


ARTICLE

The microbiome in adult acute appendicitis

Mei Sze Lee^{1,2,*} , Arielle Sulit^{1,3}, Frank Frizelle^{1,2} and Rachel Purcell¹

¹Department of Surgery, University of Otago, Christchurch, New Zealand

²Department of General Surgery, Christchurch, New Zealand

³Massey University, Auckland, New Zealand

*Corresponding author. Email: drmeilee@gmail.com

(Received 08 March 2022; revised 06 June 2022; accepted 26 July 2022)

Abstract

Acute appendicitis is a common acute surgical emergency; however, the pathogenesis of adult appendicitis remains poorly understood. The microbiome is increasingly thought to play a key role in inflammatory disease of the bowel and similarly, may play a role in appendicitis. This study aimed to characterise the microbiome of adult acute appendicitis in a prospective cohort. We recruited 60 adults with acute appendicitis and 20 healthy controls. Rectal swabs were taken from each patient. After DNA extraction, 16S rRNA amplicon sequencing was carried out for analysis of diversity and taxonomic abundance. Phylogenetic sequencing of the samples indicated that there is a difference between the microbial composition of those with acute appendicitis and healthy controls, with a statistically significant decrease in alpha diversity in rectal swabs of appendicitis patients compared to healthy controls. At the genus level, we saw an increased abundance of potential pathogens, for example, *Parvimonas* and *Acinetobacter*, and a decrease in commensal taxa such as *Faecalibacterium*, *Blautia* and *Lachnospiraceae* in appendicitis patients compared to healthy controls. There was a reduction in diversity and loss of commensals in the microbiome of those with acute appendicitis, which may play a role in the cascade leading to acute appendicitis or the result of this.

Keywords: Acute appendicitis; gut microbiome; 16S rRNA sequencing; short chain fatty acids

Introduction

Acute appendicitis is a common acute surgical emergency, with an estimated 17.7 million cases worldwide per year (Wickramasinghe et al., 2021) and a lifetime risk of appendicitis of approximately 1 in 8 (Lee et al., 2010). Multiple studies (Bhangu et al., 2015; Shahmoradi et al., 2021) have attempted to investigate the aetiology of acute appendicitis and yet there is a lack of consensus on the aetiology. Direct luminal obstruction has been the conventional theory for the pathogenesis of acute appendicitis, commonly associated with faecoliths, lymphoid hyperplasia, parasites and very rarely caecal or appendiceal malignancy. Environmental factors such as seasonal alterations in temperature also affect the incidence and course of acute appendicitis (Wolkomir et al., 1987).

Appendix as a potential microbial reservoir, allowing repopulation of the gastrointestinal tract has been theorised but relevant data are scarce (Bhangu et al., 2015). The importance of the gut microbiome is increasingly being recognised in gut-related diseases, especially its role in inflammation (Duvall et al., 2017; Glassner et al., 2020; Jackson et al., 2014; Oh et al., 2020). Several studies using bacterial cultures of

paediatric populations have highlighted the association of various microbiota in the pathogenesis of acute appendicitis (Elhag et al. 1986; Hattori et al., 2019; Swidsinski et al., 2011). The advancement of high throughput sequencing of the human microbiome has led to a growth in knowledge of the microbiome and the role of dysbiosis in disease. Despite this, there has been limited research undertaken on adult appendicitis. The aim of this study is to characterise the microbiome of adult acute appendicitis using 16S rRNA gene sequencing in a prospective cohort.

Methods

Participants

Patients who presented with acute appendicitis and subsequently underwent laparoscopic appendectomy, between March and September 2020, were recruited prospectively at Christchurch Hospital, Canterbury, New Zealand. Demographics and relevant clinical data were collected from electronic hospital health system. Participants were 18 years and older, with acute appendicitis confirmed on histology for inclusion in the study. Patients were excluded if they had antibiotic for more than 24 hours prior to surgery or in the previous 6 weeks. Appendectomies performed in conjunction with concurrent colorectal or gynaecological disease were also excluded.

Rectal swabs were taken using DNA/RNA Shield Collection tubes with swabs (Zymo Research, Irvine, CA). Rectal swab is minimally invasive and is a suitable sampling method as proven from previously published pilot study (Turner et al., 2022). Patients had a rectal swab taken pre-operatively. Healthy volunteers had rectal swabs taken by a single clinician.

Appendicitis is categorised based on histological findings; complicated appendicitis is defined as appendicitis with perforation, gangrene and/or peri-appendicular abscess formation whereas these features are absent in simple/uncomplicated appendicitis. Complicated appendicitis is known to be associated with worse outcomes, prolonged hospitalisation and increased cost and therefore it would be useful to identify the microbiome-based markers, as a predictor of complicated disease.

This study had approval from the University of Otago Human Ethics Committee (H20/045). All participants provided written informed consent upon enrollment.

DNA extraction

All swabs were stored in DNA/RNA lysis tubes (Zymo Research, Irvine, CA) to maintain optimum stability of the DNA. DNA was extracted from samples using ZymoBIOMICS micro-DNA Kits (Zymo Research, Irvine, CA), according to the manufacturer's instructions. In brief, samples homogenised using ZR BashingBead followed by centrifugation at >10,000 rpm for a minute. Supernatant was added to DNA Binding buffer and transferred Zymo-Spin IC-Z Column in a collection tube and centrifuged at maximum for 1 minute. The column was washed several times and DNA eluted in 20 µl DNase/RNase-free water. Finally, DNA concentration and 260/280 nm absorbance were measured using the NanoDrop 2000 spectrophotometer (Thermo Fisher Scientific, Waltham, MA). DNA was stored at -20C prior to 16S rRNA sequencing.

16S rRNA sequencing

Sequencing 16S rRNA region was carried out as per previous publication (Turner et al., 2022). Specifically, Nextera XT DNA Index kit (Illumina, San Diego, CA) was used for library preparation of the V3-V4 hypervariable regions of the 16S rRNA gene, using a single-step PCR with dual index primers (16SF_V3: 5'-TATG GTAATTGGCC TACGGGA GGC AGCAG-3' and 16SR_V4: 5'AGTCAGTCAGCCGGA CTACH VGGGTWCTAAT-3'). Illumina sequencing adaptors and barcodes were then added using limited cycle PCR. The libraries were pooled by equal molarity before

loading on the Illumina MiSeq platform. PhiX was used at 20 per cent of the prepared libraries. Paired-end reads of length 250 bp were generated.

Sequences of appendicitis samples are available from the NCBI Sequencing Read Archive as BioProject PRJNA843336 while sequences of healthy samples are available as Bioproject PRJNA648177.

Microbiome analysis

Quality control

DADA2 (v1.18.0) was used to filter, trim, join sequencing reads, and remove chimeras to obtain amplicon sequence variants (Callahan et al., 2016). The SILVA database (v132) was utilised to assign taxonomy to the amplicon sequence variants (Quast et al., 2013). The taxonomy, sample metadata, and sequences were combined into a phyloseq object for subsequent analysis (McMurdie and Holmes, 2013).

Phylum-level differences

Taxa identified were agglomerated to the Phylum level and percent abundances were obtained per samples. Wilcoxon tests were used to compare Phylum differences between groups. A p -value of <0.05 is considered significant.

Diversity analysis

We rarefied samples to the smallest library size (26,882). We used phyloseq's estimate richness function for Observed (richness) and Shannon (richness and evenness) measures, and compared differences of the alpha diversity measures by Wilcoxon tests. A p -value of <0.05 is considered significant.

We used principal coordinate analysis (PCoA) to test for overall differences in the microbiome between groups, using UniFrac distances between samples.

Differential abundance

DESeq2 was used to identify differentially abundant genera between healthy and appendicitis groups. Only genera that had 10 per cent abundance in 10 per cent of samples from each group were used. We considered a fold-change between comparisons significant if it has adjusted p -value of <0.05 .

The bioinformatics analysis may be accessed at https://gitlab.com/alsulit08/adult_acute_appendicitis.

Results

A total of 60 patients with acute appendicitis and 20 healthy controls were recruited for the study. There were only 58 rectal samples in appendicitis group had sufficient materials for analysis. Of the 78 total samples, we have a total of 7,465,619 reads for analysis after DADA2 quality control, with a range of 26882–195877 reads and median 83,122.5. There were 6,569 amplicon sequence variants (ASVs) identified in the 78 samples. The median age was 35.5 years old in appendicitis group and 32.5 years old in the control group. The age range is between 18 and 74 years old. In the appendicitis group, participants were predominantly male with NZ European ethnicity. There were 31 patients with simple appendicitis and 20 patients (41 per cent) with complicated appendicitis (perforated and/or gangrenous). Patients were older and had slightly higher inflammatory markers in the complicated appendicitis subgroup (Table 1).

Comparison of the microbiome of acute appendicitis and healthy group

Bioinformatic analysis showed a statistically significant difference in the alpha diversity between the groups ($p < 0.0001$), with samples from healthy controls having higher alpha diversity. Unweighted

Table 1. Demographics of the appendicitis group.

	Simple appendicitis (n = 31)	Complicated appendicitis (n = 20)	p-value (Fisher's test)
Median age	32 years old	47 years old	
Gender – M:F ratio	1.8:1	1:2	
Male	20	7	0.049
Female	11	13	
Smoker			
Current	6	5	
Ex-smoker	8	8	0.25
Never	17	7	
Preoperative imaging			
CT scan	15	13	0.26
Ultrasound (USS)	10	4	
None	6	3	
Length of stay (mean)	2.4 days	2.5 days	
Readmission			
Yes	2	2	1.0
No	29	18	
Complications			
None	28	19	0.747
Grade 1 and 2	1	1	
Grade 3 and 4	2	0	
Laboratory (median)			
White Cell Counts (WCC)	11.9	13.1	
Neutrophils	9.5	11	
C-Reactive Protein (CRP)	27	40	

UniFrac analysis showed clustering of samples by appendicitis or health controls (Figure 1A). Wilcoxon tests on phyla between healthy and appendicitis sample showed a significant difference of *Firmicutes*, *Actinobacteria* and *Proteobacteria* phyla percent abundance (Figure 1B). Further classification at the genus level show loss of beneficial gut commensals, including *Blautia*, *Lachnospiraceae*, *Faecalibacterium* and *Ruminococcus* in swabs from appendicitis patients, compared to healthy controls (Figure 1C).

Comparison of the microbiome between simple and complicated appendicitis

Shannon diversity analysis showed no significant differences in alpha diversities between simple and complicated appendicitis in rectal samples ($p = 0.98$) (Figure 2). Phylum-level abundance were similar for both simple and complicated acute appendicitis (Figure 3). Further classification at the genus level showed no difference between the two groups.

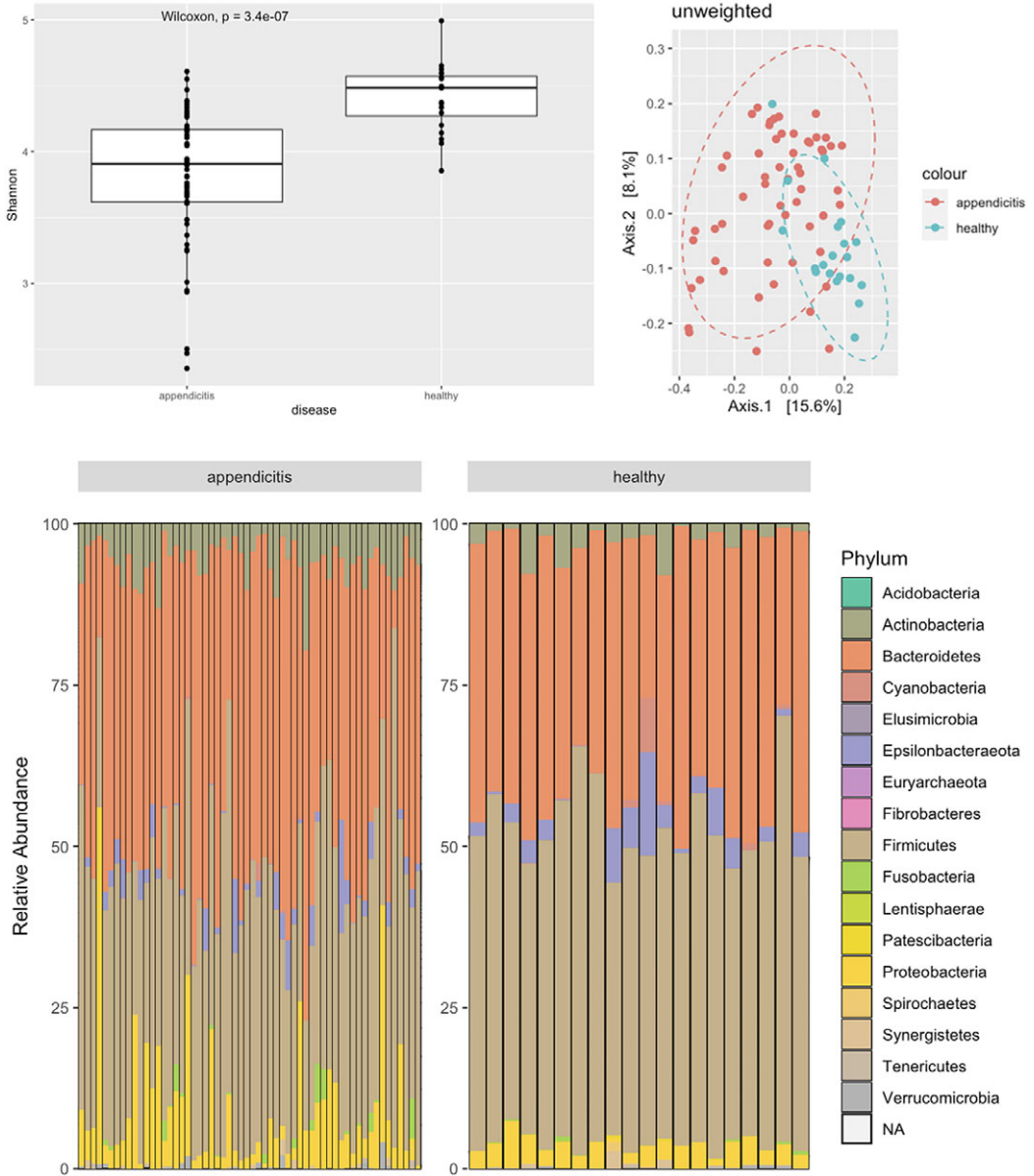


Figure 1. (A) Alpha diversity and PCoA of rectal swabs between appendicitis and healthy controls. (B) Phyla abundance between the appendicitis and healthy groups. (C) Comparison between appendicitis and healthy samples at genus level. A positive log fold change means the genus more abundant in appendicitis and a negative log fold means the genus is more abundant in healthy samples.

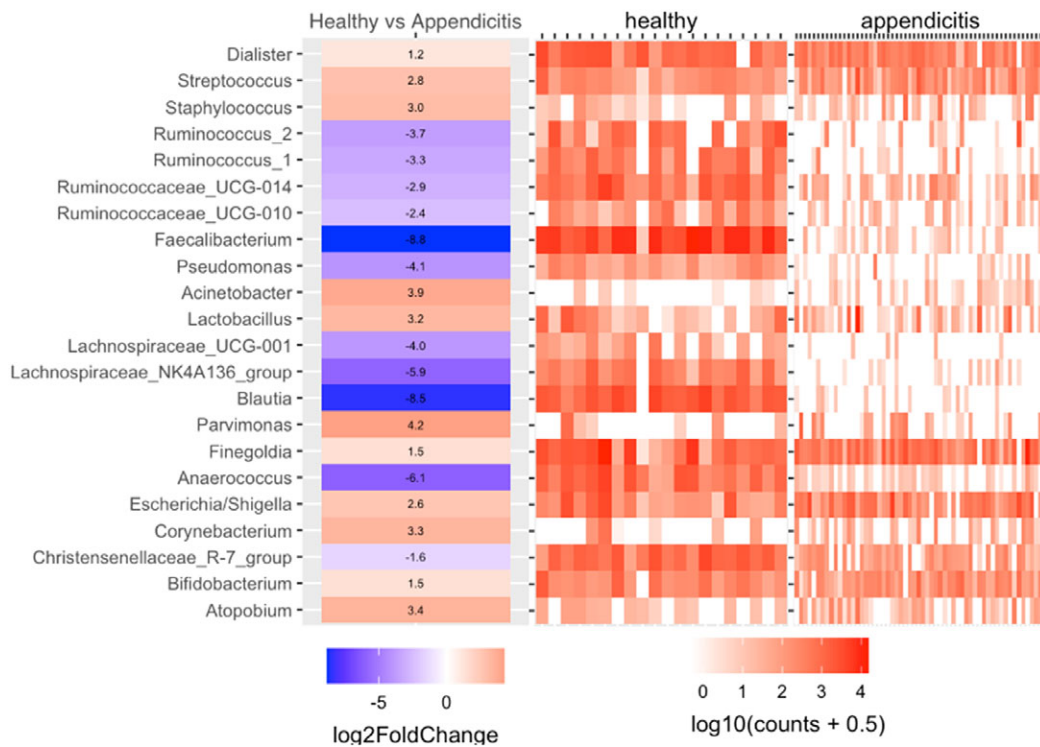


Figure 1. (Continued)

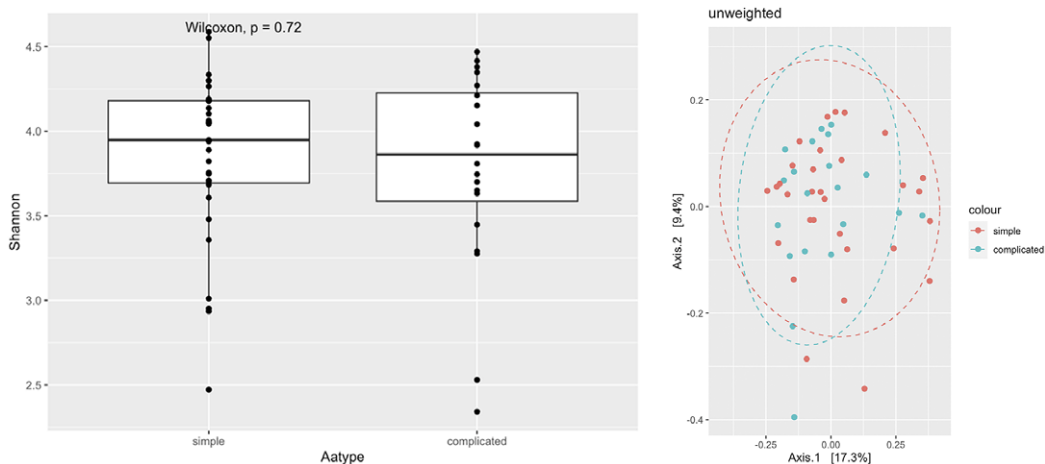


Figure 2. Alpha diversity and PCoA of rectal swabs comparing simple and complicated appendicitis.

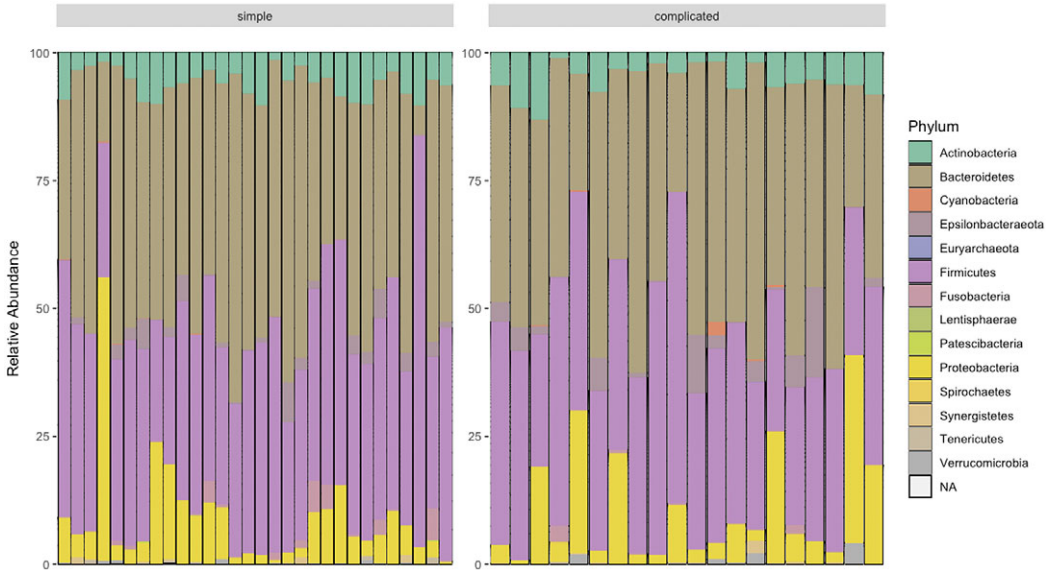


Figure 3. Phyla distribution between simple and complicated appendicitis.

Discussion

In this study, there was a difference between the microbial composition as assessed by rectal swabs of those with acute appendicitis and healthy controls. Metagenomics sequencing analysis showed a reduction in diversity and loss of commensal bacteria in people with appendicitis. The loss of commensals including phyla important to gut homeostasis, such as *Lachnospiraceae*, *Blautia* and *Faecalibacterium*, which may play a role in the cascade leading to the development of acute appendicitis. There were no significant microbiome differences found at both phylum and genus level, between patients with simple and complicated appendicitis.

Despite the frequency of acute appendicitis, we know very little for certain about its aetiology. Microbiological causes have been suggested in various studies. Guinane et al. (2013) first provided insight into the microbial composition of the appendix in a study of seven patients, aged 5–25 years old, using 16S-rRNA sequencing. Analysis of faecal and tissue samples revealed that appendices contained a high bacterial biomass and were postulated to be a reservoir for commensal bacterial. Across the seven samples, 15 different phyla were detected, dominated by five major phyla – Firmicutes, Proteobacteria, Bacteroidetes, Actinobacteria and Fusobacteria. In our study of an adult cohort, we found similar dominant phyla with the exception of Fusobacteria, due to different sampling methodologies.

Rectal swabs were used as these can be taken without delay when the patient is admitted, unlike faecal samples, where there could be a delay of up to 24 hours for sample collection. This is important as patients are routinely administered antibiotics on admission and antibiotics have been shown in multiple studies (Dudek-Wicher et al., 2018; Mu and Zhu, 2019) to affect the gut microbiome. Previously published findings from our pilot study (Turner et al., 2022) and others (Budding et al., 2014; Reyman et al., 2019; Short et al., 2021) have shown that rectal swabs are a suitable method of analysing changes in the gut microbiome between patient groups.

Several other paediatric studies of appendicitis suggest that *Fusobacterium* is associated with acute appendicitis (Salo et al., 2017; Schulin et al., 2017; Swidsinski et al., 2012; Zhong et al., 2014). Salo et al. (2017) noticed an increased abundance of *Fusobacterium* and a decreased in *Bacteroides* in complicated appendicitis compared to uncomplicated appendicitis. A further study of intraluminal samples by Schulin et al. (2017) reported that *Fusobacterium necrophorum* was mainly found in catarrhal

appendicitis, *Pseudomonas endodontalis* in phlegmonous appendicitis and *F. nucleatum* in gangrenous appendicitis. In contrast, in our study, *Fusobacterium* was detected, but was not a predominant phylum in either healthy or appendicitis groups. These findings may be due to differences between paediatric populations in these other publications and the adult population under investigations here; an adult population may have more resilient microbiome and dysbiosis may not be as pronounced.

A growth in human microbiome research has led to better understanding of disease associated microbiome change. A study on the aetiology of appendicitis in 277 children showed that *Escherichia coli* was the predominant bacteria followed by *Streptococci* (Richardson et al., 2016). In addition, Zhong et al. (2014) performed the 16S ribosomal gene sequence analysis to profile the microbiota of the luminal fluid of the appendix in children with appendicitis and normal appendices, and found an increased relative abundance of the pathogen containing genera, *Fusobacterium* and *Prevotella* with a corresponding depletion of *Bacteroides* (11.4 vs. 52.9 per cent). In our study, *Actinobacteria* and *Proteobacteria* phyla are increased in appendicitis patients with a concomitant increase in the pathogen containing *Parvimonas*, *Acinetobacter*, *E. coli*, *Cornebacterium* and *Streptococci* genera (Figure 1B,C). The apparent differences in microbiome changes between paediatric and adult population might be due to different methodologies in the various studies or other factors (ie. dietary or lifestyle factors) which may influence the dysbiosis in acute appendicitis.

Analysis of differential abundance at the genus level in our study also revealed a loss of commensal bacteria in appendicitis samples compared to healthy samples. These included *Blautia*, *Faecalibacterium* and *Lachnospiraceae*, genera that contain short-chain fatty acids (SCFA) producing bacteria. SCFAs are important metabolites that are an energy source for epithelial cells and also contribute to gut barrier function (Loh and Blaut, 2012; Parada Venegas et al., 2019) and play an anti-inflammatory role in the gut (Blaak et al., 2020; Machiels et al., 2014; Segain et al., 2000). Our observed reduction of microbial diversity has been described in other inflammatory conditions such as inflammatory bowel disease and acute appendicitis (Loh and Blaut, 2012). Ott (2004) performed a 16S rDNA study of mucosa associated colonic microflora and found a reduction of microbial diversity in both Crohn's disease and Ulcerative colitis. This demonstrates that gut inflammation has a strong association with changes in gut microbiome. This preliminary study looks at the overall changes in pattern of bacterial dysbiosis in relation to gut inflammation, neither at individual bacteria level nor specific aetiology for acute appendicitis.

This is one of the few published studies on the microbiome of acute appendicitis in adult population. This preliminary study looked at broad changes in taxa in the gut microbiomes between patients with acute appendicitis and healthy controls, and between patients with complicated versus uncomplicated appendicitis, using a well established method of analysis (16S rRNA sequencing). We have used rectal swabs instead of faecal sampling, a methodology we have previously verified (Turner et al., 2022). The methodology of this study does not however allow us to determine whether gut microbiome dysbiosis was a cause of appendicitis or just secondary to the pathology. Despite the relatively large cohort size in this study, future large-scale studies are necessary to validate the findings presented here and suggest perform functional analysis to establish bacterial signatures associated with acute appendicitis.

Conclusion

In conclusion, this study has added knowledge to the current literature on microbiome studies of acute appendicitis in adult population. We have found a reduction in diversity and loss of commensals in the microbiome of those with acute appendicitis. The loss of commensal bacteria may play a role in the cascade leading to the development of acute appendicitis.

Acknowledgement. The authors would like to thank University of Otago administrative staffs.

Notes on contributors. The authors thank our University of Otago Statistician and Laboratory staffs.

Disclosure statement. The authors have no known conflicts of interest to report.

Author contributions. Conceptualisation: F.F.; Data curation: M.S.L.; Formal analysis: A.S., R.P.; Investigations: A.S., R.P.; Methodology: M.S.L., A.S., R.P.; Project administration: F.F.; Resources: M.S.L.; Software: A.S.; Supervision: R.P., F.F.; Visualisation: M.S.L., A.S.; Writing – original draft: M.S.L.; Writing – review and editing: F.F., R.P.

Research Transparency and Reproducibility. Protocols, materials and relevant data are available for readers and can be accessed by contacting corresponding author.

References

- Bhangu A, Søreide K, Di Saverio S, Assarsson JH and Drake FT** (2015) Acute appendicitis: Modern understanding of pathogenesis, diagnosis, and management. *The Lancet* **386**(10000), 1278–1287. [https://doi.org/10.1016/s0140-6736\(15\)00275-5](https://doi.org/10.1016/s0140-6736(15)00275-5)
- Blaak EE, Canfora EE, Theis S, Frost G, Groen AK, Mithieux G, Nauta A, Scott K, Stahl B, van Harsselaar J, van Tol R, Vaughan EE and Verbeke K** (2020) Short chain fatty acids in human gut and metabolic health. *Beneficial Microbes* **11**(5), 411–455. <https://doi.org/10.3920/BM2020.0057>
- Budding AE, Grasman ME, Eck A, Bogaards JA, Vandenbroucke-Grauls CM, van Bodegraven AA and Savelkoul PH** (2014) Rectal swabs for analysis of the intestinal microbiota. *PLoS One* **9**(7), e101344. <https://doi.org/10.1371/journal.pone.0101344>
- Callahan BJ, McMurdie PJ, Rosen MJ, Han AW, Johnson AJ and Holmes SP** (2016) DADA2: High-resolution sample inference from Illumina amplicon data. *Nature Methods* **13**(7), 581–583. <https://doi.org/10.1038/nmeth.3869>
- Dudek-Wicher RK, Junka A and Bartoszewicz M** (2018) The influence of antibiotics and dietary components on gut microbiota. *Przegląd Gastroenterologiczny* **13**(2), 85–92. <https://doi.org/10.5114/pg.2018.76005>
- Duvallet C, Gibbons SM, Gurry T, Irizarry RA and Alm EJ** (2017) Meta-analysis of gut microbiome studies identifies disease-specific and shared responses. *Nature Communications* **8**(1784), 1–10. <https://doi.org/10.1038/s41467-017-01973-8>
- Elhag KM, Alwan MH, Al-Adnani MS, Sherif RA.** Bacteroides fragilis is a silent pathogen in acute appendicitis. *Journal of Medicine Microbiome*. 1986 May; **21**(3); 245–9.
- Glassner KL, Abraham BP and Quigley EMM** (2020) The microbiome and inflammatory bowel disease. *Journal of Allergy and Clinical Immunology* **145**(1), 16–27. <https://doi.org/10.1016/j.jaci.2019.11.003>
- Guinane CM, Tadrous A, Fouhy F, Ryan CA, Dempsey EM, Murphy B, Andrews E, Cotter PD, Stanton C and Ross RP** (2013) Microbial composition of human appendices from patients following appendectomy. *MBio* **4**(1), 1–6. <https://doi.org/10.1128/mBio.00366-12>
- Hattori T, Yuasa N, Ikegami S, Nishiyama H, Takeuchi E, Miyake H, Kuno R, Miyata K, Fujino M and Minami M** (2019) Culture-based bacterial evaluation of the appendix lumen in patients with and without acute appendicitis. *Journal of Infection & Chemotherapy* **25**(9), 708–713. <https://doi.org/10.1016/j.jiac.2019.03.021>
- Jackson HT, Mongodin EF, Davenport KP, Fraser CM, Sandler AD and Zeichner SL** (2014) Culture-independent evaluation of the appendix and rectum microbiomes in children with and without appendicitis. *PLoS One* **9**(4), e95414. <https://doi.org/10.1371/journal.pone.0095414>
- Lee JH, Park YS and Choi JS** (2010) The epidemiology of appendicitis and appendectomy in South Korea: National registry data. *Journal of Epidemiology* **20**(2), 97–105. <https://doi.org/10.2188/jea.je20090011>
- Loh G and Blaut M** (2012) Role of commensal gut bacteria in inflammatory bowel diseases. *Gut Microbes* **3**(6), 544–555. <https://doi.org/10.4161/gmic.22156>
- Machiels K, Joossens M, Sabino J, De Preter V, Arijs I, Eeckhaut V, Ballet V, Claes K, Van Immerseel F, Verbeke K, Ferrante M, Verhaegen J, Rutgeerts P and Vermeire S** (2014) A decrease of the butyrate-producing species *Roseburia hominis* and *Faecalibacterium prausnitzii* defines dysbiosis in patients with ulcerative colitis. *Gut* **63**(8), 1275–1283. <https://doi.org/10.1136/gutjnl-2013-304833>
- McMurdie PJ and Holmes S** (2013) Phyloseq: An R package for reproducible interactive analysis and graphics of microbiome census data. *PLoS One* **8**(4), e61217. <https://doi.org/10.1371/journal.pone.0061217>
- Mu C and Zhu W** (2019) Antibiotic effects on gut microbiota, metabolism, and beyond. *Applied Microbiology and Biotechnology* **103**(23–24), 9277–9285. <https://doi.org/10.1007/s00253-019-10165-x>
- Oh SJ, Pimentel M, Leite GGS, Celly S, Villanueva-Millan MJ, Lacsina I, Chuang B, Parodi G, Morales W, Weitsman S, Singer-Englar T, Barlow GM, Zhai J, Pichesthote N, Rezaie A, Mathur R and Pimentel M** (2020) Acute appendicitis is associated with appendiceal microbiome changes including elevated campylobacter jejuni levels. *BMJ Open Gastroenterology* **7**(1), 1–10. <https://doi.org/10.1136/bmjgast-2020-000412>
- Ott SJ** (2004) Reduction in diversity of the colonic mucosa associated bacterial microflora in patients with active inflammatory bowel disease. *Gut* **53**(5), 685–693. <https://doi.org/10.1136/gut.2003.025403>
- Parada Venegas D, De la Fuente MK, Landskron G, Gonzalez MJ, Quera R, Dijkstra G, Harmsen HJM, Faber KN and Hermoso MA** (2019) Short chain fatty acids (SCFAs)-mediated gut epithelial and immune regulation and its relevance for inflammatory bowel diseases. *Frontiers in Immunology* **10**(277), 1–16. <https://doi.org/10.3389/fimmu.2019.00277>
- Quast C, Pruesse E, Yilmaz P, Gerken J, Schweer T, Yarza P, Peplies J and Glockner FO** (2013) The SILVA ribosomal RNA gene database project: Improved data processing and web-based tools. *Nucleic Acids Research* **41**, D590–D596. <https://doi.org/10.1093/nar/gks1219>

- Reyman M, Van Houten MA, Arp K, Sanders EAM and Bogaert D (2019) Rectal swabs are a reliable proxy for faecal samples in infant gut microbiota research based on 16S-rRNA sequencing. *Scientific Reports* **9**, 16072. <https://doi.org/10.1038/s41598-019-52549-z>
- Richardson I, Schob DS, Ulmer TF, Steinau G, Neumann UP, Klink CD and Lambert A (2016) Etiology of appendicitis in children: The role of bacterial and viral pathogens. *Journal of Investigative Surgery* **29**(2), 74–79. <https://doi.org/10.3109/08941939.2015.1065300>
- Salo M, Marungruang N, Roth B, Sundberg T, Stenstrom P, Arnbjornsson E, Fak F and Ohlsson B (2017) Evaluation of the microbiome in children's appendicitis. *International Journal of Colorectal Disease* **32**(1), 19–28. <https://doi.org/10.1007/s00384-016-2639-x>
- Schulin S, Schlichting N, Blod C, Opitz S, Suttikus A, Stingu CS, Barry K, Lacher M, Buhlig U and Mayer S (2017) The intra- and extraluminal appendiceal microbiome in pediatric patients: A comparative study. *Medicine (Baltimore)* **96**(52), e9518. <https://doi.org/10.1097/MD.00000000000009518>
- Segain JP, de la Bletiere DR, Bourreille A, Leray V, Gervois N, Rosales C, Ferrier L, Bonnet C, Blottiere HM and Galmiche JP (2000) Butyrate inhibits inflammatory responses through NFκB inhibition: Implications for Crohn's disease. *Gut Journal* **47**, 397–403.
- Shahmoradi MK, Zarei F, Beiranvand M and Hosseinnia Z (2021) A retrospective descriptive study based on etiology of appendicitis among patients undergoing appendectomy. *International Journal of Surgery Open* **31**, 100326. <https://doi.org/10.1016/j.ijso.2021.100326>
- Short MI, Hudson R, Besasie BD, Reveles KR, Shah DP, Nicholson S, Johnson-Pais TL, Weldon K, Lai Z, Leach RJ, Fongang B and Liss MA (2021) Comparison of rectal swab, glove tip, and participant-collected stool techniques for gut microbiome sampling. *BMC Microbiology* **21**(1), 26. <https://doi.org/10.1186/s12866-020-02080-3>
- Swidsinski A, Dorffel Y, Loening-Baucke V, Theissig F, Ruckert JC, Ismail M, Rau WA, Gaschler D, Weizenegger M, Kuhn S, Schilling J and Dorffel WV (2011) Acute appendicitis is characterised by local invasion with fusobacterium nucleatum/necrophorum. *Gut* **60**(1), 34–40. Available at <https://ovidsp.ovid.com/ovidweb.cgi?T=JS&CSC=Y&NEWS=N&PAGE=fulltext&D=med8&AN=19926616>.
- Swidsinski A, Loening-Baucke V, Biche-ool S, Guo Y, Dörffel Y, Tertychnyy A, Stonogin S and Sun N-D (2012) Mucosal invasion by fusobacteria is a common feature of acute appendicitis in Germany, Russia, and China. *Saudi Journal of Gastroenterology* **18**(1), 55–58. <https://doi.org/10.4103/1319-3767.91734>
- Turner G, O'Grady M, Hudson D, Morgan X, Frizelle F and Purcell R (2022) Rectal swabs are a reliable method of assessing the colonic microbiome. *International Journal of Medical Microbiology* **312**(2), 151549. <https://doi.org/10.1016/j.ijmm.2022.151549>
- Wickramasinghe DP, Xavier C and Samarasekera DN (2021) The worldwide epidemiology of acute appendicitis: An analysis of the global health data exchange dataset. *World Journal of Surgery* **45**(7), 1999–2008. <https://doi.org/10.1007/s00268-021-06077-5>
- Wolkomir A, Kornak P, Elsagr M and McGovern P (1987) Seasonal variation of acute appendicitis: A 56-year study. *Southern Medical Journal* **80**(8), 958–960. 5th March 2022 Available at <https://ovidsp.ovid.com/ovidweb.cgi?T=JS&CSC=Y&NEWS=N&PAGE=fulltext&D=med2&AN=3616723>.
- Zhong D, Brower-Sinning R, Firek B and Morowitz MJ (2014) Acute appendicitis in children is associated with an abundance of bacteria from the phylum fusobacteria. *Journal of Pediatric Surgery* **49**(3), 441–446. <https://doi.org/10.1016/j.jpedsurg.2013.06.026>

Cite this article: Lee M.S., Sulit A., Frizelle F., and Purcell R. 2022. The microbiome in adult acute appendicitis. *Gut Microbiome*, **3**, e8, 1–10. <https://doi.org/10.1017/gmb.2022.7>