

Lucy E. Wilson, MD;<sup>7</sup>  
 John J. LiPuma, MD;<sup>9</sup>  
 Aaron M. Milstone, MD, MHS<sup>1,2</sup>

## A Colonization Outbreak of Penicillin-Susceptible *mecA*-Positive *Staphylococcus aureus* in a Neonatal Ward of Children's Hospital

Affiliations: 1. Department of Pediatrics, Division of Pediatric Infectious Diseases, Johns Hopkins University School of Medicine, Baltimore, Maryland; 2. Department of Hospital Epidemiology and Infection Control, The Johns Hopkins Hospital, Baltimore, Maryland; 3. Department of Anesthesia, Division of Pediatric Intensive Care Unit, Johns Hopkins University School of Medicine, Baltimore, Maryland; 4. Department of Pediatric Nursing, The Johns Hopkins Hospital, Baltimore, Maryland; 5. Department of Pathology, Division of Medical Microbiology, Johns Hopkins University School of Medicine, Baltimore, Maryland; 6. Division of Pediatric Infectious Disease, Stanford University School of Medicine, Stanford, California; 7. Infectious Disease Epidemiology and Outbreak Response Bureau, Maryland Department of Health, Baltimore, Maryland; 8. Division of Healthcare Quality Promotion, Centers for Disease Control and Prevention, Atlanta, Georgia; 9. Department of Pediatrics and Communicable Disease, University of Michigan Medical School, Ann Arbor, Michigan.

Address correspondence to Ibukunoluwa Akinboyo, 200 North Wolfe Street, Room 3155, Baltimore, MD 21287 (ibukun@jhmi.edu).

<sup>a</sup>Contributed equally as first authors.

*Infect Control Hosp Epidemiol* 2018;39:237–239

© 2018 by The Society for Healthcare Epidemiology of America. All rights reserved. 0899-823X/2018/3902-0020. DOI: 10.1017/ice.2017.265

### REFERENCES

1. Marquez L, Jones KN, Whaley EM, et al. An outbreak of *Burkholderia cepacia* complex infections associated with contaminated liquid docusate. *Infect Control Hosp Epidemiol* 2017;38:567–573.
2. US Food and Drug Administration (FDA). FDA updates on multistate outbreak of *Burkholderia cepacia* infections. FDA website. <https://www.fda.gov/Drugs/DrugSafety/ucm511527.htm>. Published 2016. Accessed November 5, 2017.
3. Office of Regulatory Affairs, US Food and Drug Administration (FDA). Safety alerts—rugby laboratories issues voluntary nationwide recall of diocto liquid and diocto syrup manufactured by PharmaTech, LLC, due to possible product contamination. FDA website. <https://www.fda.gov/safety/recalls/ucm569967.htm>. Published 2017. Accessed November 5, 2017.
4. Office of the Commissioner, US Food and Drug Administration (FDA). Safety alerts for human medical products: diocto liquid and diocto syrup by rugby laboratories: recall—possible product contamination. FDA website. <https://www.fda.gov/Safety/MedWatch/SafetyInformation/SafetyAlertsforHumanMedicalProducts/ucm570014.htm>. Published 2017. Accessed November 5, 2017.
5. Office of Regulatory Affairs, US Food and Drug Administration (FDA). Safety alerts: voluntary nationwide recall of all liquid products manufactured by Pharmatech LLC and distributed by Leader Brand, Major Pharmaceuticals, and Rugby laboratories due to possible product contamination. FDA website. <https://www.fda.gov/Safety/Recalls/ucm571001.htm>. Published 2017. Accessed November 5, 2017.
6. Daugherty Biddison L, Berkowitz KA, Courtney B, et al. Care of the critically ill and injured during pandemics and disasters: CHEST consensus statement. *Chest* 2014;146:e145S–e155S.
7. Siddiqui AH, Mulligan ME, Mahenthalingam E, et al. An episodic outbreak of genetically related *Burkholderia cepacia* among non-cystic fibrosis patients at a university hospital. *Infect Control Hosp Epidemiol* 2001;22:419–422.

*To the Editor*—We experienced a colonization outbreak of penicillin-susceptible and *mecA*-positive *Staphylococcus aureus* strain in neonatal ward. After implementation of strict precautions and decolonization, the outbreak was terminated. To our knowledge, this is the first report of penicillin-susceptible MRSA outbreak in a neonatal ward.

Nagano Children's Hospital is a tertiary pediatric hospital located in a rural area of Japan with 42 beds in the neonatal ward. Active weekly surveillance cultures of nares of inpatients of the neonatal ward have been carried out since the 1990s, especially for monitoring methicillin-resistant *Staphylococcus aureus* (MRSA). In recent years, the proportion of MRSA carriage in the neonatal ward has been approximately 0 to 5%.

In late July 2016, our surveillance system noticed an unusual surge in the colonization rate of methicillin-susceptible *S. aureus* (MSSA) in the neonatal ward. Detailed investigations revealed that this *S. aureus* strain has unique characteristics; namely, it is uniformly susceptible to penicillin but hetero-resistant to oxacillin and ceftioxin. Genetic analysis also revealed that this strain possesses the *mecA* gene; therefore, this strain was reassessed as MRSA, according to Clinical and Laboratory Standards Institute (CLSI) criteria.<sup>1</sup>

The infection control team had emergency meetings and alerted healthcare workers throughout the hospital about the outbreak. The campaign for reinforcement of hand hygiene with contact precautions, strict isolation, and cohorting the patients was carried out. However, by the end of August, the colonization rate reached its the highest level (12 of 43 patients, 28%). We then decided to implement MRSA decolonization with mupirocin ointment. In total, 17 patients (12 penicillin-susceptible [PS] MRSA patients and 5 'ordinal' MRSA carrier patients) had undergone the decolonization; 10 of 13 patients (76.9%) were confirmed as decolonized (defined as negative results for 2 consecutive cultures). Furthermore, 3 patients were not decolonized, and the other 4 patients were discharged before follow-up cultures were performed.

After these interventions, the carriage rate of PS-MRSA decreased, and no new cases of colonization were reported for 2 consecutive weeks. In late October, we declared that the outbreak had ended. Fortunately, there were no serious infections due to this PS-MRSA during this outbreak.

The outbreak strain of PS-MRSA did not yield typical cultures on MRSA-specific chromogenic media (CHROMagar II, Becton-Dickinson, Japan); on this selective medium, it yielded only a few slow-growing colonies, and sometimes the strain did not yield a culture on the medium. Antimicrobial susceptibility test showed resistance to gentamicin, erythromycin, and levofloxacin but susceptibility to vancomycin. The minimum

TABLE 1. Molecular Characteristics of the Outbreak Strain

ST	ACME	PVL Genes	Enterotoxins	Adhesins
5	...	...	<i>sed, seg, sei, sem, sen, seo, sep, selx, selw</i>	<i>icaD, eno, fnbA, ebpS, fib, clfA, clfB, sdrC, sdrD, sdrE, bbp, sak, chp</i>

NOTE: ST, sequence type; ACME, arginine catabolic mobile element; PVL, Panton-Valentine leucocidin.

inhibitory concentration (MIC) to oxacillin ranged from 0.25 to 4.0 µg/mL, and the MIC to cefoxitin ranged from 4.0 to 8.0 µg/mL, and the strain was assessed as susceptible to resistant. However, the MIC to penicillin was uniformly low, 0.03–0.12 µg/mL, and the strain was judged as susceptible according to CLSI criteria.<sup>1</sup> These MICs were measured using a broth microdilution test. The MICs to antimicrobial agents were also measured by E-test, and the results showed a similar tendency. The MICs to oxacillin, cefoxitin, and penicillin ranged from 0.25 to 8.0 µg/mL, from 4.0 to 24.0 µg/mL, and from 0.094 to 1.0 µg/mL, respectively.

Molecular subtyping and gene analyses, which were performed as described previously,<sup>2,3</sup> revealed that all the strains belonged to *coa* type IIa, SCC*mec* type I (type-1 *ccr* (A1, B1) and class B *mec*), sequence type (ST) 5, *spa* type t010, and *agr* group II. They did not possess Panton-Valentine leukocidin (PVL) or arginine catabolic mobile element (ACME) genes, but they harbored several hemolysins, enterotoxin gene clusters, and adhesins as shown in Table 1. On the other hand, these strains lacked *mecI*, *mecR1*, and *blaZ* genes.

Genetic analyses of these strains revealed that mobile gene element IS *1182* was inserted within the promoter region of *mecA* gene in the class B SCC*mec*. Therefore, the *mecA* system of this strain was suggested to be nonfunctional. The sequence data were deposited in GenBank under accession nos. MF278653 and MF278654.

To our knowledge, this MRSA outbreak strain, ST5-SCC*mec* type I, has not been reported from clinical isolates in our country. SCC*mec* type I MRSA has seldom been isolated from clinical specimens since the 1990s, and its prevalence might be ~1%–5%.<sup>4</sup>

The emergence of this type of PS-MRSA poses several clinical problems. First, PS-MRSA could not be detected using a routine MRSA selection medium, so PS-MRSA might often be misrecognized as MSSA. Therefore, genetic analysis, such as polymerase chain reaction (PCR), is necessary for the detection of the *mecA* gene from *S. aureus* isolates, at least in serious infections. Second, an appropriate antimicrobial agent for PS-MRSA remains unknown. Moreover, there might be threat of converting from PS-MRSA to true (penicillin- and oxacillin-resistant) MRSA during treatment. The usual treatment regimen for MSSA infection with β-lactam antimicrobials might lead to treatment failure.<sup>5</sup>

Exact prevalence of penicillin-susceptible or oxacillin-susceptible (PS/OS-) MRSA among clinically isolated MSSA is unknown, but it is supposed to be ~3%.<sup>5</sup> Literature on PS/OS-MRSA has been increasing all over the world.<sup>6,7</sup>

The mechanisms of anomalous antimicrobial susceptibility of PS/OS-MRSA have not been fully elucidated.<sup>8</sup> Several hypotheses have been proposed, such as amino acid changes in *Fem* proteins, which are responsible for Staphylococcal cell-wall synthesis,<sup>6</sup> partial excision of *mecA* gene,<sup>9</sup> and *bla* system dysfunction.<sup>10</sup> PS/OS-MRSA strains are also quite diverse; therefore, many other novel mechanisms might be revealed. In consideration of its clinical importance, more attention should be given to penicillin- or oxacillin-susceptible, *mecA*-positive *S. aureus*.

#### ACKNOWLEDGMENTS

*Financial support:* No financial support was provided relevant to this article.

*Potential conflicts of interest:* All authors report no conflicts of interest relevant to this article.

Kisei Minami, MD;<sup>1</sup>  
Runa Terakawa, MD;<sup>1</sup>  
Masanori Sato, MD;<sup>1</sup>  
Yasuhiro Shoji, MD;<sup>1</sup>  
Takehiko Hiroma, PhD;<sup>1</sup>  
Tomohiko Nakamura, PhD;<sup>1</sup>  
Ayaka Horiuchi, MSc;<sup>1</sup>  
Ayaka Otsuka, BSc;<sup>1</sup>  
Noriko Kubota, PhD;<sup>1</sup>  
Eiko Hidaka, PhD;<sup>1</sup>  
Yoshiyuki Kawakami, PhD;<sup>2</sup>  
Meiji Soe Aung, PhD;<sup>3</sup>  
Nobumichi Kobayashi, PhD<sup>3</sup>

Affiliations: 1. Nagano Children's Hospital, Azumino, Japan; 2. Shinshu University Graduate School of Medicine, Matsumoto, Japan; 3. Sapporo Medical University School of Medicine, Sapporo, Japan.

Address correspondence to Kisei Minami, Toyoshina 3100, Azumino, Nagano prefecture, Japan (kiseiminami@gmail.com).

PREVIOUS PRESENTATION. A part of this manuscript was presented at the 35th Annual Meeting of the European Society for Paediatric Infectious Diseases, ESPID 2017, in Madrid, Spain, on May 25–26, 2017.

*Infect Control Hosp Epidemiol* 2018;39:239–241

© 2018 by The Society for Healthcare Epidemiology of America. All rights reserved. 0899-823X/2018/3902-0021. DOI: 10.1017/ice.2017.266

#### REFERENCES

- Clinical and Laboratory Standards Institute. Performance standards for antimicrobial susceptibility testing: 25th informational supplement. CLSI document M100-S25. Wayne, PA: Clinical and Laboratory Standards Institute; 2015.
- Kawaguchiya M, Urushibara N, Ghosh S, et al. Genetic diversity of emerging Panton-Valentine leukocidine/arginine catabolic mobile element (ACME)-positive ST8 SCC*mec*-IVa methicillin resistant *Staphylococcus aureus* (MRSA) strains and ACME-positive CC5 (ST5/ST764) MRSA strains in northern Japan. *J Med Microbiol* 2013;62:1852–1863.

3. Aung MS, Urushibara N, Kawaguchiya M, et al. Virulence factors and genetic characteristics of methicillin-resistant and -susceptible *Staphylococcus aureus* isolates in Myanmar. *Microb Drug Resist* 2011;17:525–535.
4. Inomata S, Yano H, Tokuda K, et al. Microbiological and molecular epidemiological analyses of community-associated methicillin-resistant *Staphylococcus aureus* at a tertiary care hospital in Japan. *J Infect Chemother* 2015;21:729–736.
5. Proulx MK, Palace SG, Gandra S, et al. Reversion from methicillin susceptibility to methicillin resistance in *Staphylococcus aureus* during treatment of bacteremia. *J Infect Dis* 2016;213:1041–1048.
6. Pournaras S, Stathopoulos C, Tsakris A. Oxacillin-susceptible MRSA: could it become a successful MRSA type? *Future Microbiol* 2013;8:1365–1367.
7. Hososaka Y, Hanaki H, Endo H, et al. Characterization of oxacillin-susceptible mecA-positive *Staphylococcus aureus*: a new type of MRSA. *J Infect Chemother* 2007;13:79–86.
8. Lowy FD. Antimicrobial resistance: the example of *Staphylococcus aureus*. *J Clin Invest* 2003;111:1265–1273.
9. Shore AC, Rossney AS, O'Connell B, et al. Detection of Staphylococcal cassette chromosome mec-associated DNA segments in multiresistant methicillin-susceptible *Staphylococcus aureus* (MSSA) and identification of *Staphylococcus epidermidis* ccrAB4 in both methicillin-resistant *S. aureus* and MSSA. *Antimicrob Agents Chemother* 2008;52:4407–4419.
10. Sabat AJ, Pournaras S, Akkerboom V, et al. Whole-genome analysis of an oxacillin-susceptible CC80 mecA-positive *Staphylococcus aureus* clinical isolate: insights into the mechanisms of cryptic methicillin resistance. *J Antimicrob Chemother* 2015;70:2956–2964.

## Predicting Multidrug-Resistant Gram-Negative Bacterial Colonization and Associated Infection on Hospital Admission: Methodological Issues

*To the Editor*—We read with great interest the article titled “Predicting Multidrug-Resistant Gram-Negative Bacterial Colonization and Associated Infection on Hospital Admission” by Tseng et al<sup>1</sup> published in a recent issue of this journal. We would like to congratulate the authors on their valuable work; however, we think some methodological and statistical issues should be considered to avoid misinterpretation.

As shown in the Table 3 of the article, when a predictor meets a univariate criterion of  $P < .01$ , the predictor is further considered for multivariable analysis. Here, we are concerned that the authors considered a very conservative  $P$  value for univariate screening of candidate predictors. They argued that when a conservative  $P$  value (eg,  $< .01$  or  $< .05$ ) is selected in univariate analysis, only the predictors with relatively large

effect will be included in the multivariable analysis. In such a situation, the estimated regression coefficients of selected predictors can have bias away from the null,<sup>2,3</sup> which is known as testimation bias.

Considering a liberal  $P$  value (eg,  $< .10$  or  $< .20$ ) in univariable analysis can effectively compensate for testimation bias.<sup>2</sup> In other words, we can be sure that predictors with relatively large effect (eg,  $P < .01$ ) and predictors with relatively small effect (eg,  $.10 < P < .20$ ) can be tested in multivariable analysis after univariate screening with, for example,  $P < .20$ . In the study,<sup>1</sup> although long-term hemodialysis appear to be an uninteresting predictor for risk of multidrug-resistant gram-negative bacteria (MDR-GNB) colonization in univariable analysis, it may have a significant effect but only in the presence of other predictors.

We acknowledge that the study provides very interesting results, but the estimated associations for predictors of MDR-GNB colonization may be different from those reported in the study due to testimation bias.

### ACKNOWLEDGMENTS

*Financial support:* No financial support was provided relevant to this article.

*Potential conflicts of interest:* All authors report no conflicts of interest relevant to this article.

**Kamyar Mansori,<sup>1,2</sup>**  
**Erfan Ayubi,<sup>3,4</sup>**  
**Saeid Safiri<sup>5</sup>**

Affiliations: 1. Social Determinants of Health Research Center, Kurdistan University of Medical Sciences, Sanandaj, Iran; 2. Department of Epidemiology, School of Public Health, Iran University of Medical Sciences, Tehran, Iran; 3. Department of Epidemiology, School of Public Health, Shahid Beheshti University of Medical Sciences, Tehran, Iran; 4. Department of Epidemiology and Biostatistics, School of Public Health, Tehran University of Medical Sciences, Tehran, Iran; 5. Managerial Epidemiology Research Center, Department of Public Health, School of Nursing and Midwifery, Maragheh University of Medical Sciences, Maragheh, Iran.

Address correspondence to Saeid Safiri, Assistant Professor of Epidemiology, Managerial Epidemiology Research Center, Department of Public Health, School of Nursing and Midwifery, Maragheh University of Medical Sciences, Maragheh, Iran (saeidsafiri@gmail.com).

*Infect Control Hosp Epidemiol* 2018;39:241–241

© 2018 by The Society for Healthcare Epidemiology of America. All rights reserved. 0899-823X/2018/3902-0022. DOI: 10.1017/ice.2017.270

### REFERENCES

1. Tseng WP, Chen YC, Yang BJ, et al. Predicting multidrug-resistant gram-negative bacterial colonization and associated infection on hospital admission. *Infect Control Hosp Epidemiol* 2017;38:1216–1225.
2. Steyerberg E. *Clinical Prediction Models: A Practical Approach to Development, Validation, and Updating*. New York: Springer Science; 2008.
3. Pakzad R, Safiri S. Incidence and risk factors for surgical site infection posthysterectomy in a tertiary care center: methodologic issues. *Am J Infect Control* 2017;45:580–581.