

TABLE 1. Bacterial and Fungal Air Contamination Values in the Monitored Environments before (First 5 Samplings) and after (Sixth Sampling) the Corrective Action

Sampling points	N	Active sampling						Passive sampling					
		Bacteria			Fungi			Bacteria			Fungi		
		Mean	SD	Median	Mean	SD	Median	Mean	SD	Median	Mean	SD	Median
Before corrective action													
Rooms (ambient air)	15	32.5	60.3	16	20.4	27.7	14	52	48	63	398	161	440
Rooms (HVAC)	15	19.3	16.7	16	12	18	8	...	...	...	...	...	...
Corridor (ambient air)	5	135	103.2	148	2.8	1.1	2	818	1,046	472	0	0	0
After corrective action													
Rooms (ambient air)	3	2.7	3.1	2	0	0	0	0	0	0	0	0	0
Rooms (HVAC)	3	1.3	2.3	0	0	0	0	...	...	...	...	...	...
Corridor (ambient air)	1	10 <sup>a</sup>	...	...	0 <sup>a</sup>	...	...	0 <sup>a</sup>	...	...	0 <sup>a</sup>	...	...

NOTE. Active samplings are expressed as colony-forming units per cubic meter; passive samplings are expressed as the index of microbial air contamination, in colony-forming units per square meter per hour. HVAC, heating, ventilation, and air-conditioning.

<sup>a</sup> This figure refers to 1 sample.

planned and should be carried out by skilled personnel using adequate methods. Results should be properly analyzed and effectively communicated, and, most importantly, action should be taken in case of anomalies. We also advocate a closer cooperation between infection control teams and hospital engineering departments.

#### ACKNOWLEDGMENTS

*Potential conflicts of interest.* All authors report no conflicts of interest relevant to this article. All authors submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest, and the conflicts that the editors consider relevant to this article are disclosed here.

Cesira Pasquarella, MD, PhD;<sup>1</sup> Pietro Vitali, MD;<sup>2</sup>  
Elisa Saccani, MD;<sup>1</sup> Francesco Mariotti, MD;<sup>1</sup>  
Carmine Boccuni, MD;<sup>1</sup> Pietro Manotti, MD;<sup>1</sup>  
Manuela Ugolotti, BS;<sup>3</sup> Roberto Albertini, PhD<sup>3</sup>

Affiliations: 1. Department of Public Health, University of Parma, Parma, Italy; 2. Department of Hygiene and Preventive Medicine, University Hospital of Parma, Parma, Italy; 3. Laboratory of Aerobiology and Environmental Quality Control, Departments of Clinical Medicine, Nephrology, and Health Sciences, University of Parma, Parma, Italy.

Address correspondence to Cesira Pasquarella, MD, PhD, Department of Public Health, University of Parma, Via Volturno 39, 43125 Parma, Italy (ira.pasquarella@unipr.it).

*Infect Control Hosp Epidemiol* 2012;33(1):101-102

© 2012 by The Society for Healthcare Epidemiology of America. All rights reserved. 0899-823X/2012/3301-0021\$15.00. DOI: 10.1086/663646

#### REFERENCES

- National Advisory Committee on Immunization. Construction-related nosocomial infections in patient in health care facilities. *Can Commun Dis Rep* 2001;27(suppl 2).
- Vonberg RP, Gastmeier P. Nosocomial aspergillosis in outbreak settings. *J Hosp Infect* 2006;63:246-254.
- Centers for Disease Control and Prevention (CDC). *Guidelines for environmental infection control in health-care facilities, 2003.*

<http://www.cdc.gov/mmwr/preview/mmwrhtml/rr5210a1.htm>. Published June 2003. Accessed November 11, 2011.

- Tomblyn M, Chiller T, Einsele H, et al. Guidelines for preventing infectious complications among hematopoietic cell transplantation recipients: a global perspective. *Biol Blood Marrow Transplant* 2009;15:1143-1238.
- International Standardization Organization. *ISO 14698-1:2003: Clean-rooms and associated controlled environments—biocontamination control: general principles and methods*. Geneva: International Standardization Organization, 2003.
- Pasquarella C, Pitzurra O, Savino A. The index of microbial contamination. *J Hosp Infect* 2000;46:241-256.
- Eickhoff TC. Airborne nosocomial infection: a contemporary perspective. *Infect Control Hosp Epidemiol* 1994;15:663-672.
- European Commission. *EU guidelines to good manufacturing practice. Medicinal products for human and veterinary use. Annex 1. Manufacture of sterile medicinal products (corrected version)*. Brussels: European Commission, 2008.
- Landrin A, Bissery A, Kac G. Monitoring air sampling in operating theatre: can particle counting replace microbiological sampling? *J Hosp Infect* 2005;61:27-29.

## The Value of Universal versus Targeted Screening for Methicillin-Resistant *Staphylococcus aureus* among Admission Patients

*To the Editor*—We read with interest the evaluation by Leonhardt et al<sup>1</sup> of universal versus targeted screening for methicillin-resistant *Staphylococcus aureus* (MRSA) on admission to the hospital, in particular the finding that there was no impact on MRSA transmission rates. Leonhardt and colleagues used polymerase chain reaction (PCR) for detection in 2 hospitals, and their admission prevalence rates were less than 5%—that is, 1.76% and 3.24%—during the control

phase. As part of a larger study of the epidemiology of MRSA in Ireland, we have evaluated the additional yield from screening all patients on admission to 4 acute hospital wards compared with screening only patients at risk over a period of 3 years but by means of culture, not PCR.<sup>2</sup> Overall, 5% of patients were positive, but this declined from 9% in year 1 to 2% in year 3. MRSA was recovered from 4 (1%) of 340 of patients who did not have risk factors normally associated with MRSA—for example, admission to the hospital during the previous 18 months and previous MRSA colonization or infection.<sup>2</sup>

In recent years, the UK government has mandated the screening of all patients admitted to the hospital, starting with elective admissions, but this is associated with many logistical challenges.<sup>3</sup> These include additional pressure on diagnostic laboratories, the requirement for isolation when additional MRSA cases are identified, and whether additional patient screening should be undertaken in preference to other measures, such as screening for MRSA colonization among members of staff.<sup>3</sup> Interim results from universal screening in Scotland have indicated that 7.5% of patients were colonized on admission to the hospital and that 88% of patients were available for screening.<sup>4</sup> Screening was carried out using culture techniques rather than PCR, and the interval between the collection of samples and the identification of an MRSA-positive patient was often greater than the inpatient stay; hence, screening was of less value for instituting isolation and contact precautions and decolonization programs. Consequently, the potential benefits of universal screening as initially anticipated may not be realized because of a failure to achieve 100% uptake and delays in acting on positive results in the absence of molecular methods for screening.

While there remains a strong case for active screening of patients at particular risk on admission to an acute hospital—for example, previously known MRSA patients and transfers from other hospitals—the justification for screening all patients admitted to an acute hospital remains unconvincing. In addition to the low additional yield relative to that obtained from screening only at-risk patients,<sup>2</sup> universal screening results in additional expense that seems hard to justify.<sup>1,2</sup> Nonetheless, individual healthcare institutions need to assess how extensive screening should be, on the basis of the local prevalence over a reasonable period of time (ie, 1 year or more) and the likely impact that additional screening may have. In addition, before undertaking universal screening it is essential to confirm that all patients at risk are already being screened, as 27% of at-risk patients were not being screened before the start of an assessment of the use of PCR to screen for MRSA.<sup>5</sup> However, when universal screening is to be used there is a strong case for using molecular methods, which reduce the turnaround time and thus facilitate early

isolation of MRSA-positive patients or the release from pre-emptive isolation of suspected MRSA-positive patients after a negative result.<sup>5,6</sup>

In conclusion, we agree with the conclusions of Leonhardt et al<sup>1</sup> that while universal screening may increase the rate of detection, the additional expense probably does not justify its widespread implementation in most institutions.

## ACKNOWLEDGMENTS

*Financial support.* The work referred to was funded through a translational research award (TRA/004/06) from the Health Research Board (Ireland).

*Potential conflicts of interest.* H.H. reports recent research collaborations with Steris, 3M, Inov8 Science, Pfizer, and Cepheid. He also reports recently receiving lecture and other fees from 3M, Novartis, and Astellas. E.C. reports no conflicts of interest. All authors submitted the ICMJE Form for Disclosure of Potential Conflicts of Interest, and the conflicts that the editors consider relevant to this article are disclosed here.

**E. Creamer, MSc;<sup>1</sup> H. Humphreys, MD<sup>1,2</sup>**

Affiliations: 1. Department of Clinical Microbiology, Royal College of Surgeons in Ireland, Beaumont Hospital, Dublin, Ireland; 2. Department of Microbiology, Beaumont Hospital, Dublin, Ireland.

Address correspondence to H. Humphreys, MD, Department of Clinical Microbiology, RCSI Education and Research Centre, Beaumont Hospital, PO Box 9063, Dublin 9, Ireland (h Humphreys@rcsi.ie).

*Infect Control Hosp Epidemiol* 2012;33(1):102-103  
© 2012 by The Society for Healthcare Epidemiology of America. All rights reserved. 0899-823X/2012/3301-0022\$15.00. DOI: 10.1086/663647

## REFERENCES

1. Leonhardt KK, Yakusheva O, Phelan D, et al. Clinical effectiveness and cost benefit of universal versus targeted methicillin-resistant *Staphylococcus aureus* screening upon admission in hospitals. *Infect Control Hosp Epidemiol* 2011;32:797–803.
2. Creamer E, Galvin S, Dolan A, et al. Evaluation of screening risk and nonrisk patients for methicillin-resistant *Staphylococcus aureus* on admission in an acute care hospital. *Am J Infect Control* doi:10.1016/j.ajic.2011.07.008. Published September 30, 2011.
3. Dancer SJ. Considering the introduction of universal MRSA screening. *J Hosp Infect* 2008;69:315–320.
4. Reilly JS, Stewart S, Christie P, et al. Universal screening for methicillin-resistant *Staphylococcus aureus*: interim results from the NHS Scotland pathfinder project. *J Hosp Infect* 2010;74:35–41.
5. Creamer E, Dolan A, Sherlock O, et al. The effect of rapid screening for methicillin-resistant *Staphylococcus aureus* (MRSA) on the identification and early isolation of MRSA-positive patients. *Infect Control Hosp Epidemiol* 2010;31:374–381.
6. Uckay I, Sax H, Iten A, et al. Effect of screening for methicillin-resistant *Staphylococcus aureus* carriage by polymerase chain reaction on the duration of unnecessary preemptive contact isolation. *Infect Control Hosp Epidemiol* 2008;29:1077–1079.