

Moderation of antidepressant response by the serotonin transporter gene

Patricia Huezo-Diaz,* Rudolf Uher,* Rebecca Smith, Marcella Rietschel, Neven Henigsberg, Andrej Marušič, Ole Mors, Wolfgang Maier, Joanna Hauser, Daniel Souery, Anna Placentino, Astrid Zobel, Erik Roj Larsen, Piotr M. Czerski, Bhanu Gupta, Farzana Hoda, Nader Perroud, Anne Farmer, Ian Craig, Katherine J. Aitchison and Peter McGuffin

Background

There have been conflicting reports on whether the length polymorphism in the promoter of the serotonin transporter gene (5-HTTLPR) moderates the antidepressant effects of selective serotonin reuptake inhibitors (SSRIs). We hypothesised that the pharmacogenetic effect of 5-HTTLPR is modulated by gender, age and other variants in the serotonin transporter gene.

Aims

To test the hypothesis that the 5-HTTLPR differently influences response to escitalopram (an SSRI) compared with nortriptyline (a noradrenaline reuptake inhibitor).

Method

The 5-HTTLPR and 13 additional markers across the serotonin transporter gene were genotyped in 795 adults with moderate-to-severe depression treated with escitalopram or nortriptyline in the Genome Based Therapeutic Drugs for Depression (GENDEP) project.

Results

The 5-HTTLPR moderated the response to escitalopram, with long-allele carriers improving more than short-allele homozygotes. A significant three-way interaction between 5-HTTLPR, drug and gender indicated that the effect was concentrated in males treated with escitalopram. The single-nucleotide polymorphism rs2020933 also influenced outcome.

Conclusions

The effect of 5-HTTLPR on antidepressant response is SSRI specific conditional on gender and modulated by another polymorphism at the 5' end of the serotonin transporter gene.

Declaration of interest

N.H. participated in clinical trials sponsored by pharmaceutical companies including GlaxoSmithKline and Lundbeck. A.F, P.M. and K.J.A have received consultancy fees and honoraria for participating in expert panels from pharmaceutical companies, including Lundbeck and GlaxoSmithKline.

The selective serotonin reuptake inhibitor (SSRI) antidepressants are used as first-line treatment of depression, but up to 50% of people with depression do not adequately benefit from SSRI treatment. 1,2 A significant proportion of the individual differences in response to SSRI is familial.³ When searching for a genetic predictor of SSRI response, the serotonin transporter gene (SLC6A4) is the natural candidate as it encodes the molecular target of SSRI antidepressants. Several functional polymorphisms have been found in the SLC6A4 gene. The common length polymorphism constituted by a deletion of 44 base pairs in the promoter region, referred to as 5-HTTLPR, has been explored in most detail and implicated in depression aetiology.4 The long allele has been associated with more efficient transcription, resilience to depressogenic effects of adversity⁴ and better response to SSRI antidepressants.⁶ A meta-analysis of 14 studies with 1435 participants showed that long-allele carriers had higher probability of response and remission than short-allele homozygotes.⁷ Therefore, SLC6A4 was the prime candidate for response to the SSRI escitalopram and the first gene to be genotyped and tested in the Genome Based Therapeutic Drugs for Depression (GENDEP) project. However, significant heterogeneity of 5-HTTLPR effect was reported in non-European populations.^{8,7} Whereas no effect of 5-HTTLPR on efficacy of citalopram was found in a large mixed-ethnicity sample,9 an ethnicity-sensitive re-analysis of the same sample found an effect in the expected direction among White non-Hispanic participants. 10 As the

5-HTTLPR allele frequencies and its linkage disequilibrium with other polymorphisms within the *SLC6A4* gene vary markedly across populations, ¹¹ it is possible that the discrepancies are a result of the influence of another polymorphism that is in population-specific linkage disequilibrium with the

Three polymorphisms have been proposed to modulate the functionality of 5-HTTLPR: single nucleotide polymorphisms rs25531 and rs2020933, and a 17bp variable tandem repeat in the second intron (STin2). The rs25531 is located within the repeats constituting the 5-HTTLPR. Two initial reports disagreed about the location of this putatively functional A/G single nucleotide polymorphism being either within 12 or immediately upstream¹³ of the 5-HTTLPR insertion/deletion segment. However, sequence alignment has demonstrated that these two reports point to the same polymorphism, the rs25531 (A/G), which is located in the sixth repeat of the 5-HTTLPR outside the deleted segment and therefore can co-occur with either long or short variant of the 5-HTTLPR. 14 The minor G allele of the rs23551 creates a consensus binding sequence for the transcription factor AP-2, which is involved in antidepressant action. 15,16 It has been proposed that the rs25531 influences the functionality of the serotonin transporter and recoding of 5-HTTLPR long alleles based on the rs25531 has been suggested.¹⁷ The functionality of this single nucleotide polymorphism has however not been consistently replicated and it is still unclear whether the recoding of the 5-HTTLPR is warranted. 4,14,18 Therefore, in the present study, the rs25531 is evaluated separately from the 5-HTTLPR. The rs2020933 single nucleotide polymorphism is located in the

^{*} These authors contributed equally to the work.

first intron, approximately 2.5kb 3′ to the 5-HTTLPR and contributes to the variation of serotonin transporter expression. ¹⁸ The STin2 two major alleles (STin2.10 and STin2.12) correspond to 10 or 12 repeats of a 17bp sequence and the STin2.12 allele has been associated with enhanced transcription. ¹⁹

It has been proposed that the moderation of antidepressant response by the 5-HTTLPR may depend on age.²⁰ This suggestion is supported by data showing age-related variation in serotonergic function,^{21–23} but tests of interaction between age and 5-HTTLPR have not been reported in most pharmacogenetic studies.

Gender may be another important factor influencing the relationship between 5-HTTLPR genotype and antidepressant response. Ovarian steroids have a strong influence on serotonin synthesis, ²⁴ expression of serotonergic receptors, ^{25–27} and the serotonin transporter. ^{28,29} There are gender differences in 5-HTTLPR moderation of response to serotonergic challenges, including tryptophan depletion ³⁰ and to stressful life events. ^{4,31} However, previous pharmacogenetic studies have not reported a test of gender × genotype interaction.

The GENDEP project aims to identify differential predictors of response to SSRI and tricyclic antidepressants. The primary aim is to test the hypothesis that the 5-HTTLPR interacts with the drug used (escitalopram ν . nortriptyline) in treatment of depression. Specifically, it is expected that short-allele homozygotic status will be associated with poor response to escitalopram but will not influence response to nortriptyline. As the heterogeneity of findings across studies may be a result of the influence of a polymorphism that is in population-specific linkage disequilibrium with the 5-HTTLPR, we systematically explore DNA sequence variation across the *SLC6A4* gene, including the three polymorphisms with known effects on gene expression. Additionally, the moderating effects of age and gender are tested.

Method

Study design and sample

The GENDEP project is an open-label part-randomised multicentre pharmacogenetic study with two active pharmacological treatment arms.³² It was designed to establish clinical and genetic determinants of therapeutic response to two antidepressants with contrasting primary modes of action: nortriptyline (a tricyclic inhibitor of monoamine reuptake with strongest affinity for the noradrenaline transporter) and escitalopram (an SSRI). Eight hundred and eleven adults with ICD–10³³/DSM–IV³⁴ unipolar major depression of at least moderate severity³⁵ were recruited

in eight European countries: Belgium, Croatia, Denmark, Germany, Italy, Poland, Slovenia and the UK. To minimise confounding by population stratification, recruitment was restricted to individuals of White European parentage. Personal or family history of bipolar disorder or schizophrenia constituted exclusion criteria. The study was approved by ethics boards in all participating centres. All participants provided a written consent after the procedures were fully explained.

Participants included 296 men and 514 women between 19 and 72 years old (Table 1). The average participant was in her second episode of moderately severe depression and scored 28.7 (s.d.=6.7) on the Montgomery–Åsberg Depression Rating Scale (MADRS),³⁶ 21.7 (s.d.=5.3) on the 17-item Hamilton Rating Scale for Depression (HRSD–17)^{37,38} and 28.0 (s.d.=9.7) on the Beck Depression Inventory (BDI)³⁹ at baseline.

Participants with no contraindications were randomly allocated to receive flexible dosage of nortriptyline (50–150 mg daily) or escitalopram (10–30 mg daily) for 12 weeks. Individuals with relative or absolute contraindications (medical conditions incompatible with the use of the antidepressant and history of intolerance or non-response to one study medication) for one of the drugs were allocated non-randomly to the other antidepressant. Other psychotropic medication was not allowed with the exception of occasional use of hypnotics. Adherence was monitored by weekly self-report and plasma levels of antidepressants were measured at week 8. Adverse effects of medication were measured using the UKU Side-Effect Rating Scale⁴⁰ and Self-Report Antidepressant Side-Effect Checklist.⁴¹

Outcome measurement

The response to antidepressant medication is a complex phenotype that includes changes in a number of symptoms occurring over a period of up to 12 weeks following the initiation of an antidepressant. In GENDEP, the responses to escitalopram and nortriptyline were assessed by weekly administration of three established measures of depression severity: the clinician-rated 10-item MADRS, the HRSD–17 and the self-report 21-item BDI were administered at week 0 (baseline) and then weekly for 12 weeks. All raters were trained and achieved high interrater reliability, with intraclass correlation of at least 0.9.⁴² A psychometric analysis found that the MADRS was the most internally consistent and informative of the three scales and that depressive symptoms could be described in more detail by three symptom dimensions derived by categorical item factor analysis: observed mood,

	Es	citalopram (<i>n</i> = 45	0)	Nortriptyline (n = 345)			
	1/1	l/s	s/s	1/1	l/s	s/s	
Total, n (%)	162 (36)	211 (47)	77 (17)	135 (38)	157 (46)	53 (16)	
Female gender n (%)	107 (66)	127 (60)	44 (57)	86 (64)	103 (66)	36 (69)	
Age, years: mean (s.d.)	41.4 (11.5)	42.9 (11.4)	44.5 (12.5)	41.5 (11.4)	42.8 (11.5)	41.3 (13.8)	
Education, years: mean (s.d.)	12.2 (3.2)	12.2 (3.3)	11.8 (2.9)	12.4 (3.1)	11.9 (3.1)	12.1 (3.1)	
Married/cohabiting, n (%)	84 (52)	103 (49)	38 (49)	75 (56)	78 (50)	25 (47)	
Employed/student, n (%)	95 (59)	120 (57)	40 (52)	78 (58)	71 (45)	27 (51)	
Age at depression onset, years: mean (s.d.)	32.3 (9.5)	33.5 (9.8)	34.3 (9.7)	31.7 (9.8)	32.3 (10)	31.4 (10.4)	
First episode, n (%)	61 (38)	76 (36)	23 (30)	41 (30)	42 (27)	17 (32)	
Current episode duration, weeks: mean (s.d.)	18.2 (12.3)	19.8 (12.1)	19.7 (10.0)	17.1 (11.0)	20.5 (14.6)	17.7 (13.7)	
History of taking antidepressants, n (%)	62 (38)	96 (45)	30 (39)	68 (50)	84 (54)	22 (42)	
Baseline severity (MADRS), mean (s.d.)	28.5 (6.0)	28.2 (6.7)	28.4 (7.6)	28.7 (6.7)	29.3 (6.9)	29.9 (6.5)	

cognitive symptoms and neurovegetative symptoms.⁴² In the pharmacogenetic analyses we used the MADRS score as a primary continuous outcome variable. Findings of interest were further explored using the three symptom dimensions.

Genotyping

Adequate DNA samples were available for 795 of the 811 participants. A sample of 5–8 ml of blood was collected in ethylenediamine tetra-acetic acid and frozen. DNA was extracted using a standard procedure.⁴³

A two-stage method was used to type the 5HTTLPR and rs25531. In the first stage, short and long alleles were determined by polymerase chain reaction (PCR). In the second stage, the A/G alleles of rs25531 single nucleotide polymorphism within the 5HTTLPR repeats were identified using restriction fragment length polymorphism. The PCR product was digested with the Msp1 enzyme, which cuts the amplified PCR product at two or three places, depending on the rs25531 genotype. The combinations of products that determine the genotypes are shown in Appendix 1.

Five of the microsatellite markers listed on the UCSC bioinformatics website (http://genome.ucsc.edu/ accessed in September 2004) were selected for investigation and genotyped on 20 individuals, using PCR and electrophoresis, to check whether common polymorphisms at these loci are present. Only STin2 (*SLC6A4*, intron2) and STin4 (*SLC6A4*, intron4) proved to be polymorphic, and thus were carried forward for full analysis on the GENDEP sample. Fluorescent forward primers were used for amplification of these microsatellites. Genotyping of the PCR products was performed using the ABI3130 sequencer.

Single nucleotide polymorphisms were selected that tag DNA sequence variation within the SLC6A4 gene and its 1000 bp margins using the SNPTagger program (www.broad.mit.edu/mpg/tagger) run in Windows XP and HAPMAP data on the CEPH CEU population with European ancestry (CEPH NCBI Build 35/UCSC hg17/May 2004 coordinates). ⁴⁴ Criteria for single nucleotide polymorphism selection included a minimal allele frequency of 5% in the White population and a pairwise $r^2 = 0.8$. Ten single nucleotide polymorphisms met these criteria (Table 2). According to SNPTagger, these 10 single nucleotide

polymorphisms provided 92% coverage of the DNA sequence variation in the *SLC6A4* gene. The single nucleotide polymorphisms were genotyped using the SNPlexTM method. This included oligonucleotide ligation assay/PCR technology for allelic discrimination and ligation product amplification. The assay products were run on the ABI3130 sequencer and data were exported and analysed using GENEMAPPER software version 3.7 for Windows XP (Applied Biosystems). Eleven samples were genotyped in blind duplicates and the agreement was 100% for all markers.

In addition to the *SLC6A4* markers, a panel of 35 single nucleotide polymorphism markers selected as being informative for population stratification in a European population ⁴⁵ were genotyped in all individuals to establish the presence of and allow effective control for any population stratification.

The linkage disequilibrium measures (*D'* and *R*²) were calculated using the Haploview program version 4 for Windows XP. ⁴⁶ Markers were included irrespective of genotype distribution as deviations from the Hardy–Weinberg equilibrium are expected in a cases-only sample. ⁴⁷ However, Hardy–Weinberg equilibrium was tested and results are given in Table 2.

Statistical analysis

To use all available weekly data on response to antidepressants and provide unbiased estimates in the presence of missing values, effects of genotype markers were tested using linear mixed models with individual random intercept and slope and fitted with maximum likelihood. All models included the fixed effects of time (linear and quadratic), baseline depression severity, history of using antidepressants, drug, age and gender, and random effects of the individual and the recruitment centre. To account for correlations between repeated observations, the intercept and slope of time were allowed to vary randomly between participants. The results of regression are presented as standardised regression coefficients (β) with 95% confidence intervals. Standardised regression coefficients denote the number of standard deviations on the outcome measure per unit of the predictor variable. As the standard deviation of the MADRS was 7, a B of 0.14 corresponds to a difference of one point on the MADRS.

As there was strong prior evidence for the influence of 5-HTTLPR on response to serotonergic antidepressants, the

Table 2	Linkage d	isequilibriu	ım, allele	frequency	and Ha	ardy–Weinb	erg equili	brium ^a					
	HTTLPR	rs2020933 r	rs2066713	rs2020939	STin2	rs8076005 r	rs2020942	STin4	rs140700	rs4583306	rs140701	rs4325622	rs3813034
HTTLPR		0.04	0.09	0.05	0.07	0.00	0.09	0.07	0.01	0.07	0.08	0.05	0.05
rs2020933	0.90		0.01	0.03	0.00	0.17	0.01	0.04	0.04	0.03	0.04	0.04	0.04
rs2066713	0.47	0.39		0.42	0.89	0.14	0.97	0.47	0.06	0.46	0.47	0.31	0.31
rs2020939	0.25	0.72	0.94		0.38	0.14	0.42	0.88	0.06	0.86	0.86	0.71	0.71
STin2	0.40	0.27	0.99	0.84		0.11	0.92	0.43	0.06	0.43	0.43	0.27	0.27
rs8076005	0.07	0.74	1.00	0.90	0.86		0.14	0.16	0.42	0.15	0.15	0.17	0.17
rs2020942	0.46	0.38	0.99	0.93	1.00	1.00		0.46	0.06	0.46	0.47	0.31	0.31
STin4	0.29	0.81	0.99	0.94	0.91	0.95	0.99		0.08	0.97	0.98	0.81	0.81
rs140700	0.40	0.24	1.00	0.89	0.89	0.98	1.00	1.00		0.08	0.07	0.09	0.09
rs4583306	0.29	0.81	0.99	0.93	0.90	0.95	0.99	0.99	1.00		0.97	0.79	0.79
rs140701	0.29	0.81	0.99	0.93	0.90	0.95	0.99	0.99	0.96	1.00		0.80	0.80
rs4325622	0.24	0.79	0.76	0.89	0.67	0.96	0.75	0.95	1.00	0.95	0.94		1.00
rs3813034	0.24	0.79	0.76	0.89	0.67	0.96	0.75	0.95	1.00	0.95	0.94	1.00	
MAF	0.40	0.06	0.39	0.43	0.42	0.18	0.39	0.43	0.09	0.43	0.43	0.46	0.46
HWE χ^2	0.87	0.05	2.00	1.75	12.33	0.13	2.40	1.46	1.22	0.53	1.56	4.82	5.18
HWE P	0.3500	0.8292	0.1574	0.1853	0.0004	0.7201	0.1210	0.2264	0.2695	0.4659	0.2121	0.0282	0.0228

a. Linkage disequilibrium between genotyped markers is indexed by D' (below the diagonal) and R^2 (above the diagonal) estimated in Haploview. The lower part of the table shows minor allele frequency (MAF), the chi-squared statistics of the test of Hardy–Weinberg equilibrium (HWE χ^2) and the associated P (HWE P).

5-HTTLPR was first tested as a single marker without correcting for multiple testing and the model was further developed using a stepwise approach. The effect of genotype and its interactions with drug, gender and age were tested as fixed explanatory variables in the mixed linear models. Competing nested models were compared using likelihood ratio tests. The previously reported recessive short allele model^{7,6} was compared with a full genotype model. To test the hypothesis that a polymorphism in linkage disequilibrium with the 5-HTLPR modulates its effect, other markers within the SLC6A4 gene were then added in order of linkage disequilibrium with the 5-HTTLPR and retained if model fit improved significantly according to likelihood ratio tests (P < 0.05).

Haplotypes for markers within the SLC6A4 gene that were in linkage disequilibrium (D'>0.7) were estimated 12 times using a Bayesian procedure in Phase, version 2.1 for Windows XP^{48,49} and used as multiply imputed data-sets in the mixed linear models.⁵⁰ This method has been demonstrated to provide unbiased estimates and to have an adequate confidence interval coverage even when haplotypes are estimated based on cases only.⁵⁰

The ancestry-informative markers were analysed using 10 000 burn-ins and 10000 repetitions under an admixture model with correlated allele frequencies in STRUCTURE version 2.2 for Windows XP (http://pritch.bsd.uchicago.edu/software.html).⁵¹ This analysis found no clustering within the sample. Stratification was further estimated using the genomic control approach⁵² implemented in PLINK for Windows XP (http://pngu.mgh. harvard.edu/ \sim purcell/plink). Based on the same set of ancestry-informative single nucleotide polymorphisms and all candidate gene markers, PLINK estimated a genomic inflation factor of 1, confirming absence of significant stratification in the GENDEP sample.

Power analysis

The sample size required for detecting an interaction between antidepressant and the 5-HTTLPR genotype was estimated based on data from published European studies that provided continuous outcome measures by genotype group. 6,54 These studies showed that 5-HTTLPR influenced outcome with an effect size of 0.65-1.00 s.d. on a continuous outcome scale. Assuming a recessive model with minor allele frequency of 0.4, an effect size of 0.65 in participants treated with escitalopram and no effect of genotype on outcome in participants treated with nortriptyline, 270 participants per treatment arm were needed to detect a

drug × genotype interaction at $\alpha = 0.05$ with a power of 0.8. The GENDEP sample provided a power of more than 0.9.

Results

Genotyping

The 5-HTTLPR, intron 2 and intron 4 tandem repeats and 11 single nucleotide polymorphisms were typed in 795 (98%) of the 811 participants. Sample demographic and clinical characteristics at baseline were unrelated to the 5-HTTLPR genotype (all P > 0.05, Table 1). There was also no significant relationship between the 5-HTTLPR genotype and the mode of allocation (random v. non-random, $\chi^2 = 1.34$, P = 0.246) or the drug allocated among non-randomised participants ($\chi^2 = 0.0001$, P = 0.994). The 16 participants with missing or inadequate DNA samples did not differ from the genotyped participants on depression severity, age or gender (all P > 0.05). Table 2 shows the linkage disequilibrium among the 14 genotyped markers and minor allele frequencies. Frequencies of the 5-HTTLPR, s25531, rs2020933 and STin2 genotypes are given in Table 3.

The 5-HTTLPR

The 5-HTTLPR genotype moderated response to the serotonergic antidepressant in the expected direction. A significant three-way interaction between the 5-HTTLPR, drug and gender (P = 0.026; Table 4) indicated that this moderating effect is conditional on the type of antidepressant drug and on gender. Specifically, male carriers of the 5-HTTLPR long allele had a better response to escitalopram than male short-allele homozygotes (Fig. 1). The best-fitting model included a recessive effect of the 5-HTTLPR short allele, its two-way interactions with gender and drug, as well as the three-way interaction. Age did not significantly interact with genotype.

The interaction effects were qualified by significant main effects in participants treated with escitalopram and especially men treated with escitalopram. Among participants treated with escitalopram those homozygous for the short allele had significantly worse outcome than long-allele carriers ($\beta = 0.154$, 95% CI 0.003–0.305, P = 0.045). The effect of the 5-HTTLPR genotype was strong among males treated with escitalopram ($\beta = 0.333$, 95% CI 0.085–0.580, P = 0.008, Fig. 1a) but absent among females treated with escitalopram ($\beta = 0.036$, 95% CI -0.155 to 0.228, P = 0.709, Fig. 1b). The 5-HTTLPR genotype did not affect outcome among the participants treated with nortriptyline

	Escit	alopram, <i>n</i> (%) (<i>n</i> =	450)	Nortriptyline, <i>n</i> (%) (<i>n</i> = 345)			
	1/1	l/s	s/s	1/1	l/s	s/s	
s25531 genotype							
AA	126 (78)	177 (84)	75 (97)	112 (83)	139 (89)	52 (98)	
AG	33 (20)	31 (15)	2 (3)	21(16)	17 (11)	1 (2)	
GG	2 (1)	0 (0)	0 (0)	1 (1)	0 (0)	0 (0)	
s2020933 genotype							
TT	129 (80)	185 (88)	75 (97)	107 (79)	140 (89)	51 (96)	
TA	29 (18)	21 (10)	2 (3)	26 (19)	15 (10)	0 (0)	
AA	1 (1)	0 (0)	0 (0)	1 (1)	1 (1)	0 (0)	
Tin2 genotype							
12–12	36 (22)	76 (36)	40 (52)	36 (27)	59 (38)	34 (64)	
12–10	77 (48)	88 (42)	26 (34)	60 (44)	60 (38)	13 (25)	
10–10	43 (27)	33 (16)	8 (10)	34 (25)	26 (17)	4 (8)	
9–10 /9–12 /9–9	5 (3)	11 (5)	2 (3)	5 (3)	9 (6)	0 (0)	
long allele; s, short allele.							

Table 4 Results of the final best-fitting models for overall depression symptoms (Montgomery-Åsberg Depression Rating Scale (MADRS)) ^a							
MADRS	β	s.e.	Р	95% CI			
5-HTTLPR (recessive s)	0.194	0.076	0.010	0.046 to 0.343			
Drug	0.065	0.018	< 0.001	0.029 to 0.100			
Gender	0.320	0.117	0.006	0.091 to 0.549			
Interaction 5-HTTLPR × gender	-0.272	0.150	0.070	-0.567 to 0.022			
Interaction 5-HTTLPR × drug	-0.537	0.194	0.006	−0.917 to −0.157			
Interaction gender × drug	-0.219	0.095	0.021	−0.405 to −0.034			
Interaction 5-HTTLPR × drug × gender	0.532	0.240	0.027	0.061 to 1.004			
rs2020933 (additive)	0.120	0.061	0.049	0.001 to 0.240			

a. Time, baseline severity, history of taking antidepressants, age and constant were included in all models but coefficients are not shown for these terms. Beta (B) is the standardised regression coefficient and can be interpreted as effect size. Note that as a result of the inclusion of interaction terms in the model, the first-order effects have to be interpreted as conditional on the interacting variables: thus the effects of drug (advantage of escitalopram) refers only to male carriers of long alleles at the 5-HTTLPR; the effect of gender (advantage of being male) refers to 5-HTTLPR long-allele carriers treated with escitalopram. Unconditional effects of drug and gender are reported elsewhere.³²

($\beta=-0.063$, 95% CI -0.231 to 0.106, P=0.466, Fig. 1c and 1d). The 5-HTTLPR genotype explained 1.1% of variance in the repeated measurement of depression severity among men treated with escitalopram, but only 0.3% of variance in antidepressant response across the whole sample.

The additive or full genotype model provided no advantage over the recessive short-allele model (all likelihood ratio tests had P > 0.1). The 5-HTTLPR genotype was unrelated to baseline severity, antidepressant dosage, self-reported adherence, plasma levels of either nortriptyline or escitalopram and adverse effects.

rs25531

The single nucleotide polymorphism rs25531 (A/G; minor allele frequency 0.075) was in linkage disequilibrium with the 5-HTTLPR (D' = 0.82, $R^2 = 0.18$).

Given the controversies about the moderation of 5-HTTLPR functionality by the rs25531,^{17,18} we tested whether the recoding of the 5-HTTLPR based on the rs25531 presumed functionality,¹⁷ or its inclusion as a separate marker improved the model fit. This was not the case: the recoding impaired model fit and its inclusion

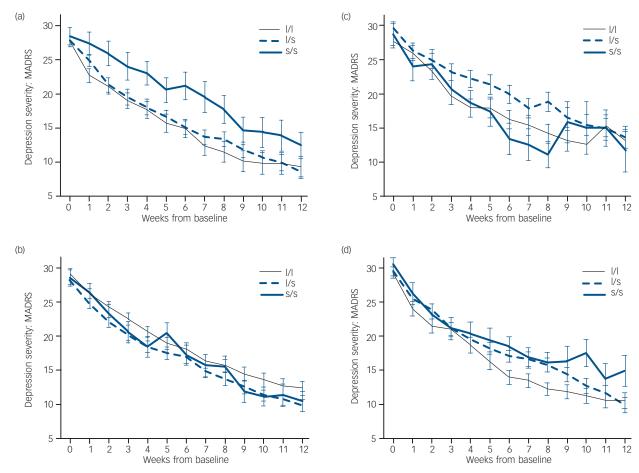


Fig. 1 Moderation of antidepressant response by 5-HTTLPR genotype and gender: (a) men treated with escitalopram, (b) women treated with escitalopram, (c) men treated with nortriptyline, (d) women treated with nortriptyline. Depression severity measured by the Montgomery-Asberg Depression Rating Scale (MADRS) in observed cases over the 12 weeks of treatment. Details of genotype are given in key; I, long allele; s, short allele. Note that values for weeks 9–12 are less reliable because of missing values resulting from drop-out and switching.

as a separate marker added no information to the prediction of antidepressant response (likelihood ratio test P > 0.1). Therefore, the rs25531 was dropped from further analyses. The rs25531-recoded 5-HTTLPR genotype was also not related to baseline severity, antidepressant dosage, self-reported adherence, plasma levels of either nortriptyline or escitalopram and adverse effects.

rs2020933

The single nucleotide polymorphism rs2020933 (A/T; minor allele frequency 0.06) was in linkage disequilibrium with the 5-HTTLPR ($D'=0.90,\ R^2=0.04$). The addition of rs2020933 (additive or dominant coding) to the model as a fixed first-order effect provided a significant improvement in the prediction of antidepressant response. Interaction between rs2020933 and 5-HTTLPR, drug, gender and age did not significantly improve the model. The final best-fitting model included both 5-HTTLPR with its interacting effects and the rs2020933 and led to a significant improvement in the prediction of antidepressant response compared with the model with no genotype data (likelihood ratio test $\chi^2(5)=13.57,\ P=0.0219,\ Table 4$).

The intron 2 simple tandem repeat

The STin2 was in weak linkage disequilibrium with the 5-HTTLPR (D' = 0.40, $R^2 = 0.07$). The STin2 did not contribute to the prediction of treatment response (likelihood ratio test P > 0.1).

Haplotype analysis

As there was significant linkage disequilibrium between the 5-HTTLPR and the rs2020933, the influence of estimated haplotypes comprising these two markers on antidepressant response was investigated. Although the haplotypes containing the 5-HTTLPR long allele were associated with better response to escitalopram, the inclusion of haplotypes provided no advantage over single marker analysis in predicting drug response.

Other markers within the SLC6A4 gene

Other markers within the *SLC6A4* gene had low linkage disequilibrium with 5-HTTLPR and their inclusion as single markers or haplotypes conferred no improvement in the prediction model.

Effects on symptom dimensions

Our previous analyses have indicated that separable dimensions of depressive symptoms may differ in their response to anti-depressants. We therefore investigated the genotype effect on change on the three symptoms dimensions. The 5-HTTLPR and rs2020933 significantly moderated response on the observed mood and cognitive symptom dimensions. The change on the neurovegetative symptom dimension was relatively independent of genotype (Table 5). The effect on the observed mood dimension was stronger than on the MADRS.

Discussion

The GENDEP data support the previously reported moderation of antidepressant response by the length polymorphism in the promoter region of the serotonin transporter gene. This effect is specific to the serotonergic mode of antidepressant action, appears to be concentrated in men and may be moderated by another polymorphism at the 5′ end of the serotonin transporter gene.

Gender effects

Within the GENDEP sample, the effect of the 5-HTTLPR on response to escitalopram was marked in men but absent in women. Among men treated with escitalopram, the 5-HTTLPR polymorphism is associated with a five-point difference on the MADRS between short-allele homozygotes and long-allele

	β	s.e.	Р	95% CI
	β	5.E.	F	93% CI
bserved mood				
5-HTTLPR (recessive s)	0.231	0.075	0.002	0.083 to 0.378
Drug	0.056	0.018	0.002	0.020 to 0.091
Gender	0.344	0.116	0.003	0.117 to 0.572
Interaction 5-HTTLPR × gender	-0.329	0.149	0.027	-0.621 to -0.03
Interaction 5-HTTLPR × drug	-0.504	0.192	0.009	-0.881 to -0.12
Interaction gender × drug	-0.163	0.094	0.083	-0.347 to 0.021
Interaction 5-HTTLPR × drug × gender	0.554	0.238	0.020	0.086 to 1.022
rs2020933 (additive)	0.134	0.061	0.027	0.015 to 0.253
ognitive				
5-HTTLPR (recessive s)	0.252	0.080	0.002	0.095 to 0.409
Drug	0.051	0.019	0.007	0.014 to 0.089
Gender	0.308	0.123	0.013	0.066 to 0.551
Interaction 5-HTTLPR × gender	-0.233	0.159	0.143	-0.545 to 0.079
Interaction 5-HTTLPR × drug	-0.518	0.204	0.011	-0.919 to -0.11
Interaction gender × drug	-0.234	0.100	0.020	-0.431 to -0.03
Interaction 5-HTTLPR × drug × gender	0.464	0.254	0.068	-0.034 to 0.962
rs2020933 (additive)	0.118	0.064	0.067	-0.009 to 0.244
leurovegetative				
5-HTTLPR (recessive s)	0.076	0.081	0.344	-0.082 to 0.235
Drug	0.079	0.019	< 0.001	0.040 to 0.117
Gender	0.235	0.125	0.060	-0.010 to 0.480
Interaction 5-HTTLPR × gender	-0.202	0.160	0.207	-0.517 to 0.112
Interaction 5-HTTLPR × drug	-0.459	0.207	0.027	−0.865 to −0.053
Interaction gender × drug	-0.270	0.101	0.008	-0.469 to -0.07
Interaction 5-HTTLPR × drug × gender	0.349	0.257	0.174	-0.154 to 0.853
rs2020933 (additive)	0.073	0.065	0.263	-0.055 to 0.200

a. Time, baseline severity, history of taking antidepressants, age and constant were included in all models but coefficients are not shown for these terms as they are of no interest for the present report. Note that as a result of the inclusion of interaction terms in the model, the first-order effects have to be interpreted as conditional on the interacting variables thus the effects of drug (advantage of escitalopram) refers only to male carriers of long alleles at the 5-HTTLPR; the effect of gender (advantage of being male) refers to 5-HTTLPR long-allele carriers treated with escitalopram. Unconditional effects of drug and gender are reported elsewhere.³²

carriers. This finding is consistent with previous reports of genderspecific genetic influences on serotonergic function.⁵⁵ It may reflect a biological interaction between the serotonergic system and ovarian hormones. Through the oestrogen alpha receptor, oestrogens stimulate the production of the 5-HT_{1A} receptor that is involved in the regulation of serotonin release and is downregulated in response to serotonin reuptake inhibitors.²⁷ Oestrogens also increase the expression of the serotonin transporter.²⁹ The impact of experimental manipulations of serotonergic function also depends on gender and hormonal status. Decrease in serotonergic function during tryptophan depletion has different effects on men and women³⁰ and oestrogen supplementation may enhance the action of antidepressants in perimenopause. 57,58 It is possible that the impact of the less functional short 5-HTTLPR allele is moderated by the oestrogen-induced stimulatory effect on serotonin transporter expression in hormonally active women. Although this accumulated evidence adds biological plausibility to the observed 5-HTTLPR × gender interaction, this finding requires replication in an independent sample.

Age effects

Although gender appears to be an important factor, age did not significantly interact with genotype in its effect on outcome. Joyce *et al*²⁰ reported interaction between age and 5-HTTLPR with age dichotomised at 25. As the GENDEP sample is older, separate analysis of individuals below 25 years of age was not feasible. Therefore, the current negative results should not be interpreted as a refutation of the findings by Joyce and colleagues.²⁰

The effect of rs2020933

The response to antidepressants in the GENDEP study was also associated with the single nucleotide polymorphism rs2020933, which is located in the first intron of the SLC6A4 gene. The major T allele was associated with a better response to both nortriptyline and escitalopram and there was no significant interaction with gender. The rs2020933 single nucleotide polymorphism has been reported to be functional in an allelic expression imbalance study.¹⁸ It is in significant linkage disequilibrium with the 5-HTTLPR, but because of a large difference in minor allele frequency, only a relatively small proportion of variance in the 5-HTTLPR is shared with the rs2020933. As this genomic region varies significantly between populations,11 the interplay between these two functional loci has the potential to explain discordant findings in samples of different ethnic origins.^{7,8} This remains to be tested in non-European samples. However, as the effect of rs2020933 was relatively weak and was not specific to the serotonergic mode of action, it is unlikely to fully explain the influence of the 5-HTTLPR on response to serotonergic antidepressants.

Drug effects on symptom dimensions

Depression is a heterogeneous group of disorders and the genetic mediation of response to antidepressants may differ between separable dimensions of depressive symptoms. For example, it has been demonstrated that change in sleep and appetite-related symptoms is relatively independent of change in core depressive symptoms such as mood and anhedonia.⁵⁹ In a psychometric analysis of the GENDEP data, we have identified partially separable dimensions of observed mood, cognitive symptoms and neurovegetative symptoms that differ in their response to the two antidepressants.^{32,38} It has been previously reported that the moderation by the 5-HTTLPR of response to antidepressants

is relatively specific to core mood symptoms and does not affect changes in sleep-related symptoms. ⁶⁰ The present analysis supports this distinction: the 5-HTTLPR and rs2020933 genotypes had the strongest influence on the observed mood dimension, comprising the core depressive symptoms and anxiety. These genotypes also moderated response on the cognitive symptom dimension. However, changes on the neurovegetative symptom dimension which comprises sleep, appetite and libido, were relatively independent of the 5-HTTLPR and rs2020933 genotypes. These findings add to the accumulating evidence indicating that mood and neurovegetative symptoms have distinct pathogenesis.

Effects of genotype on tolerability

It has been reported that tolerability rather than efficacy of antidepressants may be influenced by the 5-HTTLPR genotypes. 61,62 We have therefore explored the relationship between the 5-HTTLPR genotype and adverse effects and related variables. We found that the 5-HTTLPR genotype was not related to escitalopram or nortriptyline dosage, plasma levels, adverse effects, drop-out or self-reported adherence in the GENDEP sample. These results remained negative after recoding according to the rs25531 marker. Thus we can conclude that, in the European population, the 5-HTTLPR genotype is directly related to antidepressant efficacy and there is no significant effect on tolerability.

Strengths and limitations

The GENDEP project is the largest study to date to compare a tricyclic antidepressant with an SSRI and is the second largest sample with pharmacogenetic data on antidepressant treatment outcome. It is however only powered to detect interacting effects of moderate size. The associations between polymorphisms in the SLC6A4 gene and response to antidepressants are of small to moderate size. Subsequently, the reported findings are detected with a modest level of certainty that satisfies the nominal statistical significance threshold but would not survive corrections for the number of polymorphisms that have been reported or suggested to moderate antidepressant response. Therefore, the results of the present study must be interpreted against the background of other published findings. The association of 5-HTTLPR with response to serotonergic antidepressants appears to be a robust and replicable finding in populations of European origin. However, the concentration of effect among males and the additional moderation by the rs2020933 polymorphism are novel findings that have to be tested in other large samples.

In conclusion, the available data suggest that variations in the 5' end of the serotonin transporter gene have an effect on anti-depressant efficacy, which is of modest effect size and depends on type of antidepressant and gender. Future studies should evaluate the population-specificity of this effect and the role of other polymorphisms including the rs2020933.

Funding

The GENDEP study was funded by a European Commission Framework 6 grant, EC Contract Ref.: LSHB-CT-2003-503428. Lundbeck provided both nortriptyline and escitalopram free of charge for the GENDEP study. GlaxoSmithkline contributed by funding an add-on project in the London centre. The sponsors had no role in the design and conduct of the study, in data collection, analysis, interpretation or writing the report.

Acknowledgements

We would like to thank the following collaborators for their contribution: Helen Dean, Joanna Gray, Cerisse Gunasinghe, Amanda Elkin, Desmond Campbell, Richard J. Williamson,

Julien Mendlewicz, Mara Barreto, Thomas Schulze, Christine Schmael, Susanne Höfels, Anna Schuhmacher, Ute Pfeiffer, Sandra Weber, Lisbeth Jorgensen, Anne Schinkel Stamp, Caterina Giovannini, Dejan Kozel, Moica Z. Dernovsek, Alenka Tancic, Jerneja Sveticic, Zrnka Kovacic, Pawe Kapelski, Maria Skibiñska, Aleksandra Rajewska, Monika Dmitrzak-Weglarz, Aleksandra Szczepankiewicz, and Elzbieta Cegielska. We would like to specially acknowledge the contribution of Jorge Perez, who was the principal investigator at Brescia, Italy, and who passed away in October 2007, and the important contribution made by Dr Andrej Marušič, the principal investigator at Ljubljana, Slovenia, who passed away in June 2008.

Patricia Huezo-Diaz, PhD, Rudolf Uher, PhD, MRCPsych, Rebecca Smith, BSc, Institute of Psychiatry, King's College London, Uk; Marcella Rietschel, MD, Central Institute of Mental Health, Division of Genetic Epidemiology in Psychiatry, Mannheim, Germany; Neven Henigsberg, MD, Croatian Institute for Brain Research, Medical School, University of Zagreb, Croatia; Andrej Marušič, PhD, Institute of Public Health, Ljubljana, Slovenia; Ole Mors, PhD, Centre for Psychiatric Research, Aarhus University Hospital, Risskov, Denmark; Wolfgang Maier, MD, Department of Psychiatric Genetics, Poznan University of Medical Sciences, Poland; Daniel Souery, PhD, Université Libre de Bruxelles, Erasme Academic Hospital, Department of Psychiatry, Brussels, Belgium; Anna Placentino, PsyD, Biological Psychiatry Unit and Dual Diagnosis ward IRCCs, Centro San Giovanni di Dio, FBF, Brescia, Italy, Astrid Zobel, MD, Department of Psychiatry, University of Bonn, Germany, Erik Roj Larsen, MD, PhD, Mood Disorders Research Unit, Aarhus University Hospital, Risskov, Denmark; Piotr M. Czerski, Laboratory of Psychiatric Genetics, Poznan University of Medical Sciences, Poland; Bhanu Gupta, MRCPsych, Farzana Hoda, BSc, Nader Perroud, MD, Anne Farmer, MD, FRCPsych, Ian Craig, PhD, Katherine J. Aitchison, PhD, MRCPsych, Peter McGuffin, PhD, FRCP, FRCPsych, Institute of Psychiatry, King's College London, UK.

Correspondence: Rudolf Uher, P080, Social Genetic and Developmental Psychiatry Centre, Institute of Psychiatry, 16 De Crespigny Park, London SE5 8AF UK. Email: rudolf.uher@iop.kcl.ac.uk

First received 4 Dec 2008, final revision 12 Jan 2009, accepted 5 Feb 2009

Appendix

5-HTTLPR and rs25531 genotyping

Restriction fragment length polymorphism analysis of PCR products carrying either the long (L, 16 repeat) or short (S, 14 repeat) alleles of the 5HTTLPR (L/S) and rs25531 (a/g).

Eight combinations were identified within the GENDEP sample: LaLa (324bp, 62bp and 33bp fragment), LaLg (324bp, 174bp, 150bp), LaSa (324bp, 281bp), SaLg (281bp, 174bp, 150bp), SaSa (281bp), LgLg (174bp, 150bp) and SaSg (281bp, 150bp, 131bp).

References

- 1 Thase ME, Entsuah AR, Rudolph RL. Remission rates during treatment with venlafaxine or selective serotonin reuptake inhibitors. Br J Psychiatry 2001; 178: 234-41
- 2 Trivedi MH, Rush AJ, Wisniewski SR, Nierenberg AA, Warden D, Ritz L, et al. Evaluation of outcomes with citalopram for depression using measurement-based care in STAR*D: implications for clinical practice. Am J Psychiatry 2006; 163: 28–40.
- 3 Franchini L, Serretti A, Gasperini M, Smeraldi E. Familial concordance of fluvoxamine response as a tool for differentiating mood disorder pedigrees. J Psychiatr Res 1998; 32: 255–9.
- 4 Uher R, McGuffin P. The moderation by the serotonin transporter gene of environmental adversity in the aetiology of mental illness: review and methodological analysis. *Mol Psychiatry* 2008; 13: 131–46.
- 5 Lesch KP, Balling U, Gross J, Strauss K, Wolozin BL, Murphy DL, et al. Organization of the human serotonin transporter gene. J Neural Transm Gen Sect 1994; 95: 157–62.
- 6 Smeraldi E, Zanardi R, Benedetti F, Di BD, Perez J, Catalano M. Polymorphism within the promoter of the serotonin transporter gene and antidepressant efficacy of fluvoxamine. *Mol Psychiatry* 1998; 3: 508–11.
- 7 Serretti A, Kato M, De RD, Kinoshita T. Meta-analysis of serotonin transporter gene promoter polymorphism (5-HTTLPR) association with selective serotonin reuptake inhibitor efficacy in depressed patients. *Mol Psychiatry* 2007; 12: 247–57.
- 8 Kim H, Lim SW, Kim S, Kim JW, Chang YH, Carroll BJ, et al. Monoamine transporter gene polymorphisms and antidepressant response in Koreans with late-life depression. JAMA 2006; 296: 1609–18.
- 9 Kraft JB, Peters EJ, Slager SL, Jenkins GD, Reinalda MS, McGrath PJ, et al. Analysis of association between the serotonin transporter and antidepressant response in a large clinical sample. *Biol Psychiatry* 2007; 61: 734–42.

- 10 Mrazek DA, Rush AJ, Biernacka JM, O'Kane DJ, Cunningham JM, Wieben ED, et al. SLC6A4 variation and citalopram response. Am J Med Genet B Neuropsychiatr Genet 2009; 150B: 341–51.
- 11 Gelernter J, Cubells JF, Kidd JR, Pakstis AJ, Kidd KK. Population studies of polymorphisms of the serotonin transporter protein gene. Am J Med Genet 1999; 88: 61–6.
- 12 Hu X, Oroszi G, Chun J, Smith TL, Goldman D, Schuckit MA. An expanded evaluation of the relationship of four alleles to the level of response to alcohol and the alcoholism risk. *Alcohol Clin Exp Res* 2005; 29: 8–16.
- 13 Kraft JB, Slager SL, McGrath PJ, Hamilton SP. Sequence analysis of the serotonin transporter and associations with antidepressant response. *Biol Psychiatry* 2005; 58: 374–81.
- 14 Wendland JR, Martin BJ, Kruse MR, Lesch KP, Murphy DL. Simultaneous genotyping of four functional loci of human SLC6A4, with a reappraisal of 5-HTTLPR and rs25531. Mol Psychiatry 2006; 11: 224-6.
- 15 Braganca J, Eloranta JJ, Bamforth SD, Ibbitt JC, Hurst HC, Bhattacharya S. Physical and functional interactions among AP-2 transcription factors, p300/ CREB-binding protein, and CITED2. J Biol Chem 2003; 278: 16021–9.
- 16 Damberg M. Transcription factor AP-2 and monoaminergic functions in the central nervous system. J Neural Transm 2005; 112: 1281–96.
- 17 Hu XZ, Lipsky RH, Zhu G, Akhtar LA, Taubman J, Greenberg BD, et al. Serotonin transporter promoter gain-of-function genotypes are linked to obsessive-compulsive disorder. Am J Hum Genet 2006; 78: 815–26.
- 18 Martin J, Cleak J, Willis-Owen SA, Flint J, Shifman S. Mapping regulatory variants for the serotonin transporter gene based on allelic expression imbalance. Mol Psychiatry 2007; 12: 421–2.
- 19 MacKenzie A, Quinn J. A serotonin transporter gene intron 2 polymorphic region, correlated with affective disorders, has allele-dependent differential enhancer-like properties in the mouse embryo. *Proc Natl Acad Sci USA* 1999; 96: 15251–5
- 20 Joyce PR, Mulder RT, Luty SE, McKenzie JM, Miller AL, Rogers GR, et al. Age-dependent antidepressant pharmacogenomics: polymorphisms of the serotonin transporter and G protein beta3 subunit as predictors of response to fluoxetine and nortriptyline. Int J Neuropsychopharmacol 2003; 6: 339–46.
- 21 Tauscher J, Verhoeff NP, Christensen BK, Hussey D, Meyer JH, Kecojevic A, et al. Serotonin 5-HT1A receptor binding potential declines with age as measured by [11C]WAY-100635 and PET. Neuropsychopharmacology 2001; 24: 522–30.
- 22 van Dyck CH, Malison RT, Seibyl JP, Laruelle M, Klumpp H, Zoghbi SS, et al. Age-related decline in central serotonin transporter availability with [(123)|]beta-CIT SPECT. Neurobiol Aging 2000; 21: 497–501.
- 23 Yamamoto M, Suhara T, Okubo Y, Ichimiya T, Sudo Y, Inoue M, et al. Agerelated decline of serotonin transporters in living human brain of healthy males. *Life Sci* 2002; 71: 751–7.
- 24 Hiroi R, McDevitt RA, Neumaier JF. Estrogen selectively increases tryptophan hydroxylase-2 mRNA expression in distinct subregions of rat midbrain raphe nucleus: association between gene expression and anxiety behavior in the open field. *Biol Psychiatry* 2006; 60: 288–95.
- 25 Biegon A, McEwen BS. Modulation by estradiol of serotonin receptors in brain. *J Neurosci* 1982; 2: 199–205.
- 26 Sumner BE, Fink G. The density of 5-hydoxytryptamine2A receptors in forebrain is increased at pro-oestrus in intact female rats. *Neurosci Lett* 1997: 234: 7–10.
- 27 Wissink S, van der BB, Katzenellenbogen BS, van der Saag PT. Synergistic activation of the serotonin-1A receptor by nuclear factor-kappa B and estrogen. *Mol Endocrinol* 2001; 15: 543–52.
- 28 Lu NZ, Eshleman AJ, Janowsky A, Bethea CL. Ovarian steroid regulation of serotonin reuptake transporter (SERT) binding, distribution, and function in female macaques. *Mol Psychiatry* 2003; 8: 353–60.
- 29 McQueen JK, Wilson H, Fink G. Estradiol-17 beta increases serotonin transporter (SERT) mRNA levels and the density of SERT-binding sites in female rat brain. Brain Res Mol Brain Res 1997; 45: 13–23.
- 30 Walderhaug E, Magnusson A, Neumeister A, Lappalainen J, Lunde H, Refsum H, et al. Interactive effects of sex and 5-HTTLPR on mood and impulsivity during tryptophan depletion in healthy people. *Biol Psychiatry* 2007; 62: 593–99.
- 31 Sjoberg RL, Nilsson KW, Nordquist N, Ohrvik J, Leppert J, Lindstrom L, et al. Development of depression: sex and the interaction between environment and a promoter polymorphism of the serotonin transporter gene. *Int J Neuropsychopharmacol* 2006; 9: 443–9.
- 32 Uher R, Maier W, Hauser J, Marušič A, Schmael C, Mors O, et al. Differential efficacy of escitalopram and nortriptyline on dimensional measures of depression. Br J Psychiatry 2009; 194: 252–9.

- 33 World Health Organization. The ICD-10 Classification of Mental and Behavioural Disorders: Clinical Descriptions and Diagnostic Guidelines. WHO, 1992
- 34 American Psychiatric Association. Diagnostic and Statistical Manual of Mental Disorder (4th edn) (DSM-IV). APA, 1994.
- 35 Wing JK, Sartorius N, Ustin TB. Diagnosis and Clinical Measurement in Psychiatry. A Reference Manual for SCAN. World Health Organization, 1998.
- 36 Montgomery SA, Åsberg M. A new depression scale designed to be sensitive to change. Br J Psychiatry 1979; 134: 382–9.
- 37 Hamilton M. A rating scale for depression. J Neurol Neurosurg Psychiatry 1960; 23: 56-62.
- 38 Hamilton M. Development of a rating scale for primary depressive illness. *Br J Clin Psychol* 1967; **6**: 278–96.
- 39 Beck AT, Ward CH, Mendelson M, Mock J, Erbaugh J. An inventory for measuring depression. Arch Gen Psychiatry 1961; 4: 561–71.
- 40 Lingjaerde O, Ahlfors UG, Bech P, Dencker SJ, Elgen K. The UKU side effect rating scale. A new comprehensive rating scale for psychotropic drugs and a cross-sectional study of side effects in neuroleptic-treated patients. Acta Psychiatr Scand Suppl 1987: 334: 1–100.
- **41** Uher R, Farmer A, Henigsberg N, Rietschel M, Mors O, Maier W, et al. Adverse reactions to antidepressants. *Br J Psychiatry* in press.
- 42 Uher R, Farmer A, Maier W, Rietschel M, Hauser J, Marusic A, et al. Measuring depression: comparison and integration of three scales in the GENDEP study. *Psychol Med* 2008; 38: 289–300.
- 43 Freeman B, Smith N, Curtis C, Huckett L, Mill J, Craig IW. DNA from buccal swabs recruited by mail: evaluation of storage effects on long-term stability and suitability for multiplex polymerase chain reaction genotyping. *Behav Genet* 2003; 33: 67–72.
- 44 De Bakker PI, Yelensky R, Pe'er I, Gabriel SB, Daly MJ, Altshuler D. Efficiency and power in genetic association studies. *Nat Genet* 2005; 37: 1217–23.
- 45 Seldin MF, Shigeta R, Villoslada P, Selmi C, Tuomilehto J, Silva G, et al. European population substructure: clustering of northern and southern populations. PLoS Genet 2006; 2: e143.
- 46 Barrett JC, Fry B, Maller J, Daly MJ. Haploview: analysis and visualization of LD and haplotype maps. *Bioinformatics* 2005; 21: 263–5.
- 47 Wittke-Thompson JK, Pluzhnikov A, Cox NJ. Rational inferences about departures from Hardy-Weinberg equilibrium. Am J Hum Genet 2005; 76: 047, 84
- 48 Stephens M, Donnelly P. A comparison of Bayesian methods for haplotype reconstruction from population genotype data. Am J Hum Genet 2003; 73: 1162–9.

- 49 Stephens M, Smith NJ, Donnelly P. A new statistical method for haplotype reconstruction from population data. Am J Hum Genet 2001; 68: 978–89.
- 50 Cordell HJ. Estimation and testing of genotype and haplotype effects in casecontrol studies: comparison of weighted regression and multiple imputation procedures. *Genet Epidemiol* 2006; 30: 259–75.
- 51 Falush D, Stephens M, Pritchard JK. Inference of population structure using multilocus genotype data: linked loci and correlated allele frequencies. *Genetics* 2003; 164: 1567–87.
- 52 Devlin B, Roeder K. Genomic control for association studies. *Biometrics* 1999; 55: 997–1004
- 53 Purcell S, Neale B, Todd-Brown K, Thomas L, Ferreira MA, Bender D, et al. PLINK: a tool set for whole-genome association and population-based linkage analyses. Am J Hum Genet 2007: 81: 559–75.
- 54 Zanardi R, Serretti A, Rossini D, Franchini L, Cusin C, Lattuada E, et al. Factors affecting fluvoxamine antidepressant activity: influence of pindolol and 5-HTTLPR in delusional and nondelusional depression. *Biol Psychiatry* 2001: 50: 323–30.
- 55 Weiss LA, Abney M, Cook EH, Jr, Ober C. Sex-specific genetic architecture of whole blood serotonin levels. Am J Hum Genet 2005; 76: 33–41.
- 56 Morgan ML, Cook IA, Rapkin AJ, Leuchter AF. Estrogen augmentation of antidepressants in perimenopausal depression: a pilot study. J Clin Psychiatry 2005: 66: 774–80
- 57 Soares CN, Almeida OP, Joffe H, Cohen LS. Efficacy of estradiol for the treatment of depressive disorders in perimenopausal women: a double-blind, randomized, placebo-controlled trial. Arch Gen Psychiatry 2001; 58: 529–34.
- 58 Katz MM, Koslow SH, Frazer A. Onset of antidepressant activity: reexamining the structure of depression and multiple actions of drugs. *Depress Anxiety* 1996; 4: 257–67.
- 59 Serretti A, Mandelli L, Lorenzi C, Pirovano A, Olgiati P, Colombo C, et al. Serotonin transporter gene influences the time course of improvement of "core" depressive and somatic anxiety symptoms during treatment with SSRIs for recurrent mood disorders. Psychiatry Res 2007; 149: 185–93.
- 60 Hu XZ, Rush AJ, Charney D, Wilson AF, Sorant AJ, Papanicolaou GJ, et al. Association between a functional serotonin transporter promoter polymorphism and citalopram treatment in adult outpatients with major depression. Arch Gen Psychiatry 2007; 64: 783–92.
- 61 Murphy GM Jr, Hollander SB, Rodrigues HE, Kremer C, Schatzberg AF. Effects of the serotonin transporter gene promoter polymorphism on mirtazapine and paroxetine efficacy and adverse events in geriatric major depression. *Arch Gen Psychiatry* 2004; 61: 1163–9.