

Research Article

Cite this article: Al-Mekhlafi HM *et al.* (2019). Prevalence and risk factors of *Strongyloides stercoralis* infection among Orang Asli schoolchildren: new insights into the epidemiology, transmission and diagnosis of strongyloidiasis in Malaysia. *Parasitology* **146**, 1602–1614. <https://doi.org/10.1017/S0031182019000945>

Received: 9 May 2019

Revised: 1 July 2019

Accepted: 1 July 2019

First published online: 31 July 2019


Key words:

Malaysia; neglected tropical diseases; Orang Asli; schoolchildren; soil-transmitted helminth; *Strongyloides stercoralis*

Author for correspondence:

Hesham M. Al-Mekhlafi, E-mail: halmekhlafi@yahoo.com and Yvonne A. L. Lim, E-mail: limailian@um.edu.my

Prevalence and risk factors of *Strongyloides stercoralis* infection among Orang Asli schoolchildren: new insights into the epidemiology, transmission and diagnosis of strongyloidiasis in Malaysia

Hesham M. Al-Mekhlafi^{1,2} , Nabil A. Nasr³, Yvonne A. L. Lim³, Fatin Nur Elyana³, Hany Sady⁴, Wahib M. Atroosh^{3,5}, Salwa Dawaki⁶, Tengku Shahrul Anuar^{7,8}, Rahmah Noordin⁹ and Rohela Mahmud³

¹Medical Research Centre, Jazan University, 45142 Jazan, Kingdom of Saudi Arabia; ²Department of Parasitology, Faculty of Medicine and Health Sciences, Sana'a University, 1247 Sana'a, Yemen; ³Department of Parasitology, Faculty of Medicine, University of Malaya, 50603 Kuala Lumpur, Malaysia; ⁴Department of Medical Laboratories, Faculty of Medical Sciences, Hodeidah University, Hodeidah, Yemen; ⁵Department of Microbiology and Parasitology, Faculty of Medicine and Health Sciences, University of Aden, Aden, Yemen; ⁶School of Health Technology, Club Road, Nassarawa, Kano, 700282 Kano State, Nigeria; ⁷Centre of Medical Laboratory Technology, Faculty of Health Sciences, Universiti Teknologi MARA, Puncak Alam Campus, 42300 Selangor, Malaysia; ⁸Integrative Pharmacogenomics Institute, Universiti Teknologi MARA, Puncak Alam Campus, 42300 Selangor, Malaysia and ⁹Institute for Research in Molecular Medicine, Universiti Sains Malaysia, 11800 Pulau Pinang, Malaysia

Abstract

This cross-sectional study aimed to determine the prevalence and risk factors of *S. stercoralis* infection among 1142 Orang Asli primary schoolchildren in six different states of Peninsular Malaysia. Fecal samples were examined using direct smear, formalin-ether sedimentation (FES), agar plate culture (APC) and PCR techniques. Overall, 15.8% of the children were found to be infected with *S. stercoralis*. The prevalence was 0.2, 1.3, 15.2 and 13.7% by direct smear, FES, APC and PCR, respectively. Multivariate analysis showed that an age of >10 years, being male, belonging to a Proto-Malay tribe, belonging to the Senoi tribe, indiscriminate defecation, using an unimproved water source for drinking water and not wearing shoes when outside were the significant risk factors of infection among these children. In conclusion, we provide new evidence on the occurrence of *S. stercoralis* in Malaysia to show that there is a relatively high prevalence of infection among Orang Asli schoolchildren. Therefore, the use of specific methods for detecting *S. stercoralis* should be considered when screening these children for intestinal parasites. Moreover, prevention and control measures specific to *S. stercoralis* should be integrated into the intestinal parasitic infections control programme in Malaysia.

Introduction

Strongyloidiasis, which is caused by *Strongyloides stercoralis*, is one of the most difficult-to-diagnose soil-transmitted helminth (STH) infections. It occurs worldwide and is endemic in tropical and temperate climates (Schär *et al.*, 2013; Jourdan *et al.*, 2018). It is considered the most neglected of the neglected tropical diseases and its prevalence is largely underestimated (Olsen *et al.*, 2009; Viney, 2017). Moreover, several aspects of the epidemiology of *S. stercoralis* infection remain poorly documented (Bisoffi *et al.*, 2013; Nutman, 2017). Based on available information; it is estimated that about 370 million people are infected with *S. stercoralis* worldwide, with the prevalence rate ranging from 10 to 70% in tropical and subtropical countries where conducive conditions for transmission such as moist soil and inadequate sanitation coexist. It is particularly evident in underprivileged communities in Latin America, West Africa and Southeast Asia (Schär *et al.*, 2013; Bisoffi *et al.*, 2014; Puthiyakunnon *et al.*, 2014).

Humans acquire *S. stercoralis* infection generally through skin penetration of filariform larvae that exist in contaminated soil. Moreover, donor-derived strongyloidiasis infection in solid organ transplant recipients is increasingly being reported (Kim *et al.*, 2016; Winnicki *et al.*, 2018). Strongyloidiasis is commonly chronic and long lasting with a majority of infected individuals remaining asymptomatic, and infections could be sustained in individuals for more than 75 years (Prendki *et al.*, 2011; Junior *et al.*, 2017). Pulmonary migration of larvae results in transient eosinophilia with cough, dyspnoea (shortness of breath), wheezing and haemoptysis; known as Loeffler's syndrome which is seen in heavy infections (Al Hadidi *et al.*, 2018). Chronic *S. stercoralis* infection may present with fatigue, anorexia, vomiting, abdominal pain, diarrhoea and urticaria (Khieu *et al.*, 2013; Nutman, 2017). In immunocompetent individuals, infection produces negligible symptoms, but in immunocompromised patients the autoinfection cycle of this parasite

can be life-threatening because it can lead to *Strongyloides* hyperinfection syndrome that has a mortality rate of nearly 100% if left untreated and typically presents as intestinal or pulmonary failure (Kassalik and Mönkemüller, 2011; Nutman, 2017). In immunocompromised patients, disseminated strongyloidiasis can occur, with large numbers of parasites spreading to affect various organs, including the liver, kidneys, brain, cutaneous and subcutaneous tissue and heart, with occasional gut translocation of bacteria causing bacteraemia (Bisoffi *et al.*, 2013; Shimasaki *et al.*, 2015).

Strongyloides stercoralis diagnosis remains a challenge because individuals carrying the infection often remain asymptomatic and there is no gold-standard reference method for its diagnosis (Requena-Méndez *et al.*, 2013; Nutman, 2017). The most common diagnostic techniques that are used to detect most intestinal parasites are the direct fecal smear, formalin-ether sedimentation (FES) and Kato-Katz, but these methods often failed to detect *S. stercoralis* larvae or they have low sensitivity, particularly in the case of chronic infections (Requena-Méndez *et al.*, 2013; Buonfrate *et al.*, 2017). Hence, more-sensitive and *Strongyloides*-specific methods such as the Baermann method, Koga agar plate culture (APC) and molecular assays are now being used to detect this parasite and to update the epidemiology of strongyloidiasis worldwide (Amor *et al.*, 2016). In reality, however, these methods are not routinely used in parasitological examinations because they are time consuming and require more resources than the most commonly applied methods, particularly in the potentially endemic settings of resource-poor countries (Agrawal *et al.*, 2009; Schär *et al.*, 2013). Although serological methods are very useful in the diagnosis of strongyloidiasis and have been validated for clinical and epidemiological use, their specificity is variable and they show reduced sensitivity in the case of low-intensity infections (Bisoffi *et al.*, 2014; Buonfrate *et al.*, 2017; Fradejas *et al.*, 2018).

In Southeast Asia, high prevalence rates of *S. stercoralis* infection have been reported in Lao PDR, Cambodia and Thailand (Puthiyakunnon *et al.*, 2014; Schär *et al.*, 2016; Senephansiri *et al.*, 2017; Forrer *et al.*, 2018). In Malaysia, several previous studies are available on the prevalence of intestinal parasitic infections (IPIs) throughout the country; however, data on *S. stercoralis* infection are still very limited. The first study on *S. stercoralis* in Malaysia reported in a single case among Orang Asli in the Kelantan state; however, this study applied low-sensitivity diagnostic methods (Rahmah *et al.*, 1997). In addition, despite the high seropositive rates reported among indigenous populations in Selangor state, West Malaysia (31.5%; 17/54) and Sarawak state, East Malaysia (11%; 26/236), parasitological methods failed to detect *S. stercoralis* larvae in fecal samples among the studied participants (Ahmad *et al.*, 2013; Ngui *et al.*, 2016).

In light of the above status, the present study used multiple diagnostic methods to investigate the prevalence and risk factors of *S. stercoralis* infection among Orang Asli schoolchildren in Peninsular Malaysia. This study is crucial because it will not only provide reliable community-based evidence on the presence of *S. stercoralis* in Orang Asli communities to enable a better understanding of the epidemiology of human strongyloidiasis in Malaysia, it will also update the available information on the global distribution of strongyloidiasis.

Materials and methods

Study design and study area

A cross-sectional, community-based study was carried out among Orang Asli (aboriginal) schoolchildren in six states of Peninsular Malaysia: (Selangor, Pahang, Negeri Sembilan, Kelantan, Johor and Perak) from January to April 2017.

The Orang Asli communities that were involved in this study were residents in 11 districts as follows: Hulu Selangor (3°35'N latitude, 101°35'E longitude) and Petaling (3°05'N latitude, 101°35'E longitude), which are districts in Selangor state; Raub (3°47'N latitude, 101°51'E longitude) and Lipis (4°15'N latitude, 101°50'E longitude) in Pahang state; Jelebu (3°0'N latitude, 102°05'E longitude) and Kuala Pilah (2°44'N latitude, 102°14'E longitude) in Negeri Sembilan state; Gua Musang (4°53'N latitude, 101°58'E longitude) in Kelantan state; Segamat (2°30'N latitude, 102°55'E longitude) in Johor state; and Hulu Perak (5°20'N latitude, 101°15'E longitude), Muallim (3°50'N latitude, 101°30'E longitude) and Batang Padang (4°05'N latitude, 101°20'E longitude) in Perak state (Fig. 1). These districts were randomly selected from a list describing the distribution of Orang Asli communities in Peninsular Malaysia that was provided by the Department of Orang Asli Development (also known as JAKOA). In these Orang Asli communities, primary schools that had an enrolment of more than 100 schoolchildren were selected for this study.

Peninsular Malaysia, also known as West Malaysia, covers a land area of 1 31 587 km² that extends 740 km from north (bordering Thailand) to south (bordering Singapore) and its maximum width is 322 km. It boasts a tropical climate; warm and humid all year round with temperatures ranging from 21 to 32 °C and average humidity of 90%. Moreover, rainfall is plentiful with an average of 2300 mm per year, with thick rainforest (Wong *et al.*, 2009).

Study population

This study was carried out among Orang Asli schoolchildren who were residents in the above-mentioned districts. Orang Asli (a Malay term that can be translated as 'original people' or 'the first people') are the aboriginal minority people of Peninsular Malaysia who accounts for 0.6% of the country's total population (i.e. approximately 180 000 people) (Department of Statistics Malaysia, 2010). They comprised of three main tribal groups: Senoi, constituting the largest group (54.9%), Proto-Malay (42.3%) and Negrito (2.8%). Within these three tribal groups, there are a total of 19 ethnic groups (SyedHussain *et al.*, 2017). The largest Orang Asli population reside in the states of Pahang and Perak, while the smallest can be found in the states of Penang and Perlis. Most Orang Asli communities live in remote forest areas but some live in the vicinity of suburban areas. A majority are marginalized socioeconomically and culturally, with a significant population below the poverty line, and about half of them are classified as living in hard-core poverty (SyedHussain *et al.*, 2017). Moreover, Orang Asli have an average life expectancy of 53 years, compared to the national average of 73 years, and an infant mortality rate of 51.7 per 1000 live births compared to the national rate of 8.9 (Masron *et al.*, 2013; Odani, 2017). Some Orang Asli work for wages in rubber tapping, in oil palm plantations as labourers or as factory workers, while many other Orang Asli are engaged in other kinds of daily activities to earn a living by selling jungle products such as fruit, rattan and bamboo (Masron *et al.*, 2013).

Sample size

A randomized, open-label controlled intervention trial (Trial Registration: clinicaltrials.gov; identifier: NCT03930901) was designed with the aim of investigating the impact of an improved health education learning package in controlling IPIs among Orang Asli schoolchildren. The cross-sectional study discussed herein represents the baseline assessment before randomization and intervention.

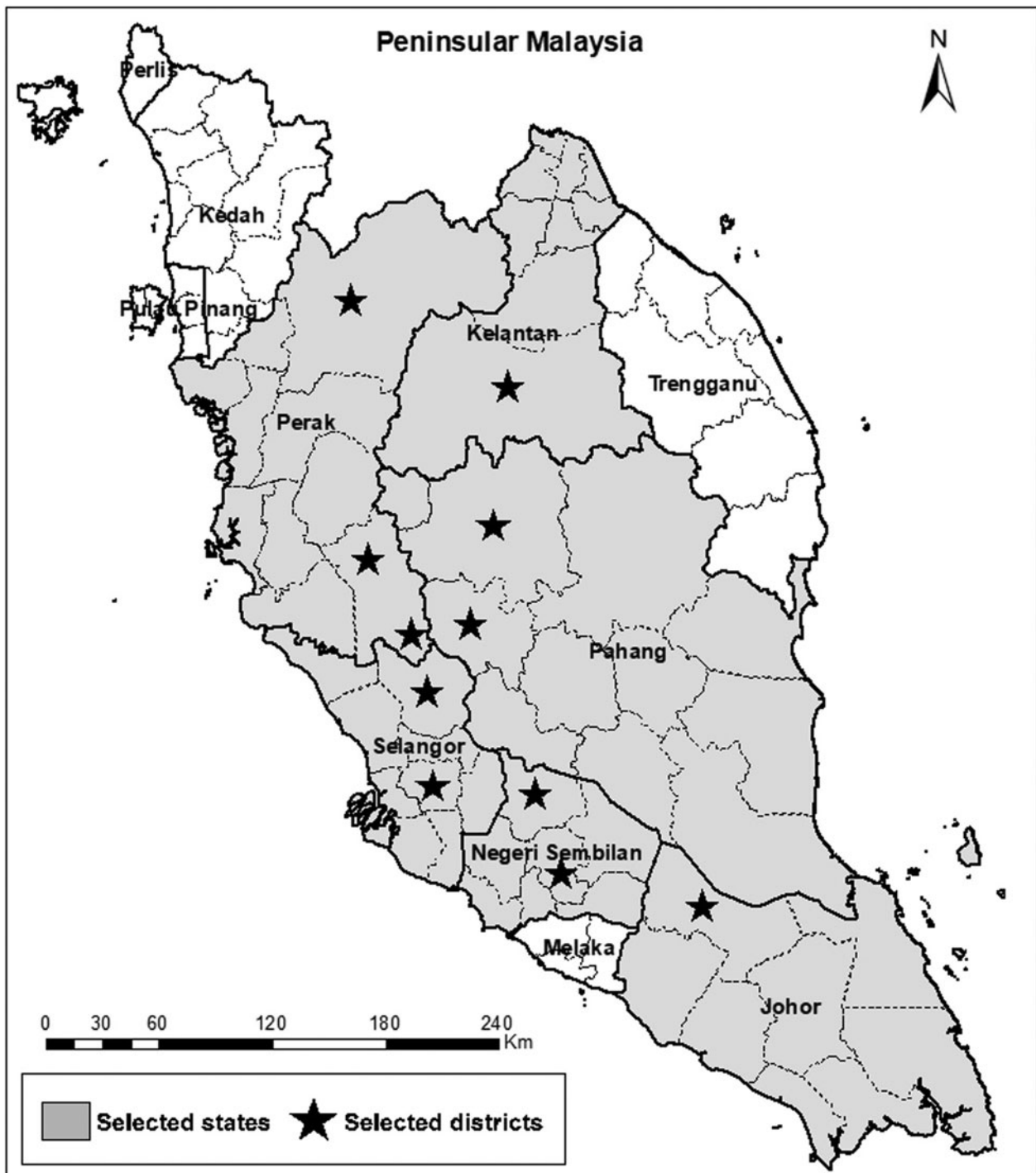


Fig. 1. A geographic map showing Peninsular Malaysia and the districts involved in the study (11 districts within six states). The map was created using the Esri ArcGIS 10.7 software.

The minimum sample size required for the main interventional study was calculated as 924 children; 462 per intervention arm (154 children from each tribe, including 10% to avoid the effects of dropouts and potential losses). It was estimated that this number would give the study at least 80% power at 5% significance to detect at least 10% difference in the prevalence and intensity of IPIs between the intervention group and the control group. This calculation assumed that 70% of Orang Asli children have IPIs (Nasr *et al.*, 2013; Al-Delaimy *et al.*, 2014; Elyana *et al.*, 2016). This sample size was judged safe enough to achieve a final sample size of 646, a figure that was calculated separately for the cross-sectional baseline survey at a 5% level of significance, a 95%

confidence level, and a design effect of 2 (Lwanga and Lemeshow, 1991). All of the eligible children who were present during our visit to each of the selected schools were invited to participate in the baseline study (universal sampling).

According to the official rolls, a total of 2763 children were enrolled in the 13 selected primary schools. However, at the time of our visits, there were 1914 children in the targeted schools all of whom were invited to participate in this study. Of these, 96 refused to participate and 676 did not deliver stool samples in the next 2 days following our visits. Hence, 1142 children (49.4% boys and 50.6% girls) who delivered suitable fecal samples for examination and complete questionnaire data were included in this study.

Questionnaire survey

A pre-tested structured questionnaire was used to collect information about the demographic (age, sex and family size) and socio-economic (e.g. parents' educational and occupational status, and family monthly income) background of the participants. Moreover, information about personal hygiene (e.g. washing hands before eating and after using the toilet, wearing shoes when outside the house, cutting nails periodically and washing vegetables/fruit before eating), health (e.g. history of IPIs and presence of any signs and symptoms particularly those related to gastrointestinal infections) and household characteristics (e.g. type of house, availability and type of toilets, sources of drinking water and presence of domestic animals) was also obtained. The types of toilet facility in the study area were categorized into two groups; improved (pour flush toilet) and unimproved (pit latrine without a slab) in line with the WHO/UNICEF criteria (WHO and UNICEF, 2015). Similarly, types of drinking water source were categorized into improved water sources (i.e. piped water supply) and unimproved water sources (e.g. wells, streams and rain).

The children were interviewed (face to face) by two assistants, one from the children's respective school and one from the Department of Parasitology, University of Malaya. Both assistants received proper training on the purpose of the study and on the administration of the questionnaire. A separate sheet containing questions to obtain demographic information about the children's parents was given to the children to hand to their parents to complete and was collected back the next day.

Fecal examination

Each child received a clean wide-mouth 100 mL screw-capped container clearly labelled with child's name and unique reference number for the collection of fecal samples. A clear explanation of how to collect the stool sample, the suitable amount of sample, and how to avoid possible contamination in the course of collection at home was provided to the participants in their classrooms. The children were instructed to bring their early morning stool samples the next day. Upon collection, the samples were placed into zipped plastic bags, kept in suitable cool boxes, and transported (within 2–8 h of collection) to the Department of Parasitology, University of Malaya, Kuala Lumpur for parasitological examination.

To detect *S. stercoralis*, the fecal samples were examined by using four different methods. First, the samples were examined using a direct smear or wet mount, using normal saline and iodine. Second, the samples were examined using the FES method (Cheesbrough, 2005). Third, the Koga APC method, which is one of the most sensitive techniques, was used to detect *S. stercoralis* and/or hookworm larvae (Koga *et al.*, 1991; Pocaterra *et al.*, 2017). *Strongyloides stercoralis* larvae were identified based on morphology (i.e. size of the buccal cavity, presence of genital primordium in rhabditiform larvae, and presence of forked tail-end in filariform larvae). Fourth, an aliquot of each fecal sample was preserved in 70% ethanol alcohol and kept at 4 °C until subsequent DNA extraction, and conventional polymerase chain reaction (PCR) assay to detect *S. stercoralis*. The samples were considered positive *via* the detection of *S. stercoralis* larvae DNA using PCR or by using any of the parasitological methods with confirmation by PCR. For those fecal samples that were found to be negative by PCR but positive by agar culture, the larvae collected from the positive agar cultures were examined by PCR to confirm the presence of *S. stercoralis*.

DNA extraction

About 500 mg of each ethanol-alcohol-preserved fecal sample was shaken vigorously and centrifuged at 1000 rpm for 2 min. The

resulting pellet was washed twice with phosphate-buffered saline (0.01 M, pH 7.2) and then used for genomic DNA extraction using the commercially available QIAamp® DNA stool Mini Kit (QIAGEN, Hilden, Germany), according to the manufacturer's instructions. The extracted DNA was then eluted with 100 µl of the AE elution buffer (included in the kit) and quantified using a Micro UV-Vis fluorescence spectrophotometer (Malcom e-spect, Tokyo, Japan). The extracted DNA was then kept at –20 °C until it was subjected to PCR amplification.

DNA amplification and sequencing

Strongyloides stercoralis-specific primer Stro18S-1530 (5'-GAA TTCCAAGTAAACGTAAGTCATTAGC-3') and Stro18S-1630 (5'-TGCCTCTGGATATTGCTCAGTTC-3') were used as an upstream and downstream oligonucleotide pair, respectively, to amplify a 101 base pair (bp) region of *S. stercoralis* 18S rRNA (GenBank accession number AF279916), as described by Verweij *et al.* (2009). The PCR was optimized and run using MyCycler thermal cycler (Bio-Rad, CA, USA) in a final volume of 25 µL of the reaction mixture that contained 1X of ExPrime Taq Master Mix (Genet Bio, Korea), 200 mM of each primer and 1 µl of DNA. The PCR thermal conditions included an initial denaturing step at 94 °C for 5 min followed by 40 cycles of 30 s at 94 °C, 60 s at 60.5 °C and 60 s at 72 °C, and a final extension step at 72 °C for 10 min. The PCR products were then visualized in 2.5% agarose gel stained with Sybr® safe DNA gel stain (Invitrogen, CA, USA) using a UV documenting system (Bio-Rad, Hercules, CA, USA).

The DNA that was extracted from the filariform larvae of *S. stercoralis* obtained from positive APC was subjected to PCR amplification and DNA sequencing in both directions. The resulting sequences were checked in NCBI BLAST to confirm the *S. stercoralis* species, and those with 100% specificity were used as a positive control.

Statistical analysis

Data analysis was performed using the IBM SPSS Statistics, version 18.0 (IBM Corporation, New York, USA). The dependent variable (i.e. *S. stercoralis* infection status) and the independent variables (i.e. demographic, socioeconomic, household and behavioural variables) were treated as categorical variables and presented as frequencies and percentages. All the variables were coded as binary dummy variables (i.e. 0 and 1). For instance, *S. stercoralis* infection (yes = 1, no = 0); age (>10 years = 1, ≤10 years = 0); gender (male = 1, female = 0); wearing shoes when outside (no = 1, yes = 0); and type of drinking water source (unimproved = 1, improved = 0). Pearson's χ^2 test was used to test the association between *S. stercoralis* infection prevalence and each of the explanatory variables. The odds ratio (OR) and 95% CI were calculated. Also, in order to identify the risk factors that were significantly associated with *S. stercoralis* infection, all the variables that showed associations with $P \leq 0.25$ in the univariate analysis were included in a multivariate logistic regression analysis, as suggested in the literature (Bendel and Afifi, 1977). In addition, the population attributable risk fraction (PARF) was calculated for the significantly associated risk factors (Rockhill *et al.*, 1998). A *P* value of <0.05 was considered as the level of significance.

Results

General characteristics of the participants

A total of 1142 schoolchildren aged 8–12 years, with a mean age of 10.19 years (s.d. = 1.36), attending 13 primary schools in six

Table 1. General characteristics of Orang Asli schoolchildren who participated in the study ($n = 1142$)

Characteristics	n (%)
Age groups	
>10 years	523 (45.8)
≤10 years	619 (54.2)
Gender	
Boys	564 (49.4)
Girls	578 (50.6)
Tribe	
Senoi	560 (49.0)
Proto-Malay	433 (37.9)
Negrito	149 (13.0)
Socioeconomic status	
Fathers' education level (at least 6 years)	603 (52.8)
Mothers' education level (at least 6 years)	626 (54.8)
Low household income (<RM500)	752 (65.8)
Working fathers	458 (40.1)
Working mothers	172 (15.1)
Large family size (≥7 members)	538 (47.1)
Improved water sources (piped water supply)	620 (54.3)
Electricity	549 (51.9)
Presence of improved toilet in the house	574 (50.3)
Presence of domestic animals at household	807 (70.7)

All values are number (%). RM, Malaysian Ringgit; US\$1 = RM4.05.

states of Peninsular Malaysia participated in this study. Of these, 49.4% were male and 50.6% were female. The highest percentage of children (49%) belonged to the Senoi tribe, while the lowest percentage belonged to the Negrito tribe (13%). About half of the mothers (54.8%) and fathers (52.8%) had completed their primary education (i.e. at least 6 years of formal education). Approximately two thirds of the participants had a low household monthly income [Malaysian Ringgit (RM) 500; US\$1 = RM4.05]. Most of the houses were made of wood or bamboo and some houses were small, single-storey, concrete terraced houses built by the government to replace the old bamboo and wooden houses. Almost half of the houses had an improved (piped, gravity-fed) water supply and electricity. Over one-third (40.1%) of the fathers worked as labourers in oil palm or rubber plantations. Full details of the general characteristics of the participants are presented in [Table 1](#).

Prevalence and distribution of *S. stercoralis* infections

Based on the above-described examination of the fecal samples for the presence of *S. stercoralis* larvae, 15.8% (180/1142) of the children were found to be infected with *S. stercoralis*. As shown in [Fig. 2](#), the results indicated that the highest detection rate was achieved by using the Koga APC method (174/1142; 15.2%; 95% CI = 13.12–17.28), followed by PCR assay, which detected *S. stercoralis* in 157 samples (13.7%; 95% CI = 11.71–15.69); however, the difference in rates by these methods as well as the combined rate was not statistically significant ($P > 0.05$). On the other hand, *S. stercoralis* was detected in only 15 (1.3%; 95% CI = 0.64–1.96) and 2 (0.2%; 95% CI = –0.06–0.46) samples by FES and direct smear, respectively. Six samples that were found to be negative

by agar culture, FES and direct smear were identified as positive by PCR assay.

[Table 2](#) shows the distribution of *S. stercoralis* among the participants. The prevalence of *S. stercoralis* infection increased significantly with age ($\chi^2 = 26.766$; $P < 0.001$), with the highest prevalence reported among children in Grade 6, aged 12 years (21.9%), while the lowest prevalence was found among children in Grade 2, aged 8 years (7.3%). Also, the prevalence of *S. stercoralis* infection among boys was significantly higher than among girls (18.8 vs 12.8%; $\chi^2 = 7.718$; $P = 0.005$). In addition, the prevalence of *S. stercoralis* infection differed significantly among the six states ($\chi^2 = 40.149$; $P < 0.001$), with the highest prevalence of *S. stercoralis* infection found among children in the states of Johor (25%) and Pahang (24.2%) and the lowest found among children in Negeri Sembilan (6.7%). Moreover, the prevalence of *S. stercoralis* infection was significantly higher among children belonging to the Proto-Malay (17.8%) and Senoi (16.2%) tribes compared to those belonging to the Negrito tribe (8.1%) ($\chi^2 = 8.100$; $P = 0.017$). Furthermore, children belonging to the Jakun sub-tribe of the Proto-Malay tribal group had the highest prevalence of infection (25.0%) compared to children of other sub-tribes ($\chi^2 = 18.349$; $P = 0.001$).

Factors associated with *S. stercoralis* infection

[Table 3](#) shows the result of the univariate analysis conducted to determine the association of *S. stercoralis* infection with the demographic, socioeconomic, housing and behavioural variables. The results showed that children aged >10 years had a significantly higher prevalence of strongyloidiasis than younger children (21.0 vs 11.3%; $P < 0.001$). Also, children of the Proto-Malay (17.8 vs 8.1%; $P = 0.004$) and Senoi (16.2 vs 8.1%; $P = 0.012$) tribes had a significantly higher strongyloidiasis prevalence when compared with children of the Negrito tribe. Similarly, a higher prevalence of strongyloidiasis was found among children who lived in houses without improved toilet facilities (19.7 vs 11.8%; $P < 0.001$) and children who used unimproved sources of drinking water (19.5 vs 12.6%; $P = 0.001$) when compared to their counterparts with improved toilet facilities and improved sources of drinking water. With regard to personal hygiene variables, it was found that children who practised indiscriminate/open defecation had a significantly higher prevalence of strongyloidiasis than those who always used toilets (18.5 vs 8.1%; $P < 0.001$). Similarly, the prevalence was significantly higher among children who did not wear shoes when outside their house (22.2 vs 12.3%; $P < 0.001$), and children who did not wash their hands after defecation (19.8 vs 14.2%; $P = 0.020$) compared to those who wore shoes or slippers and those who practised proper hand washing after defecation.

In the multivariate logistic regression analysis, six variables were identified as significant risk factors of *S. stercoralis* infection among Orang Asli schoolchildren ([Table 4](#)). The Hosmer-Lemeshow test showed that the model fit the data well [$\chi^2 = 6.789$ (8 degrees of freedom); $P = 0.560$]. Children aged > 10 years and boys had higher odds of being infected by strongyloidiasis compared to younger children [adjusted odds ratio (aOR) = 1.91; 95% CI = 1.36–2.68] and girls (aOR = 1.52; 95% CI = 1.09–2.13), respectively. Also, children belonging to the Proto-Malay and Senoi tribes had 2.50 (95% CI = 1.28–4.85) and 1.93 (95% CI = 1.01–3.70) higher odds of contracting strongyloidiasis compared to children belonging to the Negrito tribe. Moreover, children who did not have improved sources of drinking water in their houses had 2.91 odds (95% CI = 1.50–5.66) of having strongyloidiasis compared to those who had safe piped sources of drinking water. In addition, practising indiscriminate/open defecation and not wearing shoes when outside the house increased the

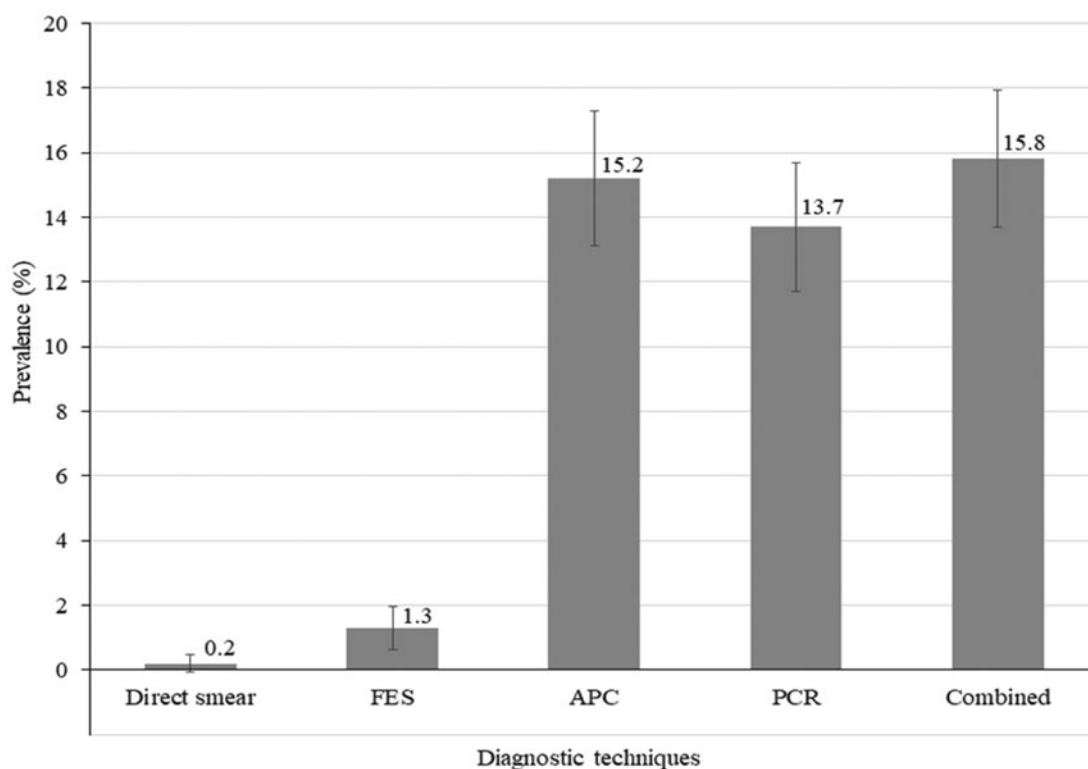


Fig. 2. Prevalence (%) of *Strongyloides stercoralis* identified by each technique, and by the combination of the four techniques among the participants ($n = 1142$). FES, formalin-ether sedimentation; APC, agar plate culture. Error bars represent 95% confidence interval of the proportion.

children's odds of having strongyloidiasis by 2.81 (95% CI = 1.75–4.50) and 1.91 times (95% CI = 1.37–2.67), respectively, compared to their counterparts.

Finally, the results of the PARF analysis showed that almost half (48.7%) of the strongyloidiasis cases among Orang Asli schoolchildren could be reduced if all children avoided indiscriminate/open defecation in rivers and surrounding areas. Moreover, the prevalence of strongyloidiasis could be reduced by 21.8% if the children in this population practised good standards of personal hygiene; particularly, wearing shoes or slippers when going or playing outside the house. In addition, 20.2% of the cases could be avoided if these children were provided with improved sources of drinking water in their houses.

Discussion

This study revealed a relatively high prevalence (15.8%; 180/1142) of strongyloidiasis among the study population, which consisted of Orang Asli schoolchildren in Peninsular Malaysia. The prevalence of this infection was detected by using different diagnostic techniques, namely the direct smear, FES, Koga APC and PCR. Hence, the current study provides reliable and comprehensive evidence on the occurrence of *S. stercoralis* among Orang Asli communities. Moreover, it is the first in Malaysia to use the Koga APC method on fecal samples.

The prevalence of strongyloidiasis reported by the current study is significantly higher than that reported in previous studies conducted in Malaysia. For instance, a recent community-based study that attempted to screen 236 participants in indigenous communities in Sarawak (East Malaysia) for *S. stercoralis* infection showed that 26 participants (11%) were seropositive for strongyloidiasis by ELISA assay, while none of the collected fecal samples was positive for *S. stercoralis* larvae by direct smear and FES (Ngui *et al.*, 2016). On the other hand, a recent study among Negrito communities reported that only seven out of 416 (1.7%) fecal

samples were found positive for *S. stercoralis*; however, the methods used (direct smear, FES and Kato-Katz techniques) have a low sensitivity for *S. stercoralis* (Muslim *et al.*, 2019). By using PCR, the current study found 157 *S. stercoralis*-positive cases (equating to 13.7% of the participants), whereas only five and three cases were reported by previous studies in Sarawak and Selangor, respectively (Ahmad *et al.*, 2013; Ngui *et al.*, 2016). Using a pentaplex real-time PCR analysis, Basuni *et al.* revealed that 30 out of 77 (39%) individuals presenting with abdominal symptoms in two district hospitals in Sarawak, East Malaysia were positive for *S. stercoralis* (Basuni *et al.*, 2011).

In neighbouring Southeast Asian countries, higher prevalence rates of *S. stercoralis* infection have been reported in Cambodia (Forrer *et al.*, 2018), Lao PDR (Vonghachack *et al.*, 2015) and Thailand (Laoraksawong *et al.*, 2018), while much lower prevalence rates were reported in Indonesia (Wiria *et al.*, 2013) and Myanmar (Aung *et al.*, 2018). Nevertheless, it is believed that strongyloidiasis is likely to have a much higher burden in Southeast Asia than currently indicated in the available literature (Schär *et al.*, 2016) not least because the region exhibits conditions that favour high transmission of strongyloidiasis, such as humid and wet climates, inadequate sanitary conditions and poverty (Al-Mekhlafi *et al.*, 2013; Schär *et al.*, 2013).

Elsewhere, strongyloidiasis is hyperendemic among aboriginal populations in Australia, with a reported prevalence ranging from 35 to 60% (Kearns *et al.*, 2017). In addition, endemic foci for *S. stercoralis* have been reported in Europe and North America (Asundi *et al.*, 2019). For instance, until 2018, strongyloidiasis was reported in a total of 1083 Spanish-born individuals without a history of travel to endemic areas (Barroso *et al.*, 2019). Also, an earlier study in Canada revealed a prevalence rate of 76.6 and 11.8% among Cambodian and Vietnamese refugees, respectively (Gyorkos *et al.*, 1990).

In the current study, the APC method showed superior sensitivity (174 cases) in detecting *S. stercoralis* larvae in fecal samples

Table 2. Distribution of *Strongyloides stercoralis* infection among Orang Asli schoolchildren in Malaysia (n = 1142)

Variable	<i>Strongyloides stercoralis</i> infection		χ^2	P
	No. examined	Infected n (%)		
Grade			26.766	<0.001
2	164	12 (7.3)		
3	171	14 (8.2)		
4	284	45 (15.8)		
5	267	53 (19.9)		
6	256	56 (21.9)		
Location (State)			40.149	<0.001
Johor	140	35 (25.0)		
Kelantan	197	26 (13.2)		
Negeri Sembilan	135	9 (6.7)		
Pahang	186	45 (24.2)		
Perak	326	32 (9.8)		
Selangor	158	33 (20.9)		
Tribe			8.100	0.017
Senoi	560	91 (16.2)		
Proto-Malay	433	77 (17.8)		
Negrito	149	12 (8.1)		
Sub-tribe			18.349	0.001
Semai	363	65 (17.9)		
Temiar	197	26 (13.2)		
Jakun	140	35 (25.0)		
Temuan	293	42 (14.3)		
Jahai	149	12 (8.1)		

compared to direct smear (two cases), FES (15 cases) and conventional PCR (157 cases), with six samples found positive only by PCR. These results are consistent with those reported elsewhere (Requena-Méndez *et al.*, 2013; Amor *et al.*, 2016; Aung *et al.*, 2018; Tuyizere *et al.*, 2018). It is also interesting to note that the number of positive cases detected by the combined methods was not significantly different from using APC. Although 17 agar culture-positive fecal samples were found to be negative by PCR, these samples were confirmed by amplification of *S. stercoralis* larvae collected from the agar culture. The differing results by APC and PCR could be explained by the low intensity of the larvae in those samples, as shown by the agar culture, as well as the presence of fecal inhibitors that might not have been completely removed prior to PCR (Knopp *et al.*, 2014; Requena-Méndez *et al.*, 2014).

The current study investigated the possible risk factors of *S. stercoralis* infection among the study participants and revealed three sets of key risk factors; (1) demographic: the age of >10 years, tribe (Senoi and Proto-Malay) and male gender; (2) socio-economic: using an unimproved water source for drinking water; and (3) behavioural: practising indiscriminate defecation and not wearing shoes when outside the house. The age-related findings of the current study showed that children aged >10 years are generally at a higher risk of *S. stercoralis* infection compared to younger children, which is consistent with the general conclusion drawn in prior research (Schär *et al.*, 2013). This finding is also in

agreement with many previous studies that have linked this infection to increasing age because *S. stercoralis* can be sustained in infected individuals for decades by means of autoinfection (Prendki *et al.*, 2011; Conlan *et al.*, 2012; Khieu *et al.*, 2014; Aung *et al.*, 2018). Hence, a higher prevalence of *S. stercoralis* infection is expected among adult individuals in Orang Asli communities. However, further studies are required to confirm this conjecture.

The results of the current study also showed that boys were more prone to carry the infection than girls, which is consistent with previous studies in different countries (Steinmann *et al.*, 2007; Conlan *et al.*, 2012; Khieu *et al.*, 2014; Tuyizere *et al.*, 2018; Gétaz *et al.*, 2019). Interestingly, previous studies have reported a consistent male-bias in helminth infections, particularly hookworm infection, and have suggested sex-related differences in susceptibility to infection arising from immunosuppression associated with male hormones (Poulin, 1996; Moore and Wilson, 2002; Brooker *et al.*, 2004). However, the reported sex-dependent difference could be also attributed to higher exposure among males who are more involved in outdoor activities such as playing football, swimming in streams or ponds as well as in helping their parents in farming activities.

The findings also showed that children belonging to the Proto-Malay (17.8%) and Senoi (16.2%) tribes were more likely to be infected with *S. stercoralis* compared to those in the Negrito tribe (8.1%). Indeed, previous studies have shown that IPIs other than *S. stercoralis* are prevalent to varying degrees in all Orang Asli communities throughout Peninsular Malaysia. For instance, the prevalence of *Trichuris trichiura* infection is significantly higher among the Negrito tribe and that of the *Ascaris lumbricoides* infection is significantly higher among the Senoi tribe, whereas the prevalence of hookworm infection is comparable among the tribes (Anuar *et al.*, 2014). A similar profile of infections was also found according to location, with Pahang and Selangor states having the highest prevalence. As *S. stercoralis* was reported in all targeted states, the significant differences could be explained by the different socioeconomic status and behavioural factors among the tribes rather than climatic or environmental factors. Most of the Senoi and Proto-Malay tribes including the Jakun, Semai, Temiar and Temuan communities practise permanent agriculture and manage their own rubber, oil palm or cocoa farms or engage in 'shifting cultivation' (hill rice cultivation), in which human/animal fecal materials are used as fertilizer (night soil). They also prefer to live in remote areas located deep in the jungle, which have inadequate sanitary facilities and are far away from healthcare facilities (Masron *et al.*, 2013; Choy *et al.*, 2014). These conditions provide the ideal ecological and economic setting for a high burden of *S. stercoralis* infection, and this may explain the significantly higher prevalence among these tribes compared to the Negrito tribe.

With regards to the socioeconomic and behavioural risk factors of *S. stercoralis* infection, this study found that children who practised indiscriminate defecation used an unimproved water source for drinking water, and walked barefooted when outside the house were more likely to be infected than their counterparts. These findings are consistent with those of previous studies in other Asian countries (Khieu *et al.*, 2014; Senephansiri *et al.*, 2017; Aung *et al.*, 2018). Moreover, several studies have identified poor personal hygienic practices as significant predictors of IPIs among Orang Asli populations (Al-Mekhlafi *et al.*, 2006; Ahmed *et al.*, 2012; Anuar *et al.*, 2012; Elyana *et al.*, 2016; Muslim *et al.*, 2019).

Given the fact that the filariform infective larvae of *S. stercoralis* are found in soil and mainly infect humans through skin penetration, it is not particularly surprising that the result of the PARF analysis showed that wearing shoes when outside the

Table 3. Univariate analysis of factors associated with *Strongyloides stercoralis* infection among Orang Asli schoolchildren in Malaysia (n = 1142)

Variables	<i>S. stercoralis</i> infection		OR	95% CI	P
	No. examined	% Infected			
<i>Demographic factors</i>					
Age group (years)					
>10	523	21.0	2.09	1.51–2.89	<0.001*
≤10	619	11.3	1		
Gender					
Boys	564	18.8	1.58	1.14–2.18	0.005*
Girls	578	12.8	1		
Tribe					
Senoi	560	16.2	2.22	1.18–4.17	0.012*
Proto-Malay	433	17.8	2.47	1.30–4.68	0.004*
Negrato	149	8.1	1		
Family size					
≥7 members (large)	538	15.4	0.95	0.69–1.31	0.770
<7 members	604	16.1	1		
<i>Socioeconomic factors</i>					
Father's educational level					
Non educated	539	14.7	0.85	0.62–1.18	0.333
Educated (≥6 years)	603	16.7	1		
Mother's educational level					
Non educated	516	15.1	0.92	0.66–1.26	0.587
Educated (≥6 years)	626	16.3	1		
Father's employment status					
Not working	684	16.8	1.22	0.88–1.70	0.234
Working	458	14.2	1		
Mother's employment status					
Not working	970	15.6	0.91	0.59–1.41	0.668
Working	172	16.9	1		
Household monthly income					
<RM500	752	15.3	0.90	0.65–1.26	0.564
≥RM500	390	16.7	1		
Presence of improved toilet in the house					
No	568	19.7	1.83	1.32–2.53	<0.001*
Yes	574	11.8	1		
Source of drinking water					
Unimproved source (river, rain)	522	19.5	1.69	1.22–2.33	0.001*
Improved source (pipe)	620	12.6	1		
Source of domestic water					
Unimproved source (river, rain)	642	17.1	1.27	0.92–1.76	0.149
Improved source (pipe)	500	14.0	1		
Presence of domestic animals					
Yes	807	16.5	1.21	0.84–1.73	0.301
No	335	14.0	1		
Presence of electricity					
No	593	15.7	1.01	0.74–1.39	0.939

(Continued)

Table 3. (Continued.)

Variables	<i>S. stercoralis</i> infection		OR	95% CI	<i>P</i>
	No. examined	% Infected			
Yes	549	15.8	1		
<i>Personal hygiene factors</i>					
Washing hands before eating					
No	473	13.5	0.75	0.54–1.04	0.082
Yes	669	17.3	1		
Washing hands after defecation					
No	324	19.8	1.50	1.06–2.09	0.020*
Yes	818	14.2	1		
Indiscriminate defecation					
Yes	845	18.5	2.58	1.64–4.05	<0.001*
No	297	8.1	1		
Eating soil habit (geophagy)					
Yes	198	13.1	0.78	0.50–1.21	0.264
No	944	16.3	1		
Cutting nails periodically					
No	576	16.7	1.15	0.83–1.58	0.397
Yes	566	14.8	1		
Wearing shoes when going outside					
No	396	22.2	2.03	1.47–2.80	<0.001*
Yes	746	12.3	1		
Washing fruits before eating					
No	370	15.7	0.99	0.71–1.40	0.956
Yes	772	15.8	1		
Washing vegetables before eating					
No	508	15.7	0.99	0.73–1.38	0.991
Yes	634	15.8	1		

RM, Malaysian Ringgit; US\$1 = RM4.05. OR, odds ratio. CI, confidence interval.
*Significant association ($P < 0.05$).

house will help in preventing strongyloidiasis. However, this study showed that using an unimproved water source for drinking water (aOR = 2.91) and indiscriminate defecation (aOR = 2.81) was the most significant risk factors of *S. stercoralis* infection. Based on the PARF results, about half (48.7%) and one fifth (21.7%) of the strongyloidiasis cases could be prevented if all children frequently used improved toilets for defecation and had an improved source for drinking water in their households, respectively. In Malaysia, rivers are considered the lifeblood of Orang Asli populations and are still the main water source for drinking water and for domestic use (e.g. bathing, washing and swimming). Unfortunately, rivers are also the preferred site for defecation, particularly among Orang Asli children who have also been noted to defecate indiscriminately close to their houses or within the village confines (Al-Mekhlafi *et al.*, 2008; Elyana *et al.*, 2016). Young children commonly select shallow areas of rivers and streams where the water flows more slowly so they can sit, individually or in small groups, to defecate and then cleanse themselves after defecation. Thus, this untreated water is likely to be highly contaminated with intestinal parasitic ova, cysts, oocysts and/or larvae (Lim and Ahmad, 2004; Lee *et al.*, 2014). In such epidemiological situation, water, sanitation and hygiene may

have a crucial role in the prevention and control of STH infections, including strongyloidiasis.

Although not found to be significant in this study, contact with domestic animals (mainly dogs) has also been identified as a significant risk factor of *S. stercoralis* infection, and thus zoonotic strongyloidiasis, with dogs as reservoirs, has been suggested (Thamsborg *et al.*, 2017). Moreover, clinical and subclinical cases of *S. stercoralis* infection have been increasingly reported among dogs in Europe (Paradies *et al.*, 2017; Iatta *et al.*, 2019). Similarly, dogs in rural Cambodia have been found to carry two populations of *S. stercoralis*, one of which is shared with humans (Jaleta *et al.*, 2017). In Malaysia, *S. stercoralis* larvae have been detected in fecal samples collected from dogs in rural Orang Asli communities and in soil samples collected from an urban area in Kuala Lumpur (Azian *et al.*, 2008). Interestingly, larvae of *Strongyloides spp.* have been detected in domestic pig-tailed macaques working in the harvesting of coconuts in Kelantan state (Choong *et al.*, 2019). Furthermore, *S. stercoralis* larvae have been detected in common vegetables and herbs in the city of Kota Bharu in Kelantan (Zeehaida *et al.*, 2011).

Based on current and previous findings, it is possible to suggest a number of different scenarios for the transmission of *S. stercoralis*

Table 4. Multivariate analysis of factors associated with *Strongyloides stercoralis* infection among Orang Asli schoolchildren in Malaysia (n = 1142)

Variables	<i>S. stercoralis</i> infection			Wald-test <i>P</i> value
	aOR	95% CI	Wald	
Age group (>10 years)	1.91	1.36–2.68	14.862	<0.001*
Gender (male)	1.52	1.09–2.13	6.105	0.014*
Tribe (Senoi)	1.93	1.01–3.70	3.957	0.047*
Tribe (Proto-Malay)	2.50	1.28–4.85	7.335	0.007*
Father's employment status (not working)	1.17	0.83–1.66	0.849	0.357
Presence of improved toilet in house (no)	1.33	0.89–1.97	1.923	0.165
Source of drinking water (unimproved source)	2.91	1.50–5.66	9.915	0.002*
Source of domestic water (unimproved sources)	0.54	0.28–1.06	3.186	0.074
Washing hands before eating (no)	0.72	0.51–1.02	3.546	0.059
Washing hands after defecation (no)	1.39	0.97–1.99	3.118	0.077
Indiscriminate defecation (yes)	2.81	1.75–4.50	17.603	<0.001*
Wearing shoes when outside (no)	1.91	1.37–2.67	14.439	0.001*

aOR, adjusted odds ratio. CI, confidence interval.

*Significant risk factors of *S. stercoralis* infection ($P < 0.05$).

infection among the studied Orang Asli children. First, filariform larvae in the soil can penetrate the intact skin of the feet when children walk or play barefooted or help their parents in farms fertilized by human and animal excreta. Second, while defecating in the river, filariform larvae in the water may penetrate the exposed skin of the feet, legs, anal or buttock area and hands. Third, recalling some valiant self-experimentation in the early 1900s to induce *S. stercoralis* infection by oral ingestion of larvae in water (Grove, 1996), water-borne transmission by ingesting the larvae in untreated drinking water collected from rivers may occur among these children. Fourth, food-borne transmission through the ingestion of larvae in contaminated vegetables or fruit may also take place. However, except for the first scenario, which has been documented as the exclusive mode of transmission, further studies are required to confirm the other suggested scenarios.

The strengths of this study include the screening of a large number of children ($n = 1142$) from the main three Orang Asli tribes and the use of a combination of screening methods to increase the sensitivity of *S. stercoralis* diagnosis. However, we also acknowledge some limitations that should be considered when interpreting the results of this study. For instance, the study design (i.e. cross-sectional) limited our ability to confirm the existence of a causal association between *S. stercoralis* infection and the reported significant risk factors. Moreover, only one fecal sample from each participant was examined for *S. stercoralis* instead of the three repeated samples due to the level of cooperation of the children and the cultural beliefs of the Orang Asli that oppose the giving of fecal samples. Previous studies have shown that there is a high risk that a single-sample examination will miss an infection because of the day-to-day variation in larval excretion, particularly in asymptomatic light *S. stercoralis* infections (Dreyer *et al.*, 1996; Uparanukraw *et al.*, 1999). Thus, the overall prevalence of *S. stercoralis* infection reported by the current study is likely to have been significantly underestimated.

In conclusion, this study revealed that *S. stercoralis* infection is prevalent among Orang Asli schoolchildren in Malaysia, particularly among Proto-Malay and Senoi tribes, and is thus a matter of serious concern. An age of >10 years, the male gender, indiscriminate defecation, using unimproved sources for drinking water and not wearing shoes when outside were identified as the significant risk factors of infection in the studied population. Orang Asli

populations in Peninsular Malaysia share similar epidemiological characteristics. Thus, the findings reported herein can be generalized to Orang Asli schoolchildren in other communities that were not included in this study. However, there is still a need for further studies among preschool children and adults.

Orang Asli children are vulnerable to many infectious diseases and can be considered partially immunocompromised as a result of a high level of malnutrition (Al-Mekhlafi *et al.*, 2005; Khor and Zalilah, 2008; Ahmed *et al.*, 2012). In such a situation, *S. stercoralis* infection may have severe consequences. Therefore, access to adequate diagnosis and treatment of strongyloidiasis is urgently needed and should be a public health priority for Orang Asli population. Moreover, specific measures to control this infection should be implemented in any efforts to improve the quality of life of the Orang Asli population as a whole. Proper health education regarding good personal hygiene, good sanitary practices, provision of improved water supply and adequate sanitation, as well as the implementation of periodic mass drug administration, will help in reducing the prevalence of *S. stercoralis* infection in these communities.

Acknowledgements. The authors wish to acknowledge the Department of Orang Asli Development (JAKOA), Ministry of Rural and Regional Development, Kuala Lumpur, Malaysia and the Departments of Education in the targeted states for their generous collaboration during this study. The authors also would like to express their appreciation to the support and cooperation of the headmasters, teachers and staff of the targeted schools. Special thanks also go to the children and their parents/guardians for their voluntary participation in this study.

Financial support. The work presented in this paper was funded by the University of Malaya High Impact Research Fund, HIR-MOHE (H-20001-00-E00051), and also by the University of Malaya Research Grants (RG331-15AFR). The funders had no role in study design, data collection and analysis, decision to publish, or preparation of the manuscript.

Conflict of interest. None.

Ethical standards. This study was carried out according to the guidelines laid down in the Declaration of Helsinki. The study protocol was approved by the Medical Ethics Committee of the University of Malaya Medical Centre, Malaysia (Reference number: 201731-4985). Before recruiting participants, meetings were held with the headmasters and teachers of schools to provide important information about the aims and methods of this study and their consents were obtained. In addition, the research team visited Orang

Asli communities in the targeted areas and met the heads of the villages and the parents of schoolchildren to inform them about the objectives and to request the involvement of their children in the study. They were informed that the involvement of their children in this study was totally voluntary and that they have the right to withdraw from the study at any point without having to give a reason. Written and signed or thumb-printed informed consents were collected from parents or guardians on behalf of their children before the commencement of the survey.

References

- Agrawal V, Agarwal T and Ghoshal UC (2009) Intestinal strongyloidiasis: a diagnosis frequently missed in the tropics. *Transactions of the Royal Society of Tropical Medicine and Hygiene* **103**, 242–246.
- Ahmed A, Al-Mekhlafi HM, Al-Adhroey AH, Ithoi I, Abdulsalam AM and Surin J (2012) The nutritional impacts of soil-transmitted helminths infections among Orang Asli schoolchildren in rural Malaysia. *Parasites and Vectors* **5**, 119.
- Ahmad AF, Hadip F, Ngui R, Lim YA and Mahmud R (2013) Serological and molecular detection of *Strongyloides stercoralis* infection among Orang Asli community in Malaysia. *Parasitology Research* **112**, 2811–2816.
- Al-Delaimy AK, Al-Mekhlafi HM, Lim YA, Nasr NA, Sady H, Atroosh WM and Mahmud R (2014) Developing and evaluating health education learning package (HELP) to control soil-transmitted helminth infections among Orang Asli children in Malaysia. *Parasites and Vectors* **7**, 416.
- Al Hadidi M, Shaaban H, Jumean KH and Peralta P (2018) Loeffler's syndrome secondary to hyperinfection by *Strongyloides stercoralis* associated with methotrexate in a patient with rheumatoid arthritis. *Journal of Global Infectious Diseases* **10**, 29–30.
- Al-Mekhlafi HM, Azlin M, Aini UN, Shaikh A, Sa'iah A, Fatmah MS, Ismail MG, Ahmad F, Aisah MY, Rozlida AR and Norhayati M (2005) Protein-energy malnutrition and soil-transmitted helminthiasis among Orang Asli children in Selangor, Malaysia. *Asia Pacific Journal of Clinical Nutrition* **14**, 188–194.
- Al-Mekhlafi MS, Azlin M, Nor Aini U, Shaikh A, Sa'iah A, Fatmah MS, Ismail MG, Firdaus MS, Aisah MY, Rozlida AR and Norhayati M (2006) Prevalence and distribution of soil-transmitted helminthiasis among Orang Asli children living in peripheral Selangor, Malaysia. *Southeast Asian Journal of Tropical Medicine and Public Health* **37**, 40–47.
- Al-Mekhlafi HMS, Surin J, Atiya AS, Ariffin WAW, Mahdy AKM and Abdullah HC (2008) Pattern and predictors of soil-transmitted helminth re-infection among aboriginal school children in rural peninsular Malaysia. *Acta Tropica* **107**, 200–204.
- Al-Mekhlafi HM, Lim YAL, Moktar N and Ngui R (2013) Soil-transmitted helminths: the neglected parasites. In Lim YAL and Vythilingam I (eds), *Parasites and Their Vectors*. Vienna: Springer, pp. 205–232.
- Amor A, Rodriguez E, Saugar JM, Arroyo A, López-Quintana B, Abera B, Yimer M, Yizengaw E, Zewdie D, Ayeubuzi Z, Hailu T, Mulu W, Echazú A, Krolewiecki AJ, Aparicio P, Herrador Z, Anegargie M and Benito A (2016) High prevalence of *Strongyloides stercoralis* in school-aged children in a rural highland of north-western Ethiopia: the role of intensive diagnostic work-up. *Parasites and Vectors* **9**, 617.
- Anuar TS, Al-Mekhlafi HM, Abdul Ghani MK, Abu Bakar E, Azreen SN, Salleh FM, Ghazali N, Bernadus M and Moktar N (2012) Molecular epidemiology of amoebiasis in Malaysia: highlighting the different risk factors of *Entamoeba histolytica* and *Entamoeba dispar* infections among Orang Asli communities. *International Journal for Parasitology* **42**, 1165–1175.
- Anuar TS, Salleh FM and Moktar N (2014) Soil-transmitted helminth infections and associated risk factors in three Orang Asli tribes in Peninsular Malaysia. *Scientific Reports* **4**, 4101.
- Asundi A, Beliaevsky A, Liu XJ, Akaberi A, Schwarzer G, Bisoffi Z, Requena-Méndez A, Shrier I and Greenaway C (2019) Prevalence of strongyloidiasis and schistosomiasis among migrants: a systematic review and meta-analysis. *Lancet Global Health* **7**, e236–e248.
- Aung MPPTH, Hino A, Oo KM, Win KK, Maruyama H, Htike WW and Nagayasu E (2018) Prevalence and associated risk factors of *Strongyloides stercoralis* infection in lower Myanmar. *Tropical Medicine and Health* **46**, 43.
- Azian MY, Sakhone L, Hakim SL, Yusri MY, Nurulysamzawaty Y, Zuhazam AH, Rodi IM and Maslawaty MN (2008) Detection of helminth infections in dogs and soil contamination in rural and urban areas. *Southeast Asian Journal of Tropical Medicine and Public Health* **39**, 205–212.
- Barroso M, Salvador F, Sánchez-Montalvá A, Bosch-Nicolau P and Molina I (2019) *Strongyloides stercoralis* infection: a systematic review of endemic cases in Spain. *PLoS Neglected Tropical Diseases* **13**, e0007230.
- Basuni M, Muhi J, Othman N, Verweij JJ, Ahmad M, Miswan N, Rahumatullah A, Aziz FA, Zainudin NS and Noordin R (2011) A pentaplex real-time polymerase chain reaction assay for detection of four species of soil-transmitted helminths. *American Journal of Tropical Medicine and Hygiene* **84**, 338–343.
- Bendel RB and Afifi AA (1977) Comparison of stopping rules in forward 'stepwise' regression. *Journal of American Statistics Association* **72**, 46–53.
- Bisoffi Z, Buonfrate D, Montresor A, Requena-Méndez A, Muñoz J, Krolewiecki AJ, Gotuzzo E, Mena MA, Chiodini PL, Anselmi M, Moreira J and Albonico M (2013) *Strongyloides stercoralis*: a plea for action. *PLoS Neglected Tropical Diseases* **7**, e2214.
- Bisoffi Z, Buonfrate D, Sequi M, Mejia R, Cimino RO, Krolewiecki AJ, Albonico M, Gobbo M, Bonafini S, Angheben A, Requena-Mendez A, Muñoz J and Nutman TB (2014) Diagnostic accuracy of five serologic tests for *Strongyloides stercoralis* infection. *PLoS Neglected Tropical Diseases* **8**, e2640.
- Brooker S, Bethony J and Hotez PJ (2004) Human hookworm infection in the 21st century. *Advance Parasitology* **58**, 197–288.
- Buonfrate D, Perandin F, Formenti F and Bisoffi Z (2017) A retrospective study comparing agar plate culture, indirect immunofluorescence and real-time PCR for the diagnosis of *Strongyloides stercoralis* infection. *Parasitology* **144**, 812–816.
- Cheesbrough M (2005) *District Laboratory Practice in Tropical Countries: Part 1*, 2nd edn. London: Cambridge University Press.
- Choong SS, Mimi Armiladiana M, Ruhil HH and Peng TL (2019) Prevalence of parasites in working pig-tailed Macaques (*Macaca nemestrina*) in Kelantan, Malaysia. *Journal of Medical Primatology* **48**, 207–210.
- Choy SH, Al-Mekhlafi HM, Mahdy MA, Nasr NN, Sulaiman M, Lim YA and Surin J (2014) Prevalence and associated risk factors of *Giardia* infection among indigenous communities in rural Malaysia. *Scientific Reports* **4**, 6909.
- Conlan JV, Khamlome B, Vongxay K, Elliot A, Pallant L, Sripa B, Blacksell SD, Fenwick S and Thompson RC (2012) Soil-transmitted helminthiasis in Laos: a community-wide cross-sectional study of humans and dogs in a mass drug administration environment. *American Journal of Tropical Medicine and Hygiene* **86**, 624–634.
- Department of Statistics Malaysia (2010) Profile of Orang Asli in Peninsular Malaysia, Kuala Lumpur. Available at <http://www.statistics.gov.my> (Accessed 23 January 2019).
- Dreyer G, Fernandes-Silva E, Alves S, Rocha A, Albuquerque R and Addiss D (1996) Patterns of detection of *Strongyloides stercoralis* in stool specimens: implications for diagnosis and clinical trials. *Journal of Clinical Microbiology* **34**, 2569–2571.
- Elyana FN, Al-Mekhlafi HM, Ithoi I, Abdulsalam AM, Dawaki S, Nasr NA, Atroosh WM, Abd-Basher MH, Al-Areeqi MA, Sady H, Subramaniam LR, Anuar TS, Lau YL, Moktar N and Surin J (2016) A tale of two communities: intestinal polyparasitism among Orang Asli and Malay communities in rural Terengganu, Malaysia. *Parasites and Vectors* **9**, 398.
- Forrer A, Khieu V, Schär F, Vounatsou P, Chammartin F, Marti H, Muth S and Odermatt P (2018) *Strongyloides stercoralis* and hookworm co-infection: spatial distribution and determinants in Preah Vihear Province, Cambodia. *Parasites and Vectors* **11**, 33.
- Fradejas I, Herrero-Martínez JM, Lizasoain M, Rodríguez de Las Parras E and Pérez-Ayala A (2018) Comparative study of two commercial tests for *Strongyloides stercoralis* serologic diagnosis. *Transactions of the Royal Society of Tropical Medicine and Hygiene* **112**, 561–567.
- Gétaz L, Castro R, Zamora P, Kramer M, Gareca N, Torrico-Espinoza MDC, Macias J, Lisarazu-Velásquez S, Rodríguez G, Valencia-Rivero C, Perneger T and Chappuis F (2019) Epidemiology of *Strongyloides stercoralis* infection in Bolivian patients at high risk of complications. *PLoS Neglected Tropical Diseases* **13**, e0007028.
- Grove DI (1996) Human strongyloidiasis. *Advanced Parasitology* **38**, 251–309.
- Gyorkos TW, Genta RM, Viens P and MacLean JD (1990) Seroepidemiology of *Strongyloides* infection in the Southeast Asian refugee population in Canada. *American Journal of Epidemiology* **132**, 257–264.

- Iatta R, Buonfrate D, Paradies P, Cavalera MA, Capogna A, Iarussi F, Šlapeta J, Giorli G, Trerotoli P, Bisoffi Z and Otranto D (2019) Occurrence, diagnosis and follow-up of canine strongyloidiasis in naturally infected shelter dogs. *Parasitology* **146**, 246–252.
- Jaleta TG, Zhou S, Bemm FM, Schär F, Khieu V, Muth S, Odermatt P, Lok JB and Streit A (2017) Different but overlapping populations of *Strongyloides stercoralis* in dogs and humans-dogs as a possible source for zoonotic strongyloidiasis. *PLoS Neglected Tropical Diseases* **11**, e0005752.
- Jourdan PM, Lambertson PHL, Fenwick A and Addiss DG (2018) Soil-transmitted helminth infections. *Lancet* **391**, 252–265.
- Junior KN, Zaman R, Zaman MH and Zaman T (2017) Twenty-five years of chronic strongyloidiasis in an immigrant. *Clinical Medicine Insights Case Reports* **10**, 1179547616684828.
- Kassalik M and Mönkemüller K (2011) *Strongyloides stercoralis* hyperinfection syndrome and disseminated disease. *Gastroenterology & Hepatology (NY)* **7**, 766–768.
- Kearns TM, Currie BJ, Cheng AC, McCarthy J, Carapetis JR, Holt DC, Page W, Shield J, Gundjirryirr R, Mulholland E, Ward L and Andrews RM (2017) *Strongyloides* seroprevalence before and after an ivermectin mass drug administration in a remote Australian Aboriginal community. *PLoS Neglected Tropical Diseases* **11**, e0005607.
- Khieu V, Srey S, Schär F, Muth S, Marti H and Odermatt P (2013) *Strongyloides stercoralis* is a cause of abdominal pain, diarrhea and urticaria in rural Cambodia. *BMC Research Notes* **6**, 200.
- Khieu V, Schär F, Marti H, Bless PJ, Char MC, Muth S and Odermatt P (2014) Prevalence and risk factors of *Strongyloides stercoralis* in Takeo Province, Cambodia. *Parasites and Vectors* **7**, 221.
- Khor GL and Zalilah MS (2008) The ecology of health and nutrition of 'Orang Asli' (Indigenous People) women and children in Peninsular Malaysia. *Tribes Tribals* **2**, 67–77.
- Kim JH, Kim DS, Yoon YK, Sohn JW and Kim MJ (2016) Donor-derived strongyloidiasis infection in solid organ transplant recipients: a review and pooled analysis. *Transplantation proceedings* **48**, 2442–2449.
- Knopp S, Salim N, Schindler T, Karagiannis Voules DA, Rothen J, Lweno O, Mohammed AS, Singo R, Benninghoff M, Nsojo AA, Genton B and Daubenberg C (2014) Diagnostic accuracy of Kato-Katz, FLOTAC, Baermann, and PCR methods for the detection of light-intensity hookworm and *Strongyloides stercoralis* infections in Tanzania. *American Journal of Tropical Medicine and Hygiene* **90**, 535–545.
- Koga K, Kasuya S, Khamboonruang C, Sukhavat K, Teda M, Takatsuka N, Kita K and Ohtomo H (1991) A modified agar plate method for detection of *Strongyloides stercoralis*. *American Journal of Tropical Medicine and Hygiene* **45**, 518–521.
- Laoraksawong P, Sanpool O, Rodpai R, Thanchomngang T, Kanarkard W, Maleewong W, Kraiklang R and Intapan PM (2018) Impact of the health education and preventive equipment package (HEPEP) on prevention of *Strongyloides stercoralis* infection among rural communities in Northeast Thailand: a cluster randomized controlled trial. *BMC Public Health* **18**, 1184.
- Lee SC, Ngui R, Tan TK, Roslan MA, Ithoi I and Lim YA (2014) Aquatic biomonitoring of *Giardia* cysts and *Cryptosporidium* oocysts in peninsular Malaysia. *Environmental Science and Pollution Research International* **21**, 445–453.
- Lim YA and Ahmad RA (2004) Occurrence of *Giardia* cysts and *Cryptosporidium* oocysts in the Temuan Orang Asli (aborigine) river system. *Southeast Asian Journal of Tropical Medicine and Public Health* **35**, 801–810.
- Lwanga SK and Lemeshow S (1991) *Sample Size Determination in Health Studies: A Practical Manual*. Geneva: World Health Organization.
- Masron T, Masami F and Ismail N (2013) Orang Asli in Peninsular Malaysia: population, spatial distribution and socio-economic condition. *Journal of Ritsumeikan Social Sciences and Humanities* **6**, 75–115.
- Moore SL and Wilson K (2002) Parasites as a viability cost of sexual selection in natural populations of mammals. *Science* **297**, 2015–2018.
- Muslim A, Mohd Sofian S, Shaari SA, Hoh B-P and Lim YA-L (2019) Prevalence, intensity and associated risk factors of soil-transmitted helminth infections: a comparison between negritos (indigenous) in inland jungle and those in resettlement at town peripheries. *PLoS Neglected Tropical Diseases* **13**, e0007331.
- Nasr NA, Al-Mekhlafi HM, Ahmed A, Roslan MA and Bulgiba A (2013) Towards an effective control program of soil-transmitted helminth infections among Orang Asli in rural Malaysia. Part 1: Prevalence and associated key factors. *Parasites and Vectors* **6**, 27.
- Ngui R, Halim NA, Rajoo Y, Lim YA, Ambu S, Rajoo K, Chang TS, Woon LC and Mahmud R (2016) Epidemiological characteristics of strongyloidiasis in inhabitants of indigenous communities in Borneo Island, Malaysia. *Korean Journal of Parasitology* **54**, 673–678.
- Nutman TB (2017) Human infection with *Strongyloides stercoralis* and other related *Strongyloides* species. *Parasitology* **144**, 263–273.
- Odani S (2017) Population dynamics among the Orang Asli of Peninsular Malaysia. *Senri Ethnological Studies* **95**, 151–170.
- Olsen A, van Lieshout L, Marti H, Polderman T, Polman K, Steinmann P, Stothard R, Thybo S, Verweij JJ and Magnussen P (2009) Strongyloidiasis—the most neglected of the neglected tropical diseases? *Transactions of the Royal Society of Tropical Medicine and Hygiene* **103**, 967–972.
- Paradies P, Iarussi F, Sasanelli M, Capogna A, Lia RP, Zucca D, Greco B, Cantacessi C and Otranto D (2017) Occurrence of strongyloidiasis in privately owned and sheltered dogs: clinical presentation and treatment outcome. *Parasites and Vectors* **10**, 345.
- Pocaterra LA, Ferrara G, Peñaranda R, Rojas E, Pérez-Chacón G, Hernán A, Certad G, Goldstein C and Núñez L (2017) Improved detection of *Strongyloides stercoralis* in modified agar plate cultures. *American Journal of Tropical Medicine and Hygiene* **96**, 863–865.
- Poulin R (1996) Sexual inequalities in helminth infections: a cost of being male? *The American Naturalist* **147**, 287–295.
- Prendki V, Fenaux P, Durand R, Thellier M and Bouchaud O (2011) Strongyloidiasis in man 75 years after initial exposure. *Emerging Infectious Diseases* **17**, 931–932.
- Puthiyakunnon S, Boddu S, Li Y, Zhou X, Wang C, Li J and Chen X (2014) Strongyloidiasis – an insight into its global prevalence and management. *PLoS Neglected Tropical Diseases* **8**, e3018.
- Rahmah N, Ariff RH, Abdullah B, Shariman MS, Nazli MZ and Rizal MZ (1997) Parasitic infections among aborigine children at Post Brooke, Kelantan, Malaysia. *Medical Journal of Malaysia* **52**, 412–415.
- Requena-Méndez A, Chiodini P, Bisoffi Z, Buonfrate D, Gotuzzo E and Muñoz J (2013) The laboratory diagnosis and follow up of strongyloidiasis: a systematic review. *PLoS Neglected Tropical Diseases* **7**, e2002.
- Requena-Mendez A, Buonfrate D, Bisoffi Z and Gutiérrez JM (2014) Advances in the diagnosis of human strongyloidiasis. *Current Tropical Medicine Reports* **1**, 207–215.
- Rockhill B, Newman B and Weinberg C (1998) Use and misuse of population attributable fractions. *American Journal of Public Health* **88**, 15–19.
- Schär F, Trostorf U, Giardina F, Khieu V, Muth S, Marti H, Vounatsou P and Odermatt P (2013) *Strongyloides stercoralis*: global distribution and risk factors. *PLoS Neglected Tropical Diseases* **7**, e2288.
- Schär F, Giardina F, Khieu V, Muth S, Vounatsou P, Marti H and Odermatt P (2016) Occurrence of and risk factors for *Strongyloides stercoralis* infection in South-East Asia. *Acta Tropica* **159**, 227–238.
- Senephansiri P, Laummaunwai P, Laymanivong S and Boonmar T (2017) Status and risk factors of *Strongyloides stercoralis* infection in rural communities of Xayaburi Province, Lao PDR. *Korean Journal of Parasitology* **55**, 569–573.
- Shimasaki T, Chung H and Shiiki S (2015) Five cases of recurrent meningitis associated with chronic strongyloidiasis. *American Journal of Tropical Medicine and Hygiene* **92**, 601–604.
- Steinmann P, Zhou XN, Du ZW, Jiang JY, Wang LB, Wang XZ, Li LH, Marti H and Utzinger J (2007) Occurrence of *Strongyloides stercoralis* in Yunnan Province, China, and comparison of diagnostic methods. *PLoS Neglected Tropical Diseases* **1**, e75.
- SyedHussain TPR, Krishnasamy DS and Hassan AAG (2017) Distribution and demography of the Orang Asli in Malaysia. *International Journal of Humanities and Social Science Invention* **6**, 40–45.
- Thamsborg SM, Ketzis J, Horii Y and Matthews JB (2017) Strongyloides spp. infections of veterinary importance. *Parasitology* **144**, 274–284.
- Tuyizere A, Ndayambaje A, Walker TD, Bayingana C, Ntiringanya C, Dusabemamba V and Hale DC (2018) Prevalence of *Strongyloides stercoralis* infection and other soil-transmitted helminths by cross-sectional survey in a rural community in Gisagara District, Southern Province, Rwanda. *Transactions of the Royal Society of Tropical Medicine and Hygiene* **112**, 97–102.
- Upanukraw P, Phongsri S and Morakote N (1999) Fluctuations of larval excretion in *Strongyloides stercoralis* infection. *American Journal of Tropical Medicine and Hygiene* **60**, 967–973.
- Verweij JJ, Canales M, Polman K, Ziem J, Brienen EA, Polderman AM and van Lieshout L (2009) Molecular diagnosis of *Strongyloides stercoralis* in

- faecal samples using real-time PCR. *Transactions of the Royal Society of Tropical Medicine and Hygiene* **103**, 342–346.
- Viney M (2017) Strongyloides. *Parasitology* **144**, 259–262.
- Vonghachack Y, Sayasone S, Bouakhasith D, Taisayavong K, Akkavong K and Odermatt P (2015) Epidemiology of *Strongyloides stercoralis* on Mekong islands in southern Laos. *Acta Tropica* **141**, 289–294.
- Winnicki W, Eder M, Mazal P, Mayer FJ, Sengölge G and Wagner L (2018) Prevalence of *Strongyloides stercoralis* infection and hyperinfection syndrome among renal allograft recipients in central Europe. *Scientific Reports* **8**, 15406.
- Wiria AE, Wammes LJ, Hamid F, Dekkers OM, Prasetyani MA, May L, Kaisar MM, Verweij JJ, Tamsma JT, Partono F, Sartono E, Supali T, Yazdanbakhsh M and Smit JW (2013) Relationship between carotid intima media thickness and helminth infections on Flores Island, Indonesia. *PLoS ONE* **8**, e54855.
- Wong CL, Venneker R, Uhlenbrook S, Jamil ABM and Zhou Y (2009) Variability of rainfall in Peninsular Malaysia. *Hydrology and Earth System Sciences* **6**, 5471–5503.
- World Health Organization and United Nations Children's Fund (2015) Progress on sanitation and drinking-water: 2015 update. Available at http://files.unicef.org/publications/files/Progress_on_Sanitation_and_Drinking_Water_2015_Update_.pdf (Accessed 30 April 2019).
- Zeehaida M, Zairi NZ, Rahmah N, Maimunah A and Madihah B (2011) *Strongyloides stercoralis* in common vegetables and herbs in Kota Bharu, Kelantan, Malaysia. *Tropical Biomedicine* **28**, 188–193.