

Research Article

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Exploring the nutritional potentials of wild *Vigna* legume species for neo-domestication prospects

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Abstract

Projected increases in human population suggest that 70% more food will be needed in the near future, this makes it imperative to search for alternative food and feed sources for human and animal nutrition to feed the exponentially growing human population. According to the FAO 2019 report, the immense challenge of achieving the Zero Hunger target by 2030 is persistent. Exploring the unexplored, refining unrefined traits, cultivating the uncultivated, and popularizing the unpopular remain the most adequate steps proposed by researchers to achieve the domestication of the undomesticated for food and nutrition security. In that line of thought, this study explored the proximate composition of 87 accessions of four wild unexplored *Vigna* species (*V. racemosa*, *V. ambacensis*, *V. reticulata*, *V. vexillata*) in order to reveal information leading to their future domestication and utilization. Standard procedures and methods approved by the Association of Official Analytical Chemists were used in carrying out the proximate composition (%protein, %lipid, %fibre, %ash and % moisture and % carbohydrate) of the wild *Vigna* legumes. The study revealed that the wild *Vigna* species possess a large variation range of nutrient characteristics which could be exploited in the improvement of domesticated species or guide their domestication. It was also found that some individual wild accessions have higher nutrient, content as compared with domesticated ones which could be advantageous for bio-fortification or domestication. Indications relating to the candidate accessions favourable for domestication, based on the nutrient characteristics were revealed.

Introduction

More than 820 million people in the world were still hungry in 2018, underscoring the immense challenge of achieving the Zero Hunger target by 2030 (FAO *et al.*, 2019). Additionally, it is unanimously recognized by many researchers and organizations that only 12 crops contribute most to the current global food production, with only three of them (rice, wheat and maize) providing more than 50% of the world's calories (Singh *et al.*, 2019). A rapid reduction of the gene pool in both plant and animal genetic resources is observed as only a dozen species of animals provide 90% of the animal protein consumed globally and just four crop species provide half of the plant-based calories in the human diet (FAO, 2009). Hence, directing researchers attention to plant protein sources becomes a necessity as the animal protein sources face challenges of preference (for vegetarians) and affordability. As such, the contribution of legumes as plant protein source is indubitable and domestication of new species is becoming an imperative due the challenges faced by the domesticated ones (Harouna *et al.*, 2018; Singh *et al.*, 2019; Takahashi *et al.*, 2019). Legumes (family: Fabaceae) constitute the third largest family of flowering plants (Bhat and Karim, 2009). It is sometimes thought that legumes have very important nutritional value for both humans and animals and are referred to as the 'poor man's meat'. The most commonly domesticated grown and commercialized legumes such as soybeans, cowpeas, common beans and others have demonstrated considerable contribution to the global food security (Harouna *et al.*, 2018). *Phaseolus* and *Vigna* genera comprise the most widely consumed legumes, namely common beans (*Phaseolus vulgaris* L.) and cowpea (*Vigna unguiculata* (L.) Walp) (Garcia *et al.*, 1997; Gepts, 2001; Harouna *et al.*, 2018). Within each genus, there are fewer domesticated edible species than the non-domesticated wild species. Some domesticated and semi-domesticated species are being neglected and underutilized species despite the persistent decrease of the genetic diversity in the food systems. The little attention



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paid to such species, coupled to the complete ignorance of their existence by agricultural researchers, plant breeders and policy-makers has contributed enormously to their under-exploitation and utilization (Padulosi *et al.*, 2013). A deep exploration into these under-exploited species diversity could be a significantly considerable alternative option that would contribute to the mitigation of food and nutrition insecurity. The *Vigna* genus comprises more than 100 wild species, which do not have common names, apart from their scientific appellation (Tomooka *et al.*, 2014). They are given different denotations, such as the under-exploited wild *Vigna* species, undomesticated *Vigna* species, wild *Vigna* or alien species, depending on the scientist (Pratap *et al.*, 2014; Harouna *et al.*, 2018). Preliminary investigations on *V. racemosa* (G. Don) Hutch. & Dalziel, *V. ambacensis* Welw. ex Baker, *V. reticulata* Hook.f. and *V. vexillata* (L.) A.Rich. accessions were carried out in this study to unveil their proximate composition potential based on their availability and accessibility in the nearest gene bank. Earlier reports have demonstrated their potential usages, farmers' preferences, cooking and agro-morphological characteristics (Harouna *et al.*, 2019a; Harouna *et al.*, 2020). It is very important to note here that investigations focusing on the chemical composition of wild *Vigna* species accessions are scanty or not well documented. The proximate composition, fatty acid composition, total phenolic content, antioxidant activity and amino acid profile of an unknown accession of *V. racemosa* were reported in a recent study (Ade-Omowaye *et al.*, 2015). Another study also reported recently on the chemical changes during open and controlled fermentation of *V. racemosa* seed collected from their natural environment, regardless of their genetic specification (Difo *et al.*, 2015). Other studies focusing on qualitative evaluation of bioactive compounds of *V. kirkii* (Baker) Gillett, Kew Bull, *V. marina* (Burm.) Merr., *V. gracilis* (Guill. & Perr.) Hook. f., *V. heterophylla* A.Rich., *V. parkeri* Baker, *V. hosei* (Craib) Backer, *V. adenantha* (G.Mey.) Maréchal, Mascherpa & Stainier, *V. venusta* (Piper) Marichal, Mascherpa & Stainier, *V. minima* (Roxb.) Ohwi & H. Ohashi, *V. glabrascens* Maréchal, Mascherpa & Stainier and *V. triphylla* (R.Wilczek) Verdc. have revealed the presence of biochemicals such as Robinin, Kaempferol-3- rutinoside, Isorhamnetin-3- rutinoside, Hyperoside, Delphinidin and Cyanidin (Lattanzio *et al.*, 1997; Macorni *et al.*, 1997; Bravo *et al.*, 1999). Many other biochemical parameters of these wild *Vigna* species also need to be investigated to enhance their usages. Therefore, this study aimed at exploring the proximate composition of some wild *Vigna* species (*V. racemosa*, *V. ambacensis*, *V. reticulata* and *V. vexillata*) in order to detect accessions for potential domestication and/or crop improvement.

Materials and methods

Seeds preparation for proximate composition analysis

One hundred and six (106) accessions of the four species of wild *Vigna* legumes (*V. racemosa*, *V. ambacensis*, *V. reticulata* and *V. vexillata*) were obtained from genebanks as presented in Table S4. Approximately 20–100 seeds of each accession were supplied by the genebanks and planted in an experimental plot, following the augmented block design arrangement, and allowed to grow until full maturity as described in an earlier study (Harouna *et al.*, 2020).

The seeds were planted at the Tanzania Agricultural Research Institute (TARI), Selian in the Arusha region, located in the northern part of Tanzania. TARI-Selian lies at a latitude of

3°21'50.08" N and longitude of 36°38'06.29" E at an elevation of 1390 m above sea level (a.s.l.). Eighty seven (87) accessions of matured seeds of wild *Vigna* species of legumes harvested were selected based on their productivity in the field to carry out the proximate composition. In addition to the wild accessions, three domesticated *Vigna* legumes that is, cowpea (*V. unguiculata*), rice bean (*V. umbellata*), and a semi-domesticated landrace (*V. vexillata*) were used as checks. The checks were obtained from the Genetic Resource Center (GRC-IITA), Nigeria (cowpea), the National Bureau of Plant Genetic Resources (NBPGR), India (rice bean), and the Australian Grain Gene bank (AGG), Australia (semi-domesticated landrace *V. vexillata*).

The matured fruits were harvested with their pods, sun-dried and the seeds were removed from the pods, threshed and winnowed, then free from broken seeds, dust and other foreign materials to obtain clean seeds. The seeds were then stored in a plastic bags at room temperature (27 °C – 30 °C) for subsequent analysis. After that, the seed samples were grinded using a kitchen blender (3 in 1 Electric Chopper Juice Blender HB-38, 350W, Jar Capacity: 1.5L, Guangdong, China) and sieved, and the 1 mm fraction were collected for analysis.

Moisture content determination

The method employed for the determination of moisture content in the sample based on the measurement of the loss in weight due to drying at a temperature of about 105 °C as describe in the Association of Official Analytical Chemists (AOAC) methods (method 950.46) (AOAC, 2000). A watch glass was washed and dried in an oven (DRY-Line 56, STEP Systems GmbH, Nuremberg, Germany) at about 105 °C for 3 h, it was cooled in a desiccator and weighed empty.

About 2.0 g of sample was weighed into a clean watch glass. The watch glass and its content were dried in an Air-circulated oven (DRY-Line 56, STEP Systems GmbH, Nuremberg, Germany) at about 105 °C to constant weight. The watch glass and its content was cooled in a desiccator and reweighed.

The percentage moisture was obtained using the expression below;

$$\% \text{ Moisture} = \frac{\text{Loss in weight on drying} \times 100}{\text{Initial sample weight (g)}}$$

Ash content determination

The term ash refers to the residue left after combustion of the oven dried sample and is a measure of the total mineral content. Determination of the ash content was done according to AOAC method 923.03 (AOAC, 2000).

Three different crucibles were preheated in a Muffle furnace (Nabertherm GmbH, Lilienthal, Germany) at about 550 °C. Each crucible was cooled in a desiccator and weighed. Approximately 2.0 g of each sample was weighed into different crucibles. The crucibles and their contents were transferred into a Muffle furnace (Nabertherm GmbH, Lilienthal, Germany) set at 550 °C and allowed to stay for 6 h. After cooling the heated crucibles, the weights of crucibles and their content were taken and recorded. The percentage ash was calculated using the following expression;

$$\% \text{ Ash} = \frac{\text{weight of ash (g)}}{\text{weight of dry sample}} \times 100$$

Crude lipid content determination

The crude lipid content of wild *Vigna* legumes samples was determined according to the AOAC method 960.39. A Soxtec™ extraction system (Model 2043 Extraction Unit; Tecator, Sweden), and 30 ml of Petroleum ether (Mallinckrodt, Paris, KE, USA) were used to extract the oil from the samples. The amount of extracted oil was determined gravimetrically.

The percentage of lipid was obtained following the equation below;

$$\% \text{ Lipid} = \frac{\text{Weight}_{(\text{extraction cup} + \text{residue})} - \text{Weight}_{(\text{extraction cup})}}{\text{weight of dry sample}} \times 100$$

Crude protein content determination

The protein content of the wild *Vigna* legume samples was analysed according to the AOAC method 928.08 (AOAC, 2000). The samples were digested with concentrated sulphuric acid (Pharmco-AAPER, USA), Hydrogen peroxide (Fisher Scientific, Fair Lawn, NJ, USA), and two Kjeldahl catalyst tablets (FisherTab ST-35; Fisher Scientific, Sweden) using a Kjeltac block digester unit (Model 2020 Digester; Tecator, Sweden). The total nitrogen amount in the sample was determined by distillation and titration of the extracts using a Kjeltac instrument (Kjeltic™ 8200 Auto Distillation Unit) (Ng, Dunford, and Chenault, 2008). A conversion factor of 6.25 was used to convert the amount of nitrogen to amount of protein present in the samples.

The amount of protein in the samples (dry basis) is calculated from the following formula:

$$\% \text{ Protein} = \frac{(T - B) \times M \times 14.007 \times 100 \times 6.25 \times MCF}{\text{weight of dry sample (W)}}$$

where, **T**, Volume of the standard hydrochloric acid used in the sample titration. **B**, Volume of the standard hydrochloric acid used in the blank titration. **M**, Molarity of the acid in four decimal places. **W**, mass of the sample used in the determination in milligrams. **6.25**, factor used to convert per cent N to per cent crude protein. Most proteins contain 16% N, so the conversion factor is 6.25 (100/16 = 6.25). **MCF**, Moisture Correction Factor = 100/(100 - % Moisture).

Crude fibre content determination

The fibre content was evaluated using AOAC method (AOAC, 2000). Fat-free grinded wild *Vigna* samples of 1.0 g were weighed into a clean pre-weighed crucible. The crucible with sample was then transferred into the hot-extraction unit (Fibertec M6, 200–230 V, FOSS, Denmark) and the sample was left to digest for 30 min with 150 ml of solution containing 12.5% Sulphuric acid (Sigma Aldrich, Germany) and 0.25 ml of octanol (Sigma Aldrich, Germany). The condenser was switched off after 30 min and allowed to cool. The acid solution was filtered and washed with hot distilled water using suction. Then the samples were digested for 30 min with 150 ml alkali solution (12.5% NaOH) (Sigma Aldrich, Germany) and 0.25 ml of octanol to dissolve the alkali-soluble matter from the samples. The porcelain crucibles' final residues were dried at 105 °C in an oven (DRY-Line 56, STEP Systems GmbH, Nuremberg, Germany) for 1 h, cooled in a desiccator and then weighed. The final residues were dried at 105 °C in an oven for 60 min. The residues were ignited in a pre-heated Muffle furnace (Carbolite, UK) at 550 °C for 3 h and weighed. The per cent of crude fibre content

was calculated using the following equation:

$$\% \text{ Fiber} = \frac{W_2 - W_3}{W_1} \times 100$$

where, **W₁**, Sample weight. **W₂**, Crucible weight with ash. **W₃**, Empty crucible weight.

Carbohydrate content determination

The percentage carbohydrate was obtained by difference (AOAC, 2000)

Percentage carbohydrate = 100 - (% Moisture + %Protein + % Fat + %Ash + %Crude fiber)

Data analysis

The data on proximate composition parameters were collected in triplicate. Planned, single degree of freedom F test (orthogonal test) was used for mean separation. The one-way analysis of variance, the hierarchical clustering analysis and principal component analysis (PCA) were performed using XLSTAT.

Results

Proximate composition domesticated *Vigna* species used as checks

The proximate composition for the various *Vigna* species studied is summarized in Tables 1–4. The proximate composition of the three domesticated legumes included in this study for comparison is presented in the tables presenting the results for each species in order to ease the appreciation. The three domesticated legumes used here as checks are: a semi-domesticated landrace of *V. vexillata* (Check 1), cowpea (Check 2), and rice bean (Check 3). A keen examination of the proximate composition of these three checks shows that there is no significant difference in lipid, fibre and carbohydrate content of Check 1 and Check 2, which are significantly different ($P < 0.05$) from that of Check 3. Their lipid content is significantly higher than that of Check 3 while their carbohydrate and fibre contents are lower than that of Check 3. The ash and moisture contents of the three checks are significantly apparently similar. This can be elucidated by evaluating the individual minerals. The protein content of the three checks is significantly different with Check 1 having the highest protein content (Table 1–4).

Proximate composition of *Vigna ambacensis* accessions

Table 1 summarizes the proximate composition of *V. ambacensis* accessions (11 accessions). It shows that the lipid content of all the wild accessions is significantly similar to those of Check 1 and 2 while it is significantly ($P < 0.05$) higher than that of Check 3. All the accessions showed significant lower ash content than the three checks except for accessions TVNu219 and TVNu877 which is comparable to that of the checks. The moisture and protein content of the wild accessions are significantly lower than that of the checks while the carbohydrate and fibre content of the wild accessions were higher than that of the checks.

Proximate composition of *Vigna racemosa* accessions

Table 2 summarizes the proximate composition of *V. racemosa* accessions. It was found that the lipid content of AGG53597WVIG1 and AGG51603WVIG1 accessions was significantly comparable to that

Table 2. Proximate composition of *Vigna racemosa* accessions (g/100)

Accessions	Lipid	Ash	Moisture	Protein	Fiber	Carbohydrates	100-Seed weight (g)
Check 1	1.127 ± 0.128a	3.729 ± 0.126b	11.052 ± 0.281cde	27.313 ± 0.597bc	3.654 ± 0.832c	53.125 ± 1.323c	7.038
Check 2	1.215 ± 0.150a	3.716 ± 0.229b	12.663 ± 0.665ab	25.935 ± 0.938cd	3.641 ± 0.678c	52.829 ± 1.023c	9.74
Check 3	0.636 ± 0.015d	3.833 ± 0.167b	11.706 ± 0.615bcd	22.800 ± 0.589e	4.637 ± 0.589b	56.388 ± 1.234b	18.158
AGG53597WWIG1	1.041 ± 0.342ab	3.871 ± 0.432b	13.167 ± 1.457a	28.852 ± 0.765b	4.684 ± 0.752b	48.384 ± 1.398d	9.527
AGG51603WWIG1	0.981 ± 0.035abc	6.196 ± 1.432a	11.722 ± 0.765bc	36.689 ± 0.681a	6.691 ± 0.532a	37.721 ± 1.281e	8.480
Unknown_Vigna_racemosa	0.770 ± 0.123cd	3.856 ± 0.456b	10.201 ± 0.657ef	23.048 ± 0.356e	4.666 ± 0.831b	57.459 ± 1.532ab	2.644
AGG52867WWIG1	0.818 ± 0.281bcd	3.310 ± 0.532b	10.547 ± 0.557de	24.492 ± 0.345de	4.005 ± 0.05bc	56.827 ± 1.982ab	9.051
Unknown Vigna	0.711 ± 0.0432d	3.438 ± 0.346b	9.331 ± 0.327f	23.052 ± 0.312e	4.160 ± 0.281bc	59.307 ± 1.881a	2.353
F statistics	14.946	48.604	28.543	147.317	44.083	180.733	
Pr > F(Model)	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	
Pr > F	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	
Significant	Yes	Yes	Yes	Yes	Yes	Yes	

Results are represented as the mean value of triplicates ± standard error. Mean values without any letter in common within each column are significantly different ($p = 0.05$). **Check 1:** Landrace of *Vigna vexillata*; **Check 2:** Domesticated variety; **Check 3:** Domesticated variety; **Check 4:** Rice Bean (*Vigna umbellata*).

of Check 1 and Check 2 whereas 'Unknown_Vigna_racemosa', AGG52867WVIG1 and 'Unknown Vigna' showed comparable lipid content to that of Check 3. By analogy to the *V. reticulata* species, all the accessions showed comparable ash content to that of the three checks indicating that none of the accessions had higher ash content than the checks. Accession AGG53597WVIG1 showed higher moisture content as compared with Check 1, Check 3 and all the other wild accessions. Moreover, it was comparable to that of Check 2. The accession AGG51603WVIG1 (36.689%) showed substantially significant higher protein content than that of all the checks whereas accession AGG53597WVIG1 (28.852%) had a protein content comparable to that of Check 1. The rest of the accessions had significant low protein content which is comparable to that of Check 3. Furthermore, the greater number of wild accessions presents significantly higher fibre and carbohydrates contents as compared to the checks.

Proximate composition of *Vigna reticulata* accessions

The proximate composition of *V. reticulata* accessions has been summarized in Table 3 (eight accessions) to ease the readability of this article. The complete results presenting the proximate composition of the 36 studied accessions are displayed on Table S1. The results show that the lipid content of most of the wild accessions were not significantly different from those of Check 1 and Check 2. Four accessions (TVNu1394_VRe, TVNu324_VRe, TVNu57_VRe and TVNu141_VRe) exhibited comparable lipid content to that of the check 3. All the accessions showed comparable ash content to that of the three checks indicating that none of the accessions had higher ash content than that of the checks. All the accessions showed lower moisture content than the three checks. The accession TVNu1112_VRe (31.074%) had substantially higher protein content which is significantly higher than that of all the checks. On the other hand, five accessions (TVNu350_VRe, TVNu1852_VRe, TVNu324_VRe, TVNu57_VRe, and TVNu141_VRe) had protein content comparable to that of Check 1 and Check 2. The rest of the accessions had low protein content which is lower than that of Check 3. It was noticed that the greater number of wild accessions present a significantly higher fibre and carbohydrates contents respectively as compared to the checks.

Proximate composition of *Vigna vexillata* accessions

The proximate composition of *V. vexillata* accessions has been summarized in Table 4 (eight accessions) to ease the readability of this article. The complete results presenting the proximate composition of the 35 studied accessions are displayed on Table S2. It was found that the lipid content of the majority of wild accessions is significantly lower than that of Check 1 and check 2, except in the case of accessions AGG308096WVIG2, TVNu333, TVNu293 and TVNu 832 where the lipid content is statistically higher than that of the Check 3. Similar to the *Vigna reticulata* species, all the accessions showed comparable ash content to that of the three checks. A significant number of accessions showed comparable moisture content to that of the checks indicating phenotypic similarity in moisture content. The accessions TVNu832, TVNu1701, TVNu1546, AGG308101WVIG2 and AGG308099WVIG2 exhibited significant higher protein content than that of all the checks.

On the other hand, ten accessions (AGG308097WVIG2, TVNu1378, TVNu1529, TVNu1344, TVNu333, TVNu293,

TVNu178, TVNu781, TVNu120 and TVNu1629) had protein content comparable to that of Check 1 and Check 2. The rest of the accessions had low protein content which is statistically lower than that of Check 3. It is similarly noticed that the greater number of wild accessions present a significantly higher fiber and carbohydrates contents as compared to the checks.

Concomitant analysis of the four *Vigna* species

To examine the relationship that could exist between the proximate composition and the accessions, as well as the relationship between the accessions themselves, a PCA (XLSTAT) was performed using the means values for nutrient component in each accession. Confidence ellipses and correlation circle, combined with an observation chart (Biplot), were obtained as shown in Fig. 1. These were obtained from the PCA analysis of the mean values for each species taken separately. The analysis showed that the first ($F1 = 67.22\%$) and second ($F2 = 17.99\%$) PCA dimensions represent 85.21% of the initial information, which is the best combination that explains the variation among the accessions and traits (proximate composition). It was found that there is a positive correlation between the traits ash, moisture and protein, except for the lipid, fibre and carbohydrate traits, which is due to the angles between their vectors (Fig. 1 and Table S3). It was also noted that all the checks, together with a set of wild accessions, are found on the left side of the F1 axis, forming a group of accessions with lower values for the examined nutrients traits, except for the lipid, fibre and carbohydrates. Those accessions could share common features with the checks. A second group, made up of only wild accessions, was found on the right side of the F1 axis, representing the accessions with higher values for the evaluated traits (Fig. 1).

Discussion

By assessing the proximate composition of 87 accessions from four wild unexplored *Vigna* species, this study revealed that some individual wild accessions have higher nutrient content as compared with domesticated ones which could be advantageous for bio-fortification or domestication. Likewise, the study examined the differences that may exist between the checks. Therefore, it was found that Check 1 and Check 2 might be related in terms of the phenotypic traits lipid, fibre and carbohydrates content though they are of different species (*V. vexillata* and *V. unguiculata*). In addition, it should be noted that Check 1 is landrace of *V. vexillata* which has not yet been fully domesticated as it is noticed that taxonomic arrangements within the *Vigna* genus are not completed (Gore *et al.*, 2019). Phylogenetic proximity between *V. vexillata* and *V. unguiculata* has also been reported (Boukar *et al.*, 2013). However, the differences observed between the three checks or between Checks 1 and 2 with Check 3 for the nutrients evaluated can simply be attributed to their species differences.

According to Table 1, the lipid content of all the wild accessions of *V. ambacensis* is significantly similar to those of check 1 and 2 while it is significantly ($P < 0.05$) higher than that of check 3. This is in line with reports that support the idea of constituents' reduction in legumes due to domestication (Fernández-Marín *et al.*, 2014). Other differences among the checks and the wild accessions may be due to species differences and phylogenetic relationships.

In the case of *V. racemosa* accessions (Table 2), the same trend of result for the proximate composition was observed as in the cases of *V. reticulata* and *V. vexillata*. Following Table 3 and

Table 3. Proximate composition of some studied *Vigna reticulata* accessions (g/100 g)

Accessions	Lipid	Ash	Moisture	Protein	Fiber	Carbohydrates	100-Seed weight(g)
Check 1	1.127 ± 0.128abcd	3.729 ± 0.126abcde	11.052 ± 0.281bcde	27.313 ± 0.597b	3.654 ± 0.832j	53.125 ± 1.323klmn	7.038
Check 2	1.215 ± 0.150abcd	3.716 ± 0.229abcde	12.663 ± 0.665a	25.935 ± 0.938bc	3.641 ± 0.678 j	52.829 ± 1.023lmn	9.74
Check 3	0.636 ± 0.015f	3.833 ± 0.167abc	11.706 ± 0.615 ab	22.800 ± 1.046fghijk	4.637 ± 0.589ghij	56.388 ± 1.234efghijk	18.158
TVNu350_VRe	1.132 ± 0.212abcd	4.130 ± 0.313a	10.610 ± 0.600bcdefg	24.910 ± 0.800cde	7.433 ± 0.760a	51.785 ± 1.005no	8.044
TVNu56_VRe	1.419 ± 0.223a	3.509 ± 0.313abc def	10.175 ± 0.602cdefghijk	23.182 ± 0.800fghi	6.317 ± 0.505abcdefg	55.397 ± 1.034ghijklm	6.844
TVNu1522_VRe	1.108 ± 0.215abcd	3.749 ± 0.310abcde	11.585 ± 0.623abc	23.115 ± 0.802fghij	6.749 ± 0.525abcd	53.694 ± 1.006jklmn	6.888
TVNu1698_VRe	1.391 ± 0.210ab	3.513 ± 0.311abcdef	10.060 ± 0.600defghijkl	22.405 ± 0.800ghijkl	6.324 ± 0.500 abcdefg	56.307 ± 1.055fghijkl	5.488
TVNu1808_VRe	1.015 ± 0.200 cde	3.935 ± 0.310ab	9.910 ± 0.600defghijklm	23.823 ± 0.801efg	7.083 ± 0.515ab	54.234 ± 1.321ijklmn	5.088
TVNu607_VRe	1.374 ± 0.214ab	3.562 ± 0.300abcdef	9.008 ± 0.600klmn	23.525 ± 0.802efgh	6.411 ± 0.508 abcdefg	56.119 ± 1.520fghijkl	8.244
TVNu379_VRe	1.123 ± 0.200 abcd	3.513 ± 0.315abcdef	10.263 ± 0.643cdefghij	23.632 ± 0.800efgh	6.324 ± 0.507 abcdefg	55.145 ± 1.045ghijklmn	5.088
TVNu1852_VRe	1.170 ± 0.223abcd	3.375 ± 0.313abcdefg	10.927 ± 0.614bcdef	26.597 ± 0.868b	6.076 ± 0.500 abcdefh	51.856 ± 1.000mno	4.688
F	12.978	6.535	16.452	56.192	8.277	20.420	
Pr > F(Model)	< 0.0001	< 0.0001	< 0.0001	< 0.0001	< 0.0001	< 0.0001	
Pr > F	< 0.0001	< 0.0001	< 0.0001	< 0.0001	< 0.0001	< 0.0001	
Significant	Yes	Yes	Yes	Yes	Yes	Yes	Yes

Results are represented as the mean value of triplicates ± standard error. Mean values without any letter in common within each column are significantly different ($p = 0.05$). **Check 1:** Landrace of *Vigna vexillata*; **Check 2:** Domesticated variety; **Check 3:** *Vigna unguiculata*; **Check 3:** Domesticated variety; **Check 3:** *Vigna umbellata*.

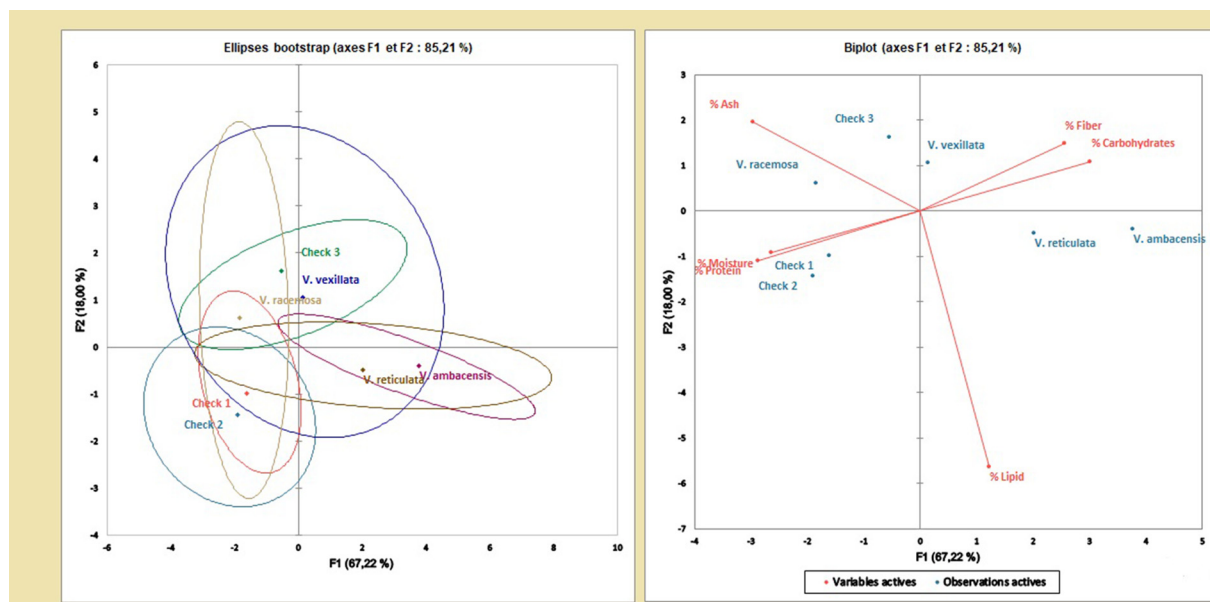


Figure 1. The PCA analysis showing correlations between nutrients contents and wild *Vigna* accessions.

Table S1, the lipid content of most of the wild accessions of *V. reticulata* is not significantly different from that of Check 1 and 2 except for few accessions (TVNu1394_VRe, TVNu324_VRe, TVNu57_VRe and TVNu141_VRe) which are comparable to Check 3. All the accessions show comparable ash content to that of the three checks indicating that none of the accessions had higher ash content than that of the checks. This can be due to species or phylogenetic proximity of *V. reticulata* with Check 1 and 2 (Table 3). All the accessions showed lower moisture content than that of the three checks. The low moisture content observed in wild accessions can be related to the seed characteristics and probably the genetic makeup of the *V. reticulata* accessions as it was earlier reported seed characteristics of wild legumes affect their composition and cooking characteristics (Ereifej, 2004; Altuntas and Demirtola, 2007; Harouna *et al.*, 2019a). The low moisture content could also be a factor of good storing quality of the seeds. The accession with highest protein content (TVNu1112_VRe, 31.074%) might be a suitable genetic material for domestication or breeding and therefore should be further investigated through molecular marker as it high protein content might be due to its genomic difference. This might have been acquired based on the environmental background. For the accessions with protein content comparable to that of Check 1 and Check 2 (TVNu1852_VRe, TVNu141_VRe, TVNu57_VRe, TVNu324_VRe, and TVNu350_VRe), phylogenetic studies as well as breeding and improvement is recommended. The rest of the accessions with very low protein content which is lower than that of Check 3 should be exploited for other nutritional elements. The greater number of wild accessions presented a significantly higher fibre and carbohydrates contents as compared to the checks. This is in line with earlier reports on wild legumes (Macorni *et al.*, 1997; Difo *et al.*, 2015). It might be due to the biosynthesis of many polysaccharides by the wild legumes in order to protect the embryo and survive in harsh environments (Smykal *et al.*, 2014). Therefore, it should be recommended to carry out sound examination of the carbohydrates and fibre fractions to ascertain the digestibility and clear their nutritive contribution in these seeds.

In Table 4 and Table S2, the proximate composition of *V. vexillata* accessions is displayed. The low lipid content found in this result could be attributed to species and genomic differences as explained in the case of *V. reticulata*. Similar to the *V. reticulata* species, all the accessions showed comparable ash content to that of the three checks which can be explained by the same reasons as elaborated earlier. A significant number of accessions showed comparable moisture content to that of the checks indicating phenotypic similarity in moisture content.

The accessions TVNu1701, and AGG30801WVIG1 with highest protein content are speculated to be suitable candidate genetic materials for domestication or breeding. Therefore further investigation should be done through molecular marker as its high protein content might be due to its genomic difference. Furthermore, this might have been acquired based on the environmental background. For the accessions with protein content comparable to that of Check 1 and Check 2, phylogenetic studies as well as breeding and improvement are recommended. As was also noticed some wild accessions present a significantly higher fibre and carbohydrates contents as compared with the checks. This finding concurred with earlier reports on wild legumes (Macorni *et al.*, 1997; Difo *et al.*, 2015). It might be due to the biosynthesis of many polysaccharides by the wild legumes in order to protect the embryo and survive in harsh environments (Smykal *et al.*, 2014) as explained earlier. Therefore, it is recommended to carry-out sound examination of the carbohydrates and fibre fraction to ascertain the digestibility and clear nutritive contribution of the carbohydrates and fibre contained in these seeds.

It is necessary to note that domestication process could also affect other nutritional and health characteristics of the domesticated product as alerted by some researchers (Smykal *et al.*, 2018). The PCA provided indications relating to the domestication of these wild legumes by grouping them based on their quantitative proximate composition traits (Fig. 1).

It was shown that most of the nutrients analysed are positively correlated, and there is a degree of commonality between the checks and a group of some wild species. The check 3 showed close relationship with the *V. vexillata* and *V. racemosa* species,

while the checks 1 and 2 showed close relationship with *V. reticulata* and *V. ambacensis* species as illustrated on the ellipses bootstrap of Fig. 1. This might be due to the relatively slight genetic similarities on their proximate composition which is presented on the biplot of Fig. 1.

Conclusion

The wild *Vigna* species studied presents a considerably high diversity in terms of proximate composition. Despite their under-exploitation for human benefits, the studied wild *Vigna* legumes demonstrated nutrient characteristics comparable with the domesticated ones. The study also demonstrated that the wild *Vigna* species possesses a large variation range of nutrient characteristics such as protein, ash and carbohydrate contents which could be exploited in the improvement of domesticated species or guide the domestication of new species. It was also found that some individual wild accessions have higher nutrients (protein, lipid, ash and carbohydrates) as compared with domesticated ones which could be advantageous for bio-fortification or domestication. The accessions that presented promising traits based on the protein content were: TVNu832, TVNu1701, AGG51603WVIG 1, AGG53597WVIG 1 and TVNu1112. Therefore, these accessions are the best suited for domestication. However, the accessions that showed poor nutrients contents should not be neglected and should be subjected to more study to unveil their potentials. Further investigations including studies involving not only their seeds content, but the other parts of the plant such as their leaves, roots and stem could be of great importance on unveiling their full potential in various aspects of life and science such as crop improvement and domestication, human and animal nutrition, biodiversity conservation and food security.

Supplementary material. The supplementary material for this article can be found at <https://doi.org/10.1017/S1479262124000029>.

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