min at 50°C followed by centrifugation at 12,000 rpm for 15 min at 4°C. Absorbance (A) of the supernatant was measured at 530 nm and the amount of anthocyanin calculated as cyanidin 3-glucoside equivalents (molar extinction coefficient ( $\epsilon$ ) 26,900  $\text{lm}^{-1} \text{mol}^{-1}$ , M.W. 484.82). The results were expressed as mg of cyanidin 3-glucoside equivalents (CGE) per g of dry weight. Phlobaphene was extracted from individual seeds using one volume of concentrated hydrochloric acid (HCl) and four volumes of dimethyl sulfoxide (DMSO) added sequentially followed by centrifugation. The clear supernatant obtained was diluted with methanol (20% final concentration) and the concentration of phlobaphene in the sample expressed as absorbance value at  $\lambda$  max (510 nm) per g of dry weight. For both experiments, samples were handled in triplicate. Additionally, for the phlobaphene estimation white kernels devoid of pigmentation were also used as control. The spectrophotometric analysis of anthocyanin and phlobaphene content were also repeated for

the top twenty performers in the cropping seasons of 2022 and 2023.

For bleaching studies, a representative sample of seeds from all the accessions were bleached by immersing the seeds in 7% sodium hypochlorite solution for 3 h. The seeds were then thoroughly rinsed with tap water and pericarp tissue discolouration was studied. For the histological studies of kernel tissues, the seeds were soaked in water for 12 h and fixed in freshly prepared 4% paraformaldehyde in phosphate buffer solution (PBS) (130 mM NaCl, 7 mM  $\text{Na}_2\text{HPO}_4$ , 3 mM  $\text{NaH}_2\text{PO}_4 \cdot \text{H}_2\text{O}$ ) at 4°C overnight. They were then rinsed in 0.85% NaCl and transferred to 70% ethanol at 4°C. Thin cross sections made manually were mounted on slides using tert-butyl alcohol to preserve anthocyanin pigment *in-situ* and photographed using LEICA DM 750 microscope.

#### Molecular diversity studies

General diversity studies for the eighty-three accessions using thirty-four SSR (Simple Sequence Repeat) markers (Naveenkumar *et al.*, 2020) was followed by a more detailed pigment diversity analysis using twenty-one pigmentation specific SSR markers (online Supplementary Table S3) on seventy-one accessions in 2021. Twelve accessions which had recorded more than 15% missing data were omitted from this analysis. Accessions with high percentage of missing data would lead to statistical inaccuracies, generate bias and misinterpretations. The marker details for the genotyping studies were obtained from the maize GDB database (Portwood *et al.*, 2018). Leaf samples collected at seedling stage were used for extraction of genomic DNA using the modified CTAB extraction protocol (Doyle and Doyle, 1990). The PCR amplification was carried out in 10 µl reaction volume using a programmable thermal cycler. An initial denaturation step at 94°C for 4 min was followed up by 35 cycles of denaturation, annealing and extension at defined temperatures. The final extension was performed at 72°C for 4 min. The amplified PCR products were then resolved in a 2% agarose gel photographed using a gel documentation unit (Alpha Imager Mini) and scored based on size relative to a standard 100 bp ladder (Gene Ruler, Fermentas).

#### Data analysis

Analysis of variance (ANOVA) and post hoc tests for anthocyanin and phlobaphene content were also worked out using XL-Stat software (Addinsoft, 2022). ANOVA for the adjusted means values of the morphological parameters was calculated using 'PBIB' function of 'agricolae' package (Felipe, 2020) in R software (R Core Team, 1993). For the adjusted means, PCA to identify the most discriminatory traits was done in the R software package ggplot2 (Wickham, 2016). Pooled ANOVA of the two cropping seasons was done using 'gamem' of metan package (Olivoto and Lúcio, 2020) in R software (R Core Team, 2020). The pooled data (seasons as random) BLUEs (Best Linear Unbiased Estimate) were generated using the META-R software (Alvarado *et al.*, 2015) of CIMMYT.

The PIC values were calculated as per Nei (1973). STRUCTURE (Version 2.3.4) (Pritchard *et al.*, 2000) was used to evaluate genetic relationship among individuals and the underlying population structure. A total of 20 independent runs were performed for each *K*, with *K* values ranging from 2 to 10 and a Markov chain Monte Carlo replication burn length of 10,000–100,000 (Evanno *et al.*, 2005). The results obtained were corroborated with the online web-based STRUCTURE HARVESTER program to calculate the optimal *K* value. Individuals with

membership probability below 0.60 were grouped into a single admixture group.

The different parameters of genetic diversity were calculated using GenAlex software version 6.5 (Peakall and Smouse, 2012). These included analysis of molecular variance (AMOVA) and Wright's fixation index ( $F_{ST}$ ) of the sub-groups obtained from population sub structuring studies. Besides, principal coordinate analysis (PCoA), observed vs expected heterozygosity, Fixation index, Shannon's Information Index, *F*- coefficients and *G*-statistics of contributing loci were also worked out. The phylogenetic tree for distance based unweighted neighbour-joining clustering method was computed using DARwin 6.0.21.

## Results

### Morphological and molecular characterization of general diversity studies

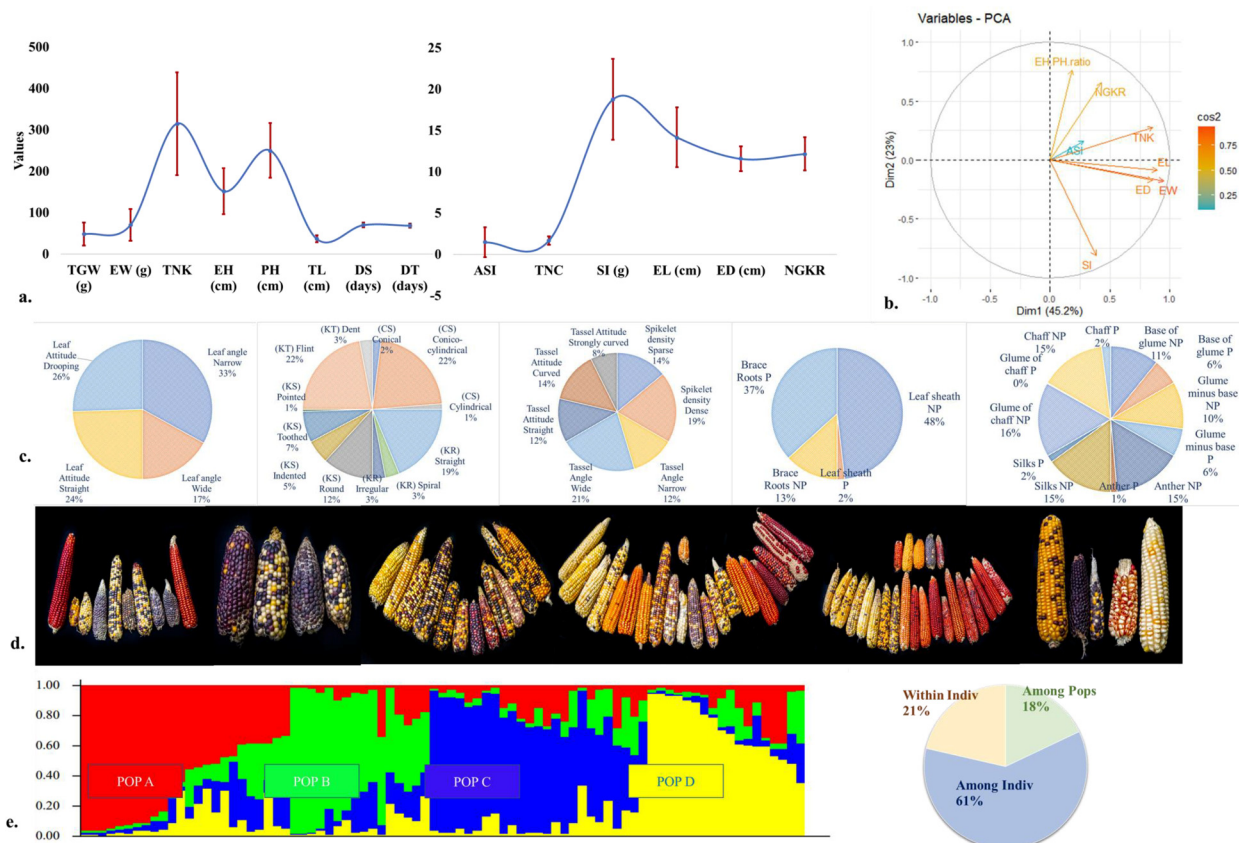
For the morphological studies (Fig. 1a), all the quantitative traits barring number of cobs exhibited high and significant variation (Table 1). A non-significant *F* value for number of cobs was because most accessions were highly prolific and as a result bore multiple cobs. Several protogynous accessions (P/AP 12, P/AP 14, P/N 3, P/N 4, P/N 6, P/MG 3, P/MG 4, P/MG 6, P/MG 8, P/MG 10, P/MG 11, W/MI 25, P/MA 3 and P/T 1) with negative ASI values were observed as opposed to protandry which is the norm. The PCA biplot (Fig. 1b) for eight highly variable traits indicated that PC1 and PC2 together could explain 68.18% of the total variation. Based on the cosine square results, PC1 had higher loadings of ear/kernel traits (EW, TNK, EL and ED) while PC2 recorded highest contribution from EH/PH ratio, SI and NGKR. Here, SI was found to be negatively correlated with EH/PH ratio and NGKR.

Pooled analysis from two seasons data indicated that mean values observed were similar with respect to the major yield and yield attributing traits studied (online Supplementary Fig. S2). PCA based on adjusted mean values of the pooled data also assigned ear traits as the highest contributor to variation in the data. However, the variation assigned to PC1 and PC2 based on pooled analysis was slightly lower in comparison to that of cropping season 1 taken alone.

The qualitative traits (Fig. 1c) which included studies of leaf traits, pigmentation of floral structures, tassel traits, cob appearance, and kernel types also displayed considerable variation. Most accessions had a narrow leaf angle, and the leaf blade attitude was either straight or drooping. Pigmentation in leaf sheath tissues was limited, but pigmentation of brace roots was more common. Variability was observed in tassel pigmentation while ears of only a few accessions were pigmented. Dense tassels were a common feature, and the angle between the tassel and lateral branches were either straight, curved or strongly curved. The cobs were mostly conico-cylindrical in appearance with a straight kernel row arrangement. Exceptions were the anthocyanin-rich cobs, which were mostly conical with an irregular kernel row arrangement. Most accessions had flint-type kernels.

The STRUCTURE analysis of the eighty-three accessions revealed four subgroups (POP A, POP B, POP C, POP D) and one admixture group (Fig. 1d). AMOVA for the four subgroups indicated that while majority of the variation was present among individuals within a population, a sizeable portion (18%) was because of differences among the populations (Fig. 1e). An overall  $F_{ST}$  value of 0.180 indicated presence of high population structuring.





**Figure 1.** General diversity studies of eighty-three maize accessions studied. (a). Line diagram illustrating mean values and variation for the fourteen yield and yield attributing traits, with error bars indicating  $\pm$  standard deviation from the mean (b). A biplot generated from principal component analysis for the eight most variable traits. Here, PC1 has a higher loading of ear/kernel traits, explaining 45.2% of the total variation while PC2 with higher loadings for EH/PH ratio, SI, and NGKR explains 23% of the variation observed (c). Pie diagrams representing per cent diversity and distribution of categorical traits related to leaf angle and attitude, tassel density, attitude and angle, kernel [Kernel row (KR), Kernel shape (KS), Kernel type (KT)] and cob [Cob shape (CS)] arrangement, and pigmentation for the studied maize accessions (d). Pictorial representation of the landraces collected from the northeastern states of Nagaland, Tripura, Meghalaya, Arunachal Pradesh, Mizoram, Manipur and Sikkim (left to right) (e). Four subgroups (POP A, POP B, POP C and POP D) obtained from population structuring studies in general diversity studies, along with a pie chart illustrating per cent variation as revealed by AMOVA.

### Biochemical, histological and molecular characterization studies of kernel pigmentation

Kernel anthocyanin content ranged between 1.43 CGE mg/g for the coloured cobs to 0 CGE mg/g in the yellow/white cobs used as control. Twenty-eight accessions with a cumulative average of 1.17 CGE mg/g recorded values above the average of 0.49 CGE mg/g. While anthocyanin content was studied for all accessions, phlobaphene studies were limited to a subset of 22 accessions with varying degrees of red. The phlobaphene content was found to range between 1.12 and 0.01 with a mean value of 0.36 A510/g. Except for a single accession (P/MA2) with pigment deposition in both the pericarp and aleurone layers, the anthocyanin content in the remaining 21 accessions was negligible. The highly significant ANOVA implied presence of genetic variability for kernel pigmentation among the accessions. Post hoc pairwise comparisons using Fisher's LSD helped to identify accessions with high values (Fig. 2). A comparison (online Supplementary Fig. S3) of the performance of the kernel anthocyanin and phlobaphene content for the top twenty performers in the first year and pooled analysis of three cropping seasons also gave near consistent results.

When maize kernels were treated with 7% sodium hypochlorite, the pigments in the kernel pericarp discoloured because of

oxidation, but the aleurone layer remained unaffected. Histological studies of the kernels in the first year further revealed that phlobaphene responsible for red pigmentation and anthocyanin, responsible for purple pigmentation, were restricted to the outer and inner layers of the kernels, respectively. Bleaching and histology studies (Fig. 3a) demonstrated six variations in kernel pigmentation

- (1) Colourless Pericarp, Colourless Aleurone
- (2) Orange Pericarp, Colourless Aleurone
- (3) Colourless Pericarp, Purple Aleurone
- (4) Red Pericarp, Colourless Aleurone
- (5) Red Pericarp, Purple Aleurone and
- (6) Striped Pericarp, Colourless Aleurone.

Accessions with purple kernels were more prevalent than those with red (Fig. 3b). The endosperm was either white or yellow.

For the twenty-one pigment specific SSR markers used on the seventy-one accessions, marker *p-Orp1* was monomorphic while SSR *p-phi031* of bin 6.04 was the least informative with a PIC value of 0.11. For the remaining 19 SSRs the PIC content ranged between 0.35 and 0.37. STRUCTURE grouped the accessions (Fig. 4a) into three subgroups – POP I, POP II and POP III

**Table 1.** Mean sum of squares based on ANOVA for the yield and pigmentation parameters under study

Parameters	Sources of variation			
	Accessions	Blocks within replication	Residuals	Replication
Phenological traits				
DT	160.22***	38.14*	7.46	128.63
DS	181.84***	35.83*	7.83	80.1
ASI	1.6646	0.5746	1.2412	3.1488
Morphological traits				
PH	10,117***	233	1167	12.1
EH	6642***	5184*	768	2508
TL	177.0***	426.8**	51	28
Ear/Kernel traits				
EW	3044.6***	97.8	425.5	1738.4
EL	30.758***	0.165	4.605	0.044
ED	9.541***	1.579	1.986	1.632
TGW	1558.2***	10.9	435.7	590.6
SI	56.40***	4.34	11.3	0.76
NGKR	11.246***	1.86	1.204	5.462
TNK	32,856***	45	5674	1176

DT, Days to tasselling (days); DS, Days to silking (days); ASI, Anthesis Silking Interval (days); PH, Plant height (cm); TL, Tassel length (cm); EH, Ear height (cm); TNC, total number of cobs (number); EW, Ear weight (g); SI, Seed index (g); TGW, Total grain weight (g); TNK, Total number of kernels (number); EL, Ear length (cm); ED, Ear diameter (cm); NGKR, Number of grain kernel rows (number).

$\alpha \leq 0.05$ , \*  $\alpha \leq 0.01$ , \*\*  $\alpha \leq 0.001$ \*\*\*.

and one admixture group. The grouping pattern based on average anthocyanin and phlobaphene content of individuals accessions within the subgroups indicated that POP I had accessions with the highest anthocyanin values. POP II on the other hand had grouped individuals with high phlobaphene content. Analysis of Nei Genetic Distance and Pairwise Population  $F_{ST}$  values however indicated that subgroups POP I and POP III were more divergent (Table 2) and that accessions with higher anthocyanin content were confined to subgroup POP I.

AMOVA indicated significant genetic differences for the subgroups with much of the variance concentrated in the 'Among Individuals' component. An 'Among Population' molecular variance of 10% which translates to an  $F_{ST}$  of 0.103 indicated presence of moderate genetic structuring for pigmentation between the subgroups identified (Fig. 4b). The first two axes of the principal co-ordinate analysis could explain 21.57% of the molecular variation detected with members of the divergent POP III and POP I loading heavily on PC 1 (11.37%) and PC 2 (10.20%) respectively. Individuals of POP II were scattered along either axis (Fig. 4c). The clustering pattern obtained from the distance-based un-weighted neighbour joining method (Fig. 4d) was comparable to the PCoA results obtained from STRUCTURE.

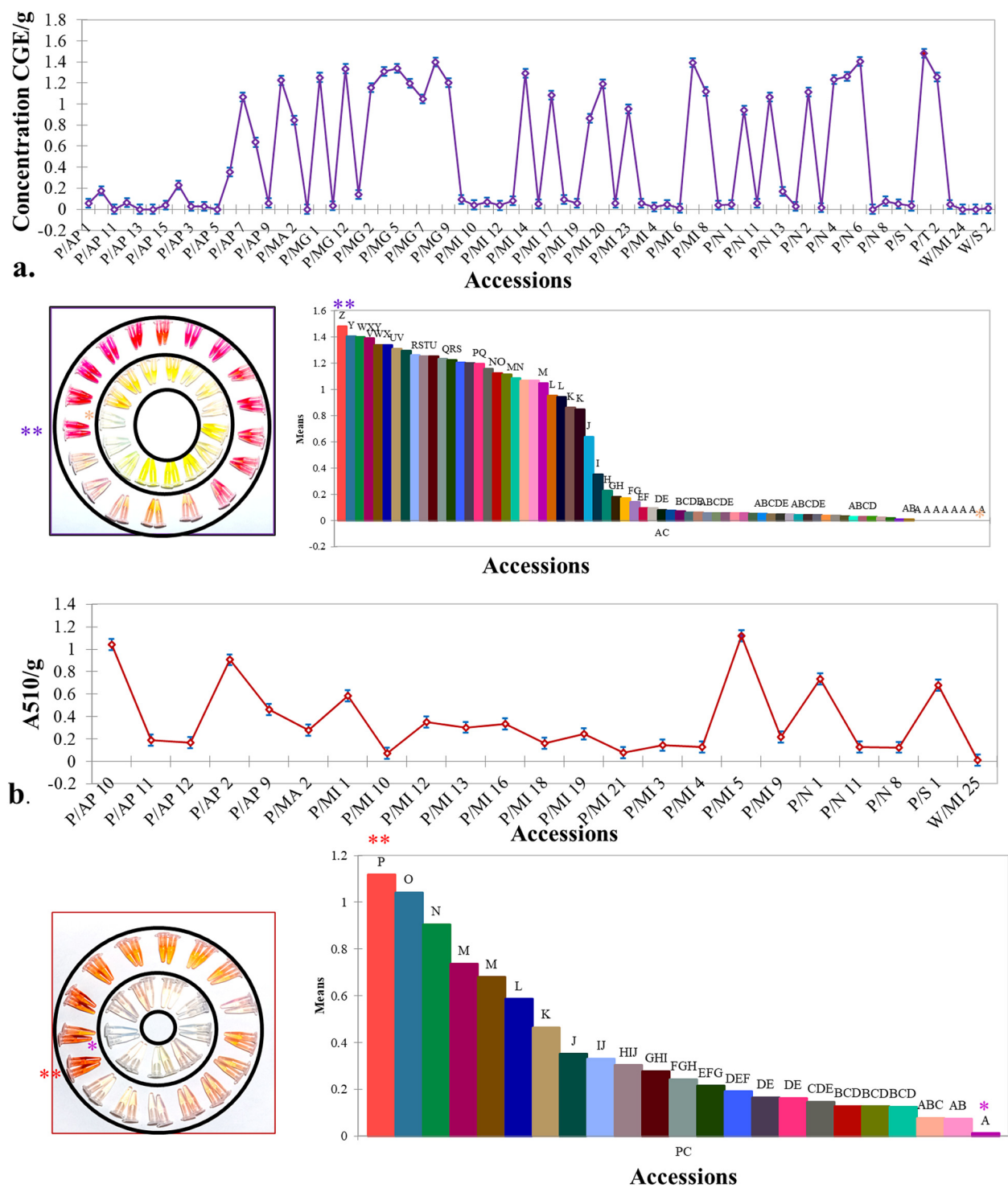
The observed heterozygosity was significantly lower than the expected heterozygosity as reflected by the high levels of fixation index in all the three subgroups. The fixation index was highest for subgroup POP III while comparatively lower values of Shannon's Information Index for POP I indicated that higher amounts of diversity among the members of this subgroup (online Supplementary Fig. S4). In all, twelve of the nineteen loci with very high to moderate  $F_{ST}$  values contributed to the

genetic differentiation of the three subgroups. Of these, seven SSR markers with  $F_{ST}$  values between 0.465 and 0.111 were the highest contributors (Fig. 4e). The high  $F_{ST}$  values for these loci were comparable with the estimated  $G'_{st}$  and  $G''_{st}$  values reaffirming the contribution of these loci in genetic structuring of the accessions under study. The Nm values which represent gene flow were lower for loci with higher  $F_{ST}$  values.

Based on un-weighted neighbour joining analysis, the highest genetic distance of 0.93 was detected was between accessions P/MI 23 and P/AP 10. Accession P/MI 23 had recorded above average anthocyanin content (0.96 CGE mg/g) while the anthocyanin content (0.17 CGE mg/g) in P/AP 10 was negligible. The accessions with the highest anthocyanin (P/T 1) and phlobaphene content (P/MI 5) were placed at a genetic distance of 0.045. Accessions P/MI 23 and P/T 1 belong to POP I while P/AP 10 and P/MI 5 belong to POP II.

## Discussion

From the current study, a common observation was that cobs with the highest anthocyanin content consistently recorded low ear weight in comparison to the other pigmented accessions. Additionally, anthocyanin content in the kernels was negatively but significantly correlated with seed index and ear length (data not shown). Most of these cobs also showed an irregular kernel row arrangement. These findings may explain why ears with high levels of anthocyanin are considered less productive and did not find preference during the course of domestication (Petroni *et al.*, 2014; Lago *et al.*, 2015). In what appears to be a trade-off between quality and yield, the challenge will be to

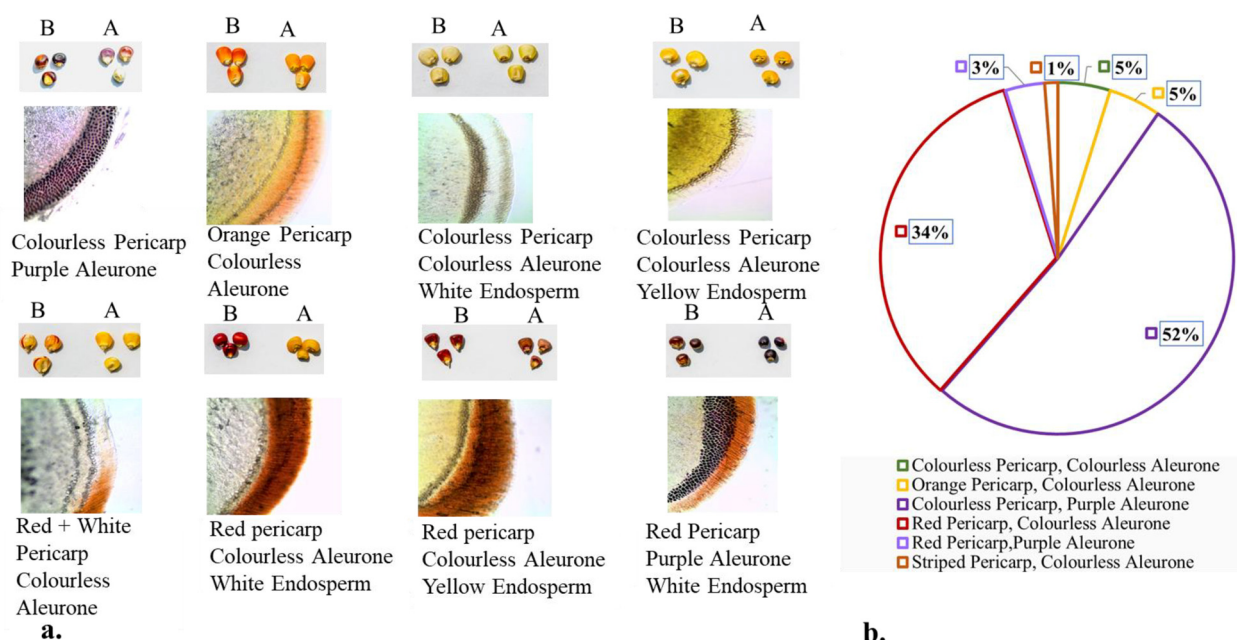


**Figure 2.** Spectrophotometric results based on colorimetric assay for (a). anthocyanin (AC) expressed as CGE/g and (b). phlobaphene (PC) content expressed as A510/g depicting the variation among the different accessions under study. The tubes placed within the circles represent the different shades of the two pigments extracted from kernels of the accessions studied. The bar diagram depicts all pairwise comparisons based on Fisher's LSD Error bars indicate  $\pm$  standard error.

develop lines with anthocyanin rich kernels and acceptable ear/kernel traits such as ear length, diameter, and kernel weight since they are critical components influencing grain yield (Fan *et al.*, 2008). Recurrent selection programmes for population improvement as suggested by Lago *et al.* (2014) and Chatham and Juvik (2021) to simultaneously accumulate desirable alleles

for enhancing pigmentation and yield into a common gene pool is one such way forward. Therefore, although the primary aim of this study was to identify accessions with high anthocyanin/phlobaphene content to initiate a breeding programme focused on nutritional parameters, general diversity studies were also simultaneously undertaken to





**Figure 3.** (a). Bleaching results before (B) and after (A) along with histological sections of the different types of pericarp/aleurone pigmentation seen among all the eighty-three accessions used in the study. Phlobaphene is confined to the pericarp while anthocyanin is limited to the aleurone layer (b). The pie chart depicts the per cent distribution of classes of pigmented kernels where of the eighty-three accessions under study, the highest numbers were of the colourless pericarp + purple aleurone type.

assess the overall variation present in the studied accessions. The significant variation observed among the accessions for the yield attributing traits suggests that selection has a genetic basis and would be effective. PCA studies specifically indicated that ear weight was the most variable. Previous studies on genetic variability of maize landraces from the NEHR had also reported on the highly significant contribution of cob weight to the overall variation (Sanjenbam *et al.*, 2018; Naveenkumar *et al.*, 2020).

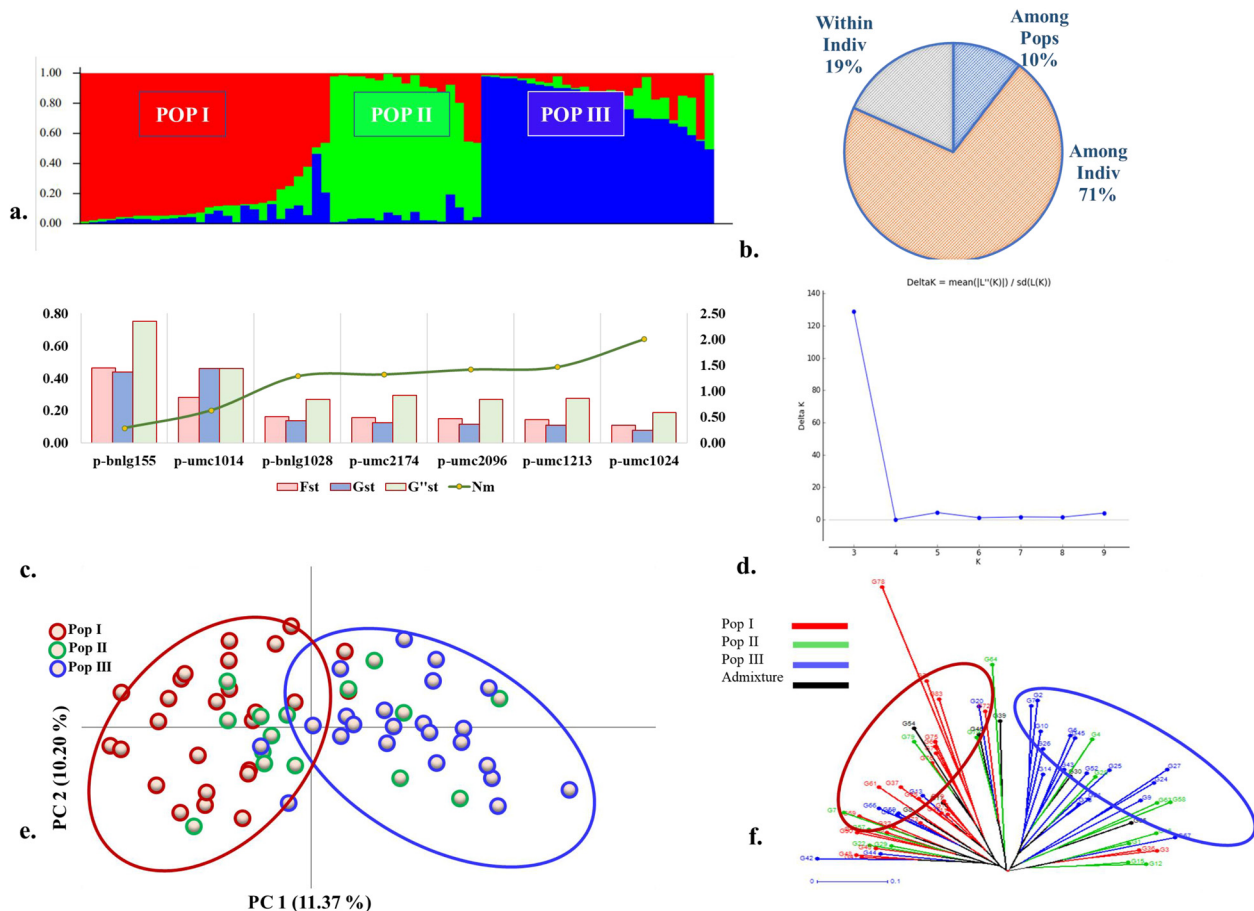
Since knowledge of interrelationship between grain yield and its associated independent variables improve the efficiency of breeding programmes (Mohammadi and Prasanna, 2003), yield association with other traits besides ear weight were also explored. A negative relationship between seed index and the EH/PH ratio observed meant that selection of accessions with a lower EH/PH ratio would lead to higher seed index. Low EH/PH ratio are advantageous since they impart stalk resistance and reduce the risk of lodging (Sandhu and Dhillon, 2021). Multi-cobbing – an expression of reproductive plasticity in combination with short ASI and protogyny observed for several of the accessions are key components for increasing yield under moisture stress conditions. Together, they are reported to promote synchronous pollination leading to higher kernel set (Uribelarrea *et al.*, 2002; Ngugi *et al.*, 2013; Adhikari *et al.*, 2021). Narrow leaf architecture with a straight leaf orientation observed in the germplasm are reported to contribute to increased light absorption and higher dry matter accumulation/unit land area (Stewart *et al.*, 2003; Araus *et al.*, 2008; Ku *et al.*, 2010). Accessions with sparse spikelet density allow higher portion of photo assimilate to be allocated for ear development and ensure light interception in high density populations (Gao *et al.*, 2020; Sandhu and Dhillon, 2021).

Although significant in both, Wright's  $F_{ST}$  and AMOVA was substantially higher for the general diversity studies as compared to pigment diversity studies. This is possibly because general diversity studies involve a wider set of traits and therefore may

be subject to higher selection pressures, leading to greater genetic differentiation among the subgroups. In contrast, pigment diversity focuses specifically on genetic variation related to anthocyanin/phlobaphene content among other pigments which are governed by a comparatively smaller number of genes leading to lower population structuring. In either case, a significant  $F_{ST}$  and AMOVA implied a deviation from random mating for the accessions under study (Excoffier *et al.*, 1992; Goudet, 1995; Meirmans, 2006; Holsinger and Weir, 2009). This is interesting since the accessions had been collected randomly from farmers across NEHR – kernel pigmentation being the only shared attribute.

Significant variation observed for kernel anthocyanin/phlobaphene content in the accessions indicated a genetic basis and meant that variation beyond what has been documented in the present accessions exists for landraces growing in the NEHR. Bleaching and histological examinations had confirmed that when present, anthocyanin was confined to the aleurone layer. Such maize are classified as blue maize (Chatham *et al.*, 2019; Chatham and Juvik, 2021; Paulsmeyer and Juvik 2021) as opposed to purple maize characterized by anthocyanin deposition in the pericarp of the kernels. However, this is not to say that purple maize is not found in the NEHR. As reported by Chatham and Juvik (2021), purple maize comprises a very small portion of available pigmented germplasm. A moderate population structuring was associated with a significant reduction in observed to expected heterozygosity within the subgroups.

$F_{ST}$  values also assist in identifying areas of the genome that have been targets of selection (Holsinger and Weir, 2009). For our studies, significant  $F_{ST}$  values were detected for loci *bnlg155*, *umc1014*, *bnlg1028*, *umc2174*, *umc2096*, *umc1213* and *umc1024* associated with genes *psy3*, *pl1*, *r1*, *a1*, *p2*, *in1* and *b1* respectively. While gene *psy3* fine tunes carotenoid flux in response to moisture stress in maize roots (Li *et al.*, 2008,



**Figure 4.** (a). Population subgroups POP I, POP II and POP III obtained using twenty-one SSR colour markers on seventy-one of the eighty-three accessions under study where  $k=4$ . (b). Pie chart indicating per cent molecular variation based on AMOVA (c). Loci *p-bnlgl55*, *p-bnlgl028*, *p-umc2174*, *p-umc2096*, *p-umc1213* and *p-umc1024* with the highest  $F_{ST}$ ,  $G_{ST}$  and comparatively lower  $N_m$  values (d). The 'k' value, as computed by STRUCTURE HARVESTER, also suggests that the probable count of population subgroups present is three. (e). PCoA graph depicting distribution of the seventy-one accessions characterized for pigmentation (f). A radial phylogenetic tree constructed using distance-based clustering analysis for the same seventy-one accessions. Colours and the legend depict the population groups for the accessions as obtained by STRUCTURE analysis.

2009), *pl1* of the R2R3MYB gene family encodes transcription factors (Lago *et al.*, 2014) that regulate anthocyanin biosynthesis in vegetative tissues including the seed pericarp. The *pl1* gene is also implicated in determining the response of the vegetative tissues to light (Piazza *et al.*, 2002) and a major QTL associated with root colour in maize has been reported to be located near the *pl1* gene on chromosome 6 (Li *et al.*, 2022).

For the *r1* and *b1* genes associated with loci *bnlg1028* and *umc1024*, both are functionally equivalent having arisen because of duplication of an ancestral gene (Radicella *et al.*, 1992). While *r1* alleles are part of a complex of several tightly linked genes in chromosomal bin10.06 (Lago *et al.*, 2014), *b1* alleles are encoded by a single gene in bin 2.04. The allelic diversity for the *r1/b1* complex varies greatly ranging from vegetative tissue specific anthocyanin production to seed aleurone specific anthocyanin production and penetration (Selinger and Chandler, 2001). The *intensifier 1* gene (*in1*) which bears a homology with the *r1/b1* multigene family is known to activate the production of anthocyanin pigments in the aleurone layer when in its recessive state (Burr *et al.*, 1996; Selinger and Chandler, 1999; Chatham *et al.*, 2019). Gene *a1* (*anthocyaninless1*) associated with locus *umc2174* regulates changes (absence/reduction) in aleurone pigmentation caused by activation of both autonomous

and non-autonomous transposons (Rhoades, 1938; O'Reilly *et al.*, 1985; Brown *et al.*, 1989; Peniche-Pavía *et al.*, 2022) and possibly accounts for the high number of variegated cobs in the study. The gene *p2* (Zhang *et al.*, 2000; Zhang and Peterson, 2005) codes for an MYB-related protein that regulates pigmentation in developing anthers and silks.

All in all, STRUCTURE and neighbour joining analyses produced near similar subgroupings for pigmentation diversity with no obvious geographical bias. Most of the molecular variation was ascribed to the 'Among Individuals' component which meant that the accessions within the subgroups were genetically diverse (Reif *et al.*, 2003) creating opportunities for identifying potential parental lines for exploitation of heterosis (Springer and Stupar, 2007). Additionally, the prevalence of traits such as multi-cobbing, short ASI, and protogyny, along with the highly fixed *psy3* gene, indicated that the accessions have adaptive advantages to deal with moisture stress. A possible explanation for this adaptation would be that maize landraces in the NEHR complete a good part of their life cycle under rainfed situations before the onset of monsoon. Taking all the findings together it appears that the maize landraces growing in the NEHR are diverse, come with adaptive advantages and amenable to genetic manipulation for developing high yielding coloured maize.



**Table 2.** Phenotypic characterization based on pigmentation and genetic distance based on Nei's unbiased genetic distance (represented above the diagonal) and pairwise  $F_{ST}$  values (represented below the diagonal)

Phenotypic characterization	POP I	POP II	POP III
Vegetative tissue pigmentation present (%)			
Brace root	62	<b>87</b>	70
Leaf sheath	<b>8</b>	0	4
Base of glume	15	<b>47</b>	26
Rest of glume	<b>42</b>	27	30
Anthers	8	7	<b>9</b>
Silk	12	7	<b>17</b>
Anthocyanin			
Overall average (CGE mg/g)	<b>0.82</b>	0.24	0.62
Anthocyanin			
Average of purple pigmented kernels only (CGE mg/g)	<b>1.16</b>	1.31 (Single accession only)	1.1
Phlobaphene			
Average of red pigmented kernels (A510/g)	0.20	<b>0.41</b>	0.39
Genetic distance			
POP I		0.093	<b>0.138</b>
POP II	0.076		0.079
POP III	<b>0.098</b>	0.055	

DT, Days to tasselling (days); DS, Days to silking (days); ASI, Anthesis Silking Interval (days); PH, Plant height (cm); TL, Tassel length (cm); EH, Ear height (cm); TNC, total number of cobs (number); EW, Ear weight (g); SI, Seed index (g); TGW, Total grain weight (g); TNK, Total number of kernels (number); EL, Ear length (cm); ED, Ear diameter (cm); NGKR, Number of grain kernel rows (number).

Highest values are presented in bold.

## Conclusion

To the best of our knowledge, this is the first in-depth study of coloured maize landraces from the NEHR of India and is a starting point for crop improvement aimed at enhancing anthocyanin/phlobaphene content in maize. We also understand that the genetic variation observed among the studied accessions at the morphological, biochemical, and molecular levels can be extended to the larger population of coloured maize landraces of the NEHR. Since the genetic variability found is significant it will increase the chances of combining desirable traits for pigmentation and yield into a common gene pool. For our subset we could group the accessions based on model and distance-based clustering studies into distinct groups. These can subsequently be developed as heterotic groups. The initial step of assembling and identifying genetically diverse individuals has been completed in the present study. The accessions studied come with certain advantageous yield attributing traits such as adaptiveness to moisture stress, improved stalk strength to counter lodging especially in high density planting scenarios and leaf architecture which optimize photosynthesis and can be successfully harnessed. Promising lines can next be inbred and based on their performance with divergent testers can be classified into opposite heterotic pools for exploiting heterosis. Additionally, while our primary focus was on identifying promising accessions for anthocyanin/phlobaphene, other pigment variations such as accession with orange pericarp can also be explored.

**Supplementary material.** The supplementary material for this article can be found at <https://doi.org/10.1017/S1479262124000182>.

**Acknowledgements.** The authors are grateful to the farmers of the different districts of NEHR and to the ICAR-NBPG Regional Station, Umiam for their

willingness to provide the seed material used in the present study. We are also thankful to all those individuals who facilitated the germplasm collection by serving as contact points. Thanks, are also due to Dr Mayank Rai and Dr Wricha Tyagi for their valuable suggestions from time to time. The authors are also grateful to the Central Agricultural University (Imphal) for providing all necessary infrastructure for carrying out this research work.

**Authors' contributions.** Dr Devyani Sen and Ms Sristishila Baruah contributed to the study conception, design, experimentation, data analysis and draft preparation. Ms Thounaojam Bharti phenotyped the second and third season crop data as part of her M.Sc. thesis work. Dr Duddukur Rajasekhar and Mr Harshavardhan Reddy supported the study with material preparation, field data collection and morphological data analysis. Ms Ernieca Lyngdoh Nongbri assisted with the molecular work.

**Competing interests.** None.

## References

- Addinsoft (2022) *XLSTAT Statistical and Data Analysis Solution*. New York, USA: Addinsoft. Available at <https://www.xlstat.com/en>.
- Adhikari S, Joshi A, Kumar A and Singh NK (2021) Diversification of maize (*Zea mays* L.) through teosinte (*Zea mays* subsp. *parviglumis* Iltis and Doebley) allelic. *Genetic Resources and Crop Evolution* **68**, 2983–2995. <https://doi.org/10.1007/s10722-021-01170-z>.
- Adom KK and Liu RH (2002) Antioxidant activity of grains. *Journal of Agricultural and Food Chemistry* **50**, 6182–6187. <https://doi.org/10.1021/jf0205099>.
- Alvarado G, López M, Vargas M, Pacheco-Gil RÁ, Rodríguez F, Burguenio J and Crossa J (2015) META-R (Multi Environment Trial Analysis with R for Windows) Version 6.04. <https://hdl.handle.net/11529/10201>, CIMMYT Research Data & Software Repository Network, V23.
- Andorf C, Beavis WD, Hufford M, Smith S, Suza WP, Wang K, Woodhouse M, Yu J and Lubberstedt T (2019) Technological advances in maize

- breeding: past, present and future. *Theoretical and Applied Genetics* **132**, 817–849. <https://doi.org/10.1007/s00122-019-03306-3>.
- Anonymous** (2007) Guidelines for the conduct of tests for distinctness, uniformity and stability on maize (*Zea mays* L.). The protection of plant varieties and farmers rights authority, New Delhi. *Plant Variety Journal India* **1**, 1–13.
- Araus JL, Slafer GA and Serret MD** (2008) Breeding for yield potential and stress adaptation in cereals. *Critical Reviews in Plant Science* **27**, 377–412. <https://doi.org/10.1080/07352680802467736>.
- Bennetzen J, Buckler E, Chandler V, Doebley J, Dorweiler J, Gaut B, Freeling M, Hake S, Kellogg E, Poethig RS and Walbot V** (2001) Genetic evidence and the origin of maize. *Latin American Antiquity* **12**, 84–86. <https://doi.org/10.2307/971759>.
- Bhat KV, Chandel KPS, Shivakumar TM and Sachan JKS** (1998) Isozyme diversity in Indian primitive maize landraces. *Journal of Plant Biochemistry and Biotechnology* **7**, 23–27. <https://doi.org/10.1007/BF03263028>.
- Brown JJ, Mattes MG, O'Reilly C and Shepherd NS** (1989) Molecular characterization of rDt, a maize transposon of the 'Dotted' controlling element system. *Molecular and General Genetics MGG* **215**, 239–244. <https://doi.org/10.1007/BF00339723>.
- Bruggeman SA, Horvath DP, Fennell AY, Gonzalez-Hernandez JL and Clay SA** (2020) Teosinte (*Zea mays* ssp *parviglumis*) growth and transcriptomic response to weed stress identifies similarities and differences between varieties and with modern maize varieties. *PLoS One* **15**, e0237715. <https://doi.org/10.1371/journal.pone.0237715>.
- Burr FA, Burr B, Scheffler BE, Blewitt M, Wienand U and Matz EC** (1996) The maize repressor-like gene intensifier1 shares homology with the r1/b1 multigene family of transcription factors and exhibits missplicing. *The Plant Cell* **8**, 1249–1259. <https://doi.org/10.1105/tpc.8.8.1249>.
- Cassani E, Puglisi D, Cantaluppi E, Landoni M, Giupponi L, Giorgi A and Pilu R** (2017) Genetic studies regarding the control of seed pigmentation of an ancient European pointed maize (*Zea mays* L.) rich in phlobaphenes: the 'Nero Spinoso' from the Camonica valley. *Genetic Resources and Crop Evolution* **64**, 761–773. <https://doi.org/10.1007/s10722-016-0399-7>.
- Chatham LA and Juvik JA** (2021) Linking anthocyanin diversity, hue, and genetics in purple corn. *G3* **11**, jkaa062. <https://doi.org/10.1093/g3journal/jkaa062>.
- Chatham LA, Paulsmeyer M and Juvik JA** (2019) Prospects for economical natural colorants: insights from maize. *Theoretical and Applied Genetics* **132**, 2927–2946. <https://doi.org/10.1007/s00122-019-03414-0>.
- Das OP, Morales M and Messing J** (1994) Quantitative extraction of pericarp pigments. *Maize Genetics Cooperation Newsletter* **68**, 79–83.
- de Pascual-Teresa S and Sanchez-Ballesta MT** (2008) Anthocyanins: from plant to health. *Phytochemistry Reviews* **7**, 281–299. <https://doi.org/10.1007/s11101-007-9074-0>.
- Dhawan NL** (1964) Primitive maize in Sikkim. *Maize Genetics Cooperation Newsletter* **38**, 69–70.
- Doyle JJ and Doyle JL** (1990) Isolation of plant DNA from fresh tissue. *Focus* **12**, 13–11.
- Evanno G, Regnaut S and Goudet J** (2005) Detecting the number of clusters of individuals using the software STRUCTURE: a simulation study. *Molecular Ecology* **14**, 2611–2620. <https://doi.org/10.1111/j.1365-294X.2005.02553.x>.
- Excoffier L, Smouse PE and Quattro J** (1992) Analysis of molecular variance inferred from metric distances among DNA haplotypes: application to human mitochondrial DNA restriction data. *Genetics* **131**, 479–491. <https://doi.org/10.1093/genetics/131.2.479>.
- Fan XM, Chen HM, Tan J, Xu CX, Zhang YD, Luo LM, Huang YX and Kang MS** (2008) Combining abilities for yield and yield components in maize. *Maydica* **53**, 39.
- Felipe de Mendiburu F** (2020) *Agricolae: Statistical Procedures for Agricultural Research*. R package version 1.3-3.
- Flint-Garcia SA** (2013) Genetics and consequences of crop domestication. *Journal of Agricultural and Food Chemistry* **61**, 8267–8276. <https://doi.org/10.1021/jf305511d>.
- Food and Agriculture Organization of the United Nations (FAO)** (2019) *World Food and Agriculture: Statistical Pocketbook*. Rome, Italy: FAO; ISBN 978-92-5-131849-2.
- Gao Z, Sun L, Ren JH, Liang XG, Shen S, Lin S, Zhao X, Chen XM, Wu G and Zhou SL** (2020) Detasseling increases kernel number in maize under shade stress. *Agricultural and Forest Meteorology* **280**, 107811. <https://doi.org/10.1016/j.agrformet.2019.107811>.
- Gorjanc G, Jenko J, Hearne SJ and Hickey JM** (2016) Initiating maize pre-breeding programs using genomic selection to harness polygenic variation from landrace populations. *BMC Genomics* **17**, 1–15. <https://doi.org/10.1186/s12864-015-2345-z>.
- Goudet J** (1995) FSTAT (version 1.2): a computer program to calculate F-statistics. *Journal of Heredity* **86**, 485–486.
- Hallauer AR and Carena MJ** (2009) Maize. In Carena M (ed.), *Cereals: Handbook of Plant Breeding*, Vol 3. New York: Springer, pp. 3–98. [https://doi.org/10.1007/978-0-387-72297-9\\_1](https://doi.org/10.1007/978-0-387-72297-9_1).
- Holsinger KE and Weir BS** (2009) Genetics in geographically structured populations: defining, estimating and interpreting F ST. *Nature Reviews Genetics* **10**, 639–650. <https://doi.org/10.1038/nrg2611>.
- Ilyas MZ, Park H, Jang SJ, Cho J, Sa KJ and Lee JK** (2023) Association mapping for evaluation of population structure, genetic diversity, and physiological traits in drought-stressed maize germplasm using SSR markers. *Plants* **12**, 4092. <https://doi.org/10.3390/plants12244092>.
- Kalia RK, Rai MK, Kalia S, Singh R and Dhawan AK** (2011) Microsatellite markers: an overview of the recent progress in plants. *Euphytica* **77**, 309–334. <https://doi.org/10.1007/s10681-010-0286-9>.
- Ku LX, Zhao WM, Zhang J, Wu LC, Wang CL, Wang PA, Zhang WQ and Chen YH** (2010) Quantitative trait loci mapping of leaf angle and leaf orientation value in maize (*Zea mays* L.). *Theoretical and Applied Genetics* **121**, 951–959. <https://doi.org/10.1007/s00122-010-1364-z>.
- Lago C, Cassani E, Zanzi C, Landoni M, Trovato R and Pilu R** (2014) Development and study of a maize cultivar rich in anthocyanins: coloured polenta, a new functional food. *Plant Breeding* **133**, 210–217. <https://doi.org/10.1111/pbr.12153>.
- Lago C, Landoni M, Cassani E, Cantaluppi E, Doria E, Nielsen E, Giorgi A and Pilu R** (2015) Study and characterization of an ancient European flint white maize rich in anthocyanins: Millo Corvo from Galicia. *PLoS One* **10**, e0126521. <https://doi.org/10.1371/journal.pone.0130110>.
- Li F, Vallabhaneni R and Wurtzel ET** (2008) PSY3, a new member of the phytoene synthase gene family conserved in the Poaceae and regulator of abiotic-stress-induced root carotenogenesis. *Plant Physiology* **146**, 1333–1345. <https://doi.org/10.1104/pp.107.111120>.
- Li F, Tzfadia O and Wurtzel ET** (2009) The phytoene synthase gene family in the Grasses: subfunctionalization provides tissue-specific control of carotenogenesis. *Plant Signaling and Behavior* **4**, 208–211. <https://doi.org/10.4161/psb.4.3.7798>.
- Li Y, Fang X and Lin Z** (2022) Convergent loss of anthocyanin pigments is controlled by the same MYB gene in cereals. *Journal of Experimental Botany* **73**, 6089–6102. <https://doi.org/10.1093/jxb/erac270>.
- Lia VV, Poggio L and Confalonieri VA** (2009) Microsatellite variation in maize landraces from Northwestern Argentina: genetic diversity, population structure and racial affiliations. *Theoretical and Applied Genetics* **119**, 1053–1067. <https://doi.org/10.1007/s00122-009-1108-0>.
- Matsuoka Y, Vigouroux Y, Goodman MM, Sanchez GJ, Buckler E and Doebley J** (2002) A single domestication for maize shown by multilocus microsatellite genotyping. *Proceedings of the National Academy of Sciences* **99**, 6080–6084. <https://doi.org/10.1073/pnas.052125199>.
- Meirmans PG** (2006) Using the AMOVA framework to estimate a standardized genetic differentiation measure. *Evolution* **60**, 2399–2402. <https://doi.org/10.1111/j.0014-3820.2006.tb01874.x>.
- Mohammadi SA and Prasanna BM** (2003) Analysis of genetic diversity in crop plants-salient statistical tools and considerations. *Crop Science* **43**, 1235–1248.
- Naveenkumar KL, Sen D, Vashum S and Sanjenbam M** (2020) Genetic characterization and divergence studies of maize (*Zea mays* L.) lines developed from landraces indigenous to North Eastern Hill Region (NEHR) of India. *Plant Genetic Resources* **18**, 231–242. <https://doi.org/10.1017/S1479262120000246>.
- Nei M** (1973) Analysis of gene diversity in subdivided populations. *Proceedings of the National Academy of Sciences* **70**, 3321–3323. <https://doi.org/10.1073/pnas.70.12.3321>.

- Ngugi K, Collins JO and Muchira S (2013) Combining, earliness, short anthesis to silking interval and yield based selection indices under intermittent water stress to select for drought tolerant maize. *Australian Journal of Crop Science* 7, 2014–2020.
- Olivoto T and Lúcio ADC (2020) metan: an R package for multi-environment trial analysis. *Methods in Ecology and Evolution* 11, 783–789. <https://doi.org/10.1111/2041-210X.13384>.
- O'Reilly C, Shepherd NS, Pereira A, Schwarz-Sommer Z, Bertram I, Robertson DS, Peterson PA and Saedler H (1985) Molecular cloning of the a1 locus of *Zea mays* using the transposable elements En and Mu1. *The EMBO Journal* 4, 877–882. <https://doi.org/10.1105/tpc.12.12.2311>.
- Paauw M, Koes R and Quattrocchio FM (2019) Alteration of flavonoid pigmentation patterns during domestication of food crops. *Journal of Experimental Botany* 70, 3719–3735. <https://doi.org/10.1093/jxb/erz141>.
- Paulsmeyer M and Juvik J (2021) Functional characterization of an anthocyanin dimalonyltransferase in maize. *Molecules* 26, 2020. <https://doi.org/10.3390/molecules26072020>.
- Peakall R and Smouse PE (2012) GENALEX 6.5: genetic analysis in EXCEL. Population genetic software for teaching and research – an update. *Bioinformatics* 28, 2537–2539. <https://doi.org/10.1111/j.1471-8286.2005.01155.x>.
- Peniche-Pavía HA, Guzmán TJ, Magaña-Cerino JM, Gurrola-Díaz CM and Tiessen A (2022) Maize flavonoid biosynthesis, regulation, and human health relevance: a review. *Molecules* 27, 5166. <https://doi.org/10.3390/molecules27165166>.
- Petrescu DC, Vermeir I and Petrescu-Mag RM (2020) Consumer understanding of food quality, healthiness, and environmental impact: a cross-national perspective. *International Journal of Environmental Research and Public Health* 17, 169. <https://doi.org/10.3390/ijerph17010169>.
- Petroni K, Pilu R and Tonelli C (2014) Anthocyanins in corn: a wealth of genes for human health. *Planta* 240, 901–911. <https://doi.org/10.1007/s00425-014-2131-1>.
- Piazza P, Procissi A, Jenkins GI and Tonelli C (2002) Members of the c1/pl1 regulatory gene family mediate the response of maize aleurone and mesocotyl to different light qualities and cytokinins. *Plant Physiology* 128, 1077–1086. <https://doi.org/10.1104/pp.010799>.
- Pilu R, Cassani E, Sirizzotti A, Petroni K and Tonelli C (2011) Effect of flavonoid pigments on the accumulation of fumonisin B1 in the maize kernel. *Journal of Applied Genetics* 52, 145–152. <https://doi.org/10.1007/s13353-010-0014-0>.
- Portwood JL, Woodhouse MR, Cannon EK, Gardiner JM, Harper LC, Schaeffer ML, Walsh JR, Sen TZ, Cho KT, Schott DA and Braun BL (2018) MaizeGDB 2018: the maize multi-genome genetics and genomics database. *Nucleic Acids Research* 47, D1146–D1154. <https://doi.org/10.1093/nar/gky1046>.
- Prasanna BM (2012) Diversity in global maize germplasm: characterization and utilization. *Journal of Biosciences* 37, 843–855. <https://doi.org/10.1007/s12038-012-9227-1>.
- Pritchard JK, Stephens M and Donnelly P (2000) Inference of population structure using multilocus genotype data. *Genetics* 155, 945–959. <https://doi.org/10.1534/genetics.116.195164>.
- Radicella JP, Brown D, Tolar LA and Chandler VL (1992) Allelic diversity of the maize B regulatory gene: different leader and promoter sequences of two B alleles determine distinct tissue specificities of anthocyanin production. *Genes and Development* 6, 2152–2164. <https://doi.org/10.1101/gad.6.11.2152>.
- Rathod NKK, Kumari J, Hossain F, Chhabra R, Roy S, Harish GD, Bhardwaj R, Gadag RN and Misra AK (2019) Characterization of Mimban maize landrace from North-Eastern Himalayan region using microsatellite markers. *Journal of Plant Biochemistry and Biotechnology* 29, 323–335. <https://doi.org/10.1007/s13562-019-00524-0>.
- R Core Team (1993) *R: A Language and Environment for Statistical Computing*. Vienna, Austria: R Foundation for Statistical Computing.
- R Core Team (2020) *R: A Language and Environment for Statistical Computing*. Vienna, Austria: R Foundation for Statistical Computing. Available at <https://www.r-project.org> (accessed 21 January 2024).
- Reif JC, Melchinger AE, Xia XC, Warburton ML, Hoisington DA, Vasal SK, Srinivasan G, Bohn M and Frisch M (2003) Genetic distance based on simple sequence repeats and heterosis in tropical maize populations. *Crop Science* 43, 1275–1282. <https://doi.org/10.2135/cropsci2003.1275>.
- Rhoades MM (1938) Effect of the Dt gene on the mutability of the a1 allele in maize. *Genetics* 23, 377. <https://doi.org/10.1093/genetics/23.4.377>.
- Ryu SH, Werth L, Nelson S, Scheerens JC and Pratt RC (2013) Variation of kernel anthocyanin and carotenoid pigment content in USA/Mexico borderland landraces of maize. *Economic Botany* 67, 98–109. <https://doi.org/10.1007/s12231-013-9232-9>.
- Sachan JKS and Sarkar KR (1982) Plant type of 'Sikkim Primitives'. *Maize Genetics Cooperation Newsletter* 56, 122–124.
- Sachan JS and Sarkar KR (1986) Sikkim primitive maize – an overview. *Indian Journal of Genetics and Plant Breeding* 46, 153–161.
- Sandhu S and Dhillon BS (2021) Breeding plant type for adaptation to high plant density in tropical maize – a step towards productivity enhancement. *Plant Breeding* 140, 509–518. <https://doi.org/10.1111/pbr.12949>.
- Sanjenbam M, Sen D, Tyagi W and Chand R (2018) Phenotypic diversity analysis and screening for northern corn leaf blight resistance in maize (*Zea mays* L.) landraces grown in North Eastern hill region of India. *Indian Journal of Plant Genetic Resources* 31, 286–294. <http://doi.org/10.5958/0976-1926.2018.00033.5>.
- Selinger DA and Chandler VL (1999) A mutation in the pale aleurone color1 gene identifies a novel regulator of the maize anthocyanin pathway. *The Plant Cell* 11, 5–14. <https://doi.org/10.1105/tpc.11.1.5>.
- Selinger DA and Chandler VL (2001) B-Bolivia, an allele of the maize b1 gene with variable expression, contains a high copy retrotransposon-related sequence immediately upstream. *Plant Physiology* 125, 1363–1379. <https://doi.org/10.1104/pp.125.3.1363>.
- Sharma SK and Brahmi P (2011) Gene bank curators: towards implementation of the international treaty on plant genetic resources for food and agriculture by the Indian National Gene Bank. In: Frison C, López F and Esquinas Alcazar JT (eds), *Plant Genetic Resources and Food Security: Stakeholder Perspectives on the International Treaty on Plant Genetic Resources for Food and Agriculture*. Oxon: FAO and Bioversity International, pp. 183–196.
- Sharma L, Prasanna BM and Ramesh B (2010) Analysis of phenotypic and microsatellite-based diversity of maize landraces in India, especially from the North East Himalayan region. *Genetica* 138, 619–631. <https://doi.org/10.1007/s10709-010-9436-1>.
- Singh B (1977) *Races of Maize in India*. New Delhi: Indian Council of Agricultural Research.
- Singh AR, Singh SB, Dutta SK, Boopathi T, Lungmuana, Saha S, Devi MT and Singh NH (2016) Multi cob-bearing popcorn (Pukazo maize): a unique landrace of Mizoram, North East, India. *Current Science* 110, 1392–1393.
- Springer NM and Stupar RM (2007) Allelic variation and heterosis in maize: how do two halves make more than a whole? *Genome Research* 17, 264–275. <https://doi.org/10.1101/gr.5347007>.
- Stewart DW, Costa C, Dwyer LM, Smith DL, Hamilton RI and Ma BL (2003) Canopy structure, light interception, and photosynthesis in maize. *Agronomy Journal* 95, 1465–1474. <https://doi.org/10.2134/agronj2003.1465>.
- Stitzer MC and Ross-Ibarra J (2018) Maize domestication and gene interaction. *New Phytologist* 220, 395–408. <https://doi.org/10.1111/nph.15350>.
- Uribealarea M, Cárcova J, Otegui ME and Westgate ME (2002) Pollen production, pollination dynamics, and kernel set in maize. *Crop Science* 42, 1910–1918. <https://doi.org/10.2135/cropsci2002.1910>.
- Wickham H (2016) Data analysis. In *ggplot2*. Use R!. Cham: Springer, pp. 189–201. [https://doi.org/10.1007/978-3-319-24277-4\\_9](https://doi.org/10.1007/978-3-319-24277-4_9).
- Zhang F and Peterson T (2005) Comparisons of maize pericarp color1 alleles reveal paralogous gene recombination and an organ-specific enhancer region. *The Plant Cell* 17, 903–914. <https://doi.org/10.1105/tpc.104.029660>.
- Zhang P, Chopra S and Peterson T (2000) A segmental gene duplication generated differentially expressed myb-homologous genes in maize. *The Plant Cell* 12, 2311–2322. <https://doi.org/10.1105/tpc.12.12.2311>.
- Zilic S, Serpen A, Akillioglu G, Gökmen V and Vancetovic J (2012) Phenolic compounds, carotenoids, anthocyanins, and antioxidant capacity of colored maize (*Zea mays* L.) kernels. *Journal of Agricultural and Food Chemistry* 60, 1224–1231. <https://doi.org/10.1021/jf204367z>.