

Review: Following the smoke signals: inflammatory signaling in metabolic homeostasis and homeorhesis in dairy cattle

B. J. Bradford[†]  and T. H. Swartz

Department of Animal Sciences and Industry, Kansas State University, 1530 Mid-Campus Dr. N., Manhattan, KS 66506, USA

(Received 10 April 2019; Accepted 29 July 2019)

Inflammatory cascades are a critical component of the immune response to infection or tissue damage, involving an array of signals, including water-soluble metabolites, lipid mediators and several classes of proteins. Early investigation of these signaling pathways focused largely on immune cells and acute disease models. However, more recent findings have highlighted critical roles of both immune cells and inflammatory mediators on tissue remodeling and metabolic homeostasis in healthy animals. In dairy cattle, inflammatory signals in various tissues and in circulation change rapidly and dramatically, starting just prior to and at the onset of lactation. Furthermore, several observations in healthy cows point to homeostatic control of inflammatory tone, which we define as a regulatory process to balance immune tolerance with activation to keep downstream effects under control. Recent evidence suggests that peripartum inflammatory changes influence whole-body nutrient flux of dairy cows over the course of days and months. Inflammatory mediators can suppress appetite, even at levels that do not induce acute responses (e.g. fever), thereby decreasing nutrient availability. On the other hand, inhibition of inflammatory signaling with non-steroidal anti-inflammatory drug (NSAID) treatment suppresses hepatic gluconeogenesis, leading to hypoglycemia in some cases. Over the long term, though, peripartum NSAID treatment substantially increases peak and whole-lactation milk synthesis by multiparous cows. Inflammatory regulation of nutrient flux may provide a homeorhetic mechanism to aid cows in adapting to rapid changes in metabolic demand at the onset of lactation, but excessive systemic inflammation has negative effects on metabolic homeostasis through inhibition of appetite and promotion of immune cell activity. Thus, in this review, we provide perspectives on the overlapping regulation of immune responses and metabolism by inflammatory mediators, which may provide a mechanistic underpinning for links between infectious and metabolic diseases in transition dairy cows. Moreover, we point to novel approaches to the management of this challenging phase of the production cycle.

Keywords: parturition, metabolism, resolution, non-steroidal anti-inflammatory drug, cytokine

Implications

Inflammatory signals are elevated in dairy cows in the days around parturition, but these signals are not always pathological. Physiological roles of inflammatory mediators may include directing metabolic shifts that enable adaptation to lactation, although excessive elevation of these signals likely has net negative impacts on nutrient availability through suppression of appetite and activation of immune responses. The management of inflammatory tone through dietary or pharmacological means may reduce periparturient disorders while enhancing productivity in dairy cattle.

Introduction

In dairy cattle, the transition to lactation represents a perilous time period when the incidence of disease is high. This increase in disease incidence is associated with a chronic inflammatory response (Sordillo and Raphael, 2013). At the onset of lactation, a depression of feed intake occurs simultaneously with an increase in energy demand, resulting in metabolic stress. Low-grade inflammation has been associated with metabolic and infectious diseases in early-lactation cattle (Bradford *et al.*, 2015), and therefore, the metabolic effects of inflammation have been the focus of much research in recent years. An accelerating field of research has now demonstrated that immune cells are directly involved in a surprising array of metabolic functions (Man *et al.*, 2017), including the maintenance of gastrointestinal

[†] E-mail: bjbrad@ksu.edu

function (Ferreira *et al.*, 1990; Hoytema van Konijnenburg *et al.*, 2017), control of adipose tissue lipolysis (Kosteli *et al.*, 2010) and regulation of insulin sensitivity in multiple tissues (Olefsky and Glass, 2010). In fact, the immune system is so deeply embedded in metabolic physiology that some suggest that the liver, adipose tissue and the immune system developed from a single ancestral organ (Hotamisligil, 2006). In recent years, the vast majority of research on low-grade inflammation associated with metabolic disorders, also termed metabolic inflammation, has focused on human diseases that are, in part, initiated by inflammatory signals, including diabetes, hepatic steatosis and atherosclerosis (Medzhitov, 2008). Recent advancements in bovine research have also illuminated the intersection of immune function and metabolism. Yet, many questions remain, including:

- Are there adaptive roles for metabolic inflammation during the transition to lactation?
- Is metabolic inflammation homeostatically regulated, and if so, what are the mechanisms behind this regulation?
- If these mechanisms are manipulated, what are the potential benefits and consequences?
- How can this information be harnessed on-farm to advance dairy cattle health and productivity?

Several recent reviews have provided updates on the role of immune cells in gastrointestinal tissue of cattle and downstream impacts of their activation, including on hepatic function (Steele *et al.*, 2016; Garcia *et al.*, 2017; Khiaosa-Ard and Zebeli, 2018). Therefore, this review presents some targeted perspectives, rather than an exhaustive overview, on adipose tissue and inter-organ integration of inflammatory signals. We focus on the concept of inflammatory tone, which we define as the regulatory process to strike a balance between immune activation and tolerance.

Immune cells perform an intricate inflammatory balancing act in adipose tissue

The mechanisms underlying sentinel functions of the immune system have become far better described in the past decade, particularly with respect to the need to strike an appropriate balance between immune tolerance and responsiveness to microbial or damage-associated signals. In the gut especially, this balance is critical to health, even in the absence of a true pathogenic challenge (Chistiakov *et al.*, 2015). One of the primary ways that this balance is achieved is through immune cross-talk in the gut, primarily utilizing pro- and anti-inflammatory cytokines to regulate proliferation and activity of specific immune cells (Omenetti and Pizarro, 2015). This regulatory process, which we refer to as inflammatory tone, is clearly critical to gut homeostasis. An important question is whether inflammatory tone is similarly important for organs that are not protecting an epithelial surface. Although the need for tolerance is perhaps less obvious

in tissues that are not routinely exposed to commensal microbiota (Desruisseaux *et al.*, 2007), there is strong evidence that maintaining a balance between inflammatory and regulatory immune cells is critical for the metabolic health of adipose tissue and, in turn, the animal.

Immune cells resident in many tissues regulate nutrient use on a routine basis (Iyer *et al.*, 2015) in addition to the broad systemic effects of cytokine storms that occur during disease. The mechanisms behind metabolic inflammation remain speculative in bovines; however, in rodents, the nexus of immune cells and metabolism are far better described (Figure 1). Tissue-resident macrophages play a central role in metabolic stress; and in obesity the proportion of macrophages in adipose tissue increases from approximately 10% of all cells to 50% (Weisberg *et al.*, 2003, Lumeng *et al.*, 2007). These cells can be classified as either pro-inflammatory (M1) or anti-inflammatory (M2), although these classifications have limitations as macrophages exhibit immense plasticity in adipose tissue (Morris *et al.*, 2011). In obese mice, adipocyte hypertrophy triggers the invasion of M1 macrophages into adipose tissue. Some of these adipocytes will atrophy, resulting in the release of fatty acids which are engulfed by macrophages to prevent lipotoxicity. The net effect is a low-grade inflammatory tone due to the accumulation of M1 macrophages in crown-like structures, which also include necrotic adipocytes and lipid remnants (Bai and Sun, 2015). However, in bovine, an overt inflammatory state was not found in subcutaneous adipose tissue of overfed non-pregnant non-lactating cows (Lopreato *et al.*, 2018), although a mildly inflammatory state was noted in mesenteric adipose tissue (Moisá *et al.*, 2017). Moreover, contrary to observations from rodents, macrophages do not accumulate in crown-like structures in early-lactation dairy cows (Akter *et al.*, 2012; Contreras *et al.*, 2015). Yet, a greater number of adipose tissue macrophages in diseased early-lactation dairy cattle (particularly M1 macrophages) was found compared to non-lactating dairy cows in addition to an increase in pro-inflammatory cytokine mRNA abundance, although it should be noted that there was also a smaller increase in the number of M2 macrophages (Contreras *et al.*, 2015). When comparing late-lactation cows in either positive or negative energy balance, an increase in macrophage infiltration in adipose was noted in cows experiencing negative energy balance; however, there were minimal changes in inflammatory markers between groups (Contreras *et al.*, 2016). Lastly, an invasion of macrophages in adipose was associated with a greater degree of body condition loss in transition dairy cows (Newman *et al.*, 2019). Collectively, these data suggest a role for M1 macrophages in the development of a low-grade chronic inflammatory state that can occur in diseased early-lactation dairy cattle (Contreras *et al.*, 2018). In agreement with these findings, an increased mRNA abundance of the M1 cytokine, tumor necrosis factor- α (TNF- α), which is primarily produced by M1 macrophages, was found in adipose tissue from obese rats (Hotamisligil *et al.*, 1993) and postpartum dairy cattle (Sadri *et al.*, 2010).

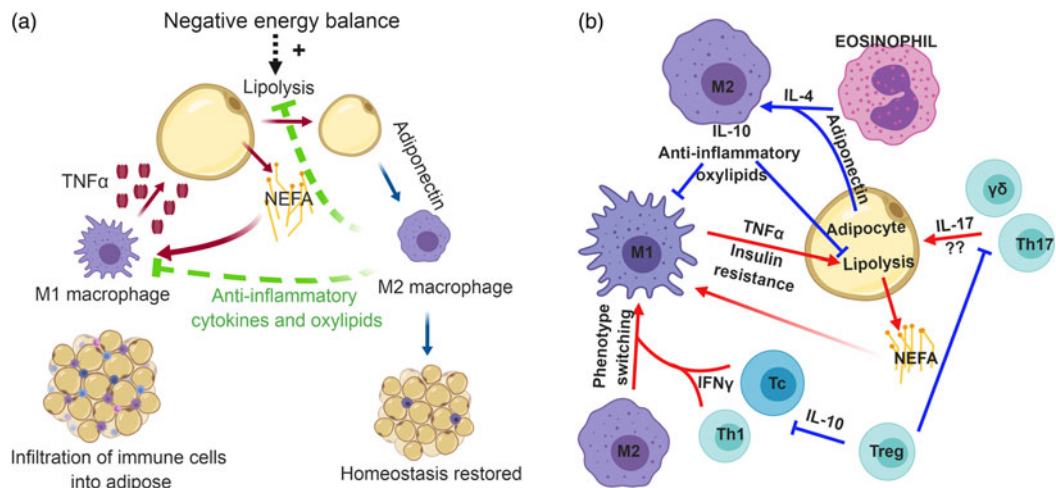


Figure 1 (Color online) Role of immune cells in mammalian adipose tissue. (a) Adipose tissue-resident macrophages exhibit a pro-inflammatory (M1) phenotype that is stimulated by the release of non-esterified fatty acids (NEFA) from adipocytes. In response to NEFA, M1 macrophages secrete the pro-inflammatory cytokine, tumor necrosis factor- α (TNF- α), which can further induce lipolysis. To maintain inflammatory tone, anti-inflammatory (M2) macrophages may be recruited into adipose tissue during negative energy balance. Adiponectin polarizes recruited macrophages toward an M2 phenotype, which produce anti-inflammatory cytokines and oxylipids to temper lipolysis and inflammation and restore homeostasis. (b) Immense cross-talk exists between a variety of immune cells to regulate inflammatory tone in adipose tissue. In obesity, cytotoxic T cells (Tc) and T helper 1 (Th1) cells in adipose tissue secrete interferon- γ (IFN- γ). This cytokine promotes phenotype switching of macrophages from an M2 phenotype to M1. Pro-inflammatory macrophages produce TNF- α , which induces insulin resistance. On the other hand, eosinophils produce interleukin-4 (IL-4) which synergizes with the anti-inflammatory adipose-derived cytokine, adiponectin, to promote M2 polarization of macrophages. This anti-inflammatory macrophage is thought to temper inflammatory responses in adipose tissue and slow lipolysis. Lastly, the role of IL-17-producing $\gamma\delta$ T cells and Th17 cells are relatively unknown, but the production of IL-17 likely contributes to the chronic, low-grade inflammatory tone seen in metabolic inflammation. Regulatory T cells (Treg) can inhibit the polarization of pro-inflammatory immune cells, thereby promoting tissue homeostasis and self-tolerance. Panel B inspired by Chatzigeorgiou *et al.* (2012). Most studies underlying these relationships have been conducted with mice, but mechanisms are proposed to be conserved across mammalian species.

Monocytes are recruited to adipose tissue via chemo-attractants, such as CC-chemokine ligand 2 (CCL2) and CCL5 (Mukesh *et al.*, 2010), which are produced by adipocytes and M1 macrophages, and additional recruitment and invasion of M1 macrophages should aid in amplifying the inflammatory response (Chawla *et al.*, 2011; Odegaard and Chawla, 2013). Considering these findings, one could expect a cascade of immune activation events snowballing into a pronounced inflammatory response. Yet, metabolic inflammation appears to have 'one foot on the gas' and simultaneously 'one foot on the brake', resulting in a smoldering, low-grade inflammatory tone. Thus, the question of how this inflammatory tone is maintained remains elusive to many researchers. It seems plausible that other immune cells are playing a critical role in maintaining a low-grade inflammatory tone in diseased cattle, preventing an aggressive immune response, and that these cells could underpin an anti-inflammatory adaptation that may occur in healthy early-lactation dairy cattle.

Eosinophils are granulocytes that are typically associated with allergy and asthma. In adipose tissue, eosinophils produce the anti-inflammatory cytokine, interleukin-4 (IL-4), that promotes differentiation of monocytes toward the IL-10 producing, M2 macrophage phenotype in mice (Wu *et al.*, 2011). Interleukin-4 synergizes with adiponectin, an adipocyte-derived anti-inflammatory cytokine (Kabara *et al.*, 2014), to improve insulin sensitivity (Chawla *et al.*, 2011), and IL-10 protects against TNF- α -induced insulin resistance (Lumeng *et al.*, 2007). As previously mentioned, CCL2 recruits

monocytes, and knocking out the CCL2 receptor in obese mice increased the number of eosinophils in adipose tissue (Bolus *et al.*, 2015). Taken together, these data suggest that during metabolic inflammation, CCL2 may play a critical role in tilting the balance toward a pro-inflammatory tone by not only recruiting M1 macrophages but also reducing the migration of anti-inflammatory eosinophils into adipose tissue. In dairy cattle, the role of eosinophils in adipose tissue has yet to be explored, but blood eosinophils are reduced in early-lactation cows (Holtenius *et al.*, 2004), suggesting a need for additional research.

T lymphocytes are also key players in regulating inflammatory tone in adipose tissue. T cells can be classified into different subsets such as regulatory T cells (Treg), helper T cells (Th), cytotoxic T cells (Tc) and $\gamma\delta$ T cells. Depending on the stimuli, T cells can be polarized into different phenotypes that are defined by the cytokines produced, and these cytokines exert either pro- or anti-inflammatory effects. For instance, pro-inflammatory interferon- γ (IFN- γ) drives immune responses that are particularly effective against intracellular microbes (type 1 response, Th1), whereas cytokines such as IL-4, IL-5 and IL-13 support antibody production from B cells, M2 polarization of macrophages and elimination of parasitic infections (type 2 response, Th2). Moreover, pro-inflammatory IL-17 induces neutrophil generation and migration (Th17), while IL-10 promotes tissue homeostasis and self-tolerance by suppressing the immune function (Treg). These cells serve to counterbalance each other. For instance, a pro-inflammatory Th1 cytokine is

counterbalanced with the anti-inflammatory Th2 cytokine; and similarly, Th17 offsets Treg cells. Thus, the balance of these cells is critical for determining inflammatory tone.

In lean mice, immunosuppressive Treg cells, which produce IL-10, were highly enriched in adipose tissue, but not so in obese mice (Feuerer *et al.*, 2009). Indeed, obesity is characterized by the production of pro-inflammatory cytokines from the major T cell subsets, including cytotoxic T cells, T helper cells and $\gamma\delta$ T cells (Feuerer *et al.*, 2009; Winer *et al.*, 2009; Mehta *et al.*, 2015). In mice fed a high-fat diet, cytotoxic T cells promoted monocyte infiltration and inflammation in adipose tissue, and depletion of these T cells ameliorated insulin resistance (Nishimura *et al.*, 2009). More specifically, IFN- γ produced by Th1 and cytotoxic T cells promotes M1 macrophage differentiation, and more recent evidence has shown that this cytokine can also induce insulin resistance directly (Rocha *et al.*, 2008; McGillicuddy *et al.*, 2009; O'Rourke *et al.*, 2012; Khan *et al.*, 2014). While it is unclear what role IL-17 plays in adipose tissue (Ahmed and Gaffen, 2010), it should be noted that obesity is associated with an increase in IL-17 production (Winer *et al.*, 2009). Th17 cells exacerbate hepatic steatosis and liver inflammation (Tang *et al.*, 2011), and $\gamma\delta$ T cells are increased in adipose tissue in obese mice (Mehta *et al.*, 2015); both of these cell types are major sources of IL-17. Moreover, when stimulated peripheral blood mononuclear cells are treated with pro-inflammatory palmitate, there is a dramatic increase in IL-17 production (McCambridge *et al.*, 2019). On the anti-inflammatory side, the abundance of Th2 cells in adipose tissue was positively associated with insulin sensitivity in humans (McLaughlin *et al.*, 2014). Moreover, Th2-associated cytokines are needed for infiltration of eosinophils in adipose tissue, which in turn promotes macrophage phenotype switching from M1 to M2 (Molofsky *et al.*, 2013). Piecing it altogether, a tilting of the scale toward Th1 and Th17 and away from Th2 and Treg responses could be another underlying mechanism behind the low-grade chronic inflammatory state found in metabolic inflammation (Figure 1b).

Effector memory T cells, which are antigen-specific T cells that secrete cytokines, have been identified in adipose tissue of dairy cattle (Oliveira *et al.*, 2019); however, cytokines produced by these cells in metabolically stressed or healthy cattle have yet to be elucidated. Extrapolation from studies conducted in mice should be done with caution, as species differences between mouse, non-human primate and humans have already been identified in the frequency of Treg cells in adipose tissue (Laparra *et al.*, 2019). Supporting this concern, postpartum dairy cattle are characterized as having an anti-inflammatory T cell phenotype compared to late-lactation cows (Shafer-Weaver and Sordillo, 1997; Shafer-Weaver *et al.*, 1999), and the production of IFN- γ from stimulated peripheral blood mononuclear cells isolated from peripartum dairy cattle was negatively correlated with non-esterified fatty acid (NEFA) concentrations (Lacetera *et al.*, 2005). Taken together, there seems to be a conflict between research conducted in obesity-induced mouse models, suggestive of a pro-inflammatory T cell

polarization in adipose, and peripartum dairy cattle, suggestive of an anti-inflammatory T cell polarization from blood samples. However, similar to observations in mouse models (Feuerer *et al.*, 2009), Treg cells were abundant in mesenteric adipose tissue from healthy, lean dairy cattle (Aylward *et al.*, 2019), although a comparison between over-fed, feed-restricted or diseased cattle was not conducted. Potentially more informative, Kosteli *et al.* (2010) identified a recruitment of M2 macrophages from circulation into adipose tissue in fasted obese mice, and that these cells played an integral role on tempering lipolysis. This model may better reflect what occurs in transition dairy cattle, especially considering that over-conditioned dairy cows experience more extreme negative energy balance during early lactation and are at a greater risk of disease. Could it be that the anti-inflammatory T cell bias found in postpartum cows is a consequence of anti-inflammatory cytokines generated by a larger population of M2 macrophages? Moreover, could these anti-inflammatory T cells exist to decelerate lipolysis? Speculation aside, much research is needed to illuminate T cell phenotypes and their role in adipose tissue in peripartum cows. $\gamma\delta$ T cells, which possess both innate and adaptive immune cell properties, are of great interest due to their abundance in adipose tissue in dairy cattle (~40% of all T cells; Oliveira *et al.*, 2019) as well as their apparent role in the recognition of lipid antigens (Champagne, 2011).

Metabolites and lipid mediators influence inflammatory tone

Negative energy balance in early-lactation dairy cows leads to the mobilization of fat reserves, increasing serum NEFA. These fatty acids can be metabolized in the liver for the purpose of energy production, ketone production (e.g. β -hydroxybutyrate, **BHB**); or in cases of excessive fat mobilization, fat can accumulate in the liver. Moreover, metabolic demands in early lactation augment the production of reactive oxygen species (**ROS**), leading to oxidative stress. Metabolites, including saturated fatty acids, as well as damage-associated molecular patterns, can influence inflammatory responses (Bradford *et al.*, 2015). Immune cells can be activated through a variety of mechanisms, including toll-like receptors (**TLR**), nuclear receptors such as peroxisome proliferator-activated receptors (**PPAR**), and intracellular sensors such as inflammasomes (e.g. nucleotide-binding domain, leucine-rich containing family, pyrin domain-containing-3, also known as **NLRP3**). While some of these receptors, such as TLR, are more typically thought of as a tool to identify pathogens via pathogen-associated molecular patterns, some metabolites can also activate these pathways. For example, saturated fatty acids activate TLR4 signaling in numerous immune cells, including macrophages (Lee *et al.*, 2001), as well as non-immune cells such as hepatocytes (Mamedova *et al.*, 2013), though perhaps not through direct receptor–ligand interaction (Lancaster *et al.*, 2018). The activation of TLR4 leads to increased production of pro-inflammatory

cytokines, including TNF- α , which in turn induces insulin resistance in parenchymal tissue to spare glucose for immune cells to use in driving oxidative burst. It is telling that inflammatory M1 macrophages obtain energy through glycolysis, whereas M2 macrophages obtain energy from oxidative phosphorylation (Galván-Peña and O'Neill, 2014). Fatty acids can also activate PPAR, which are nuclear receptors that function as transcription factors to regulate gene expression. For example, the activation of PPAR γ via n-3 polyunsaturated fatty acids attenuates inflammatory responses in mouse monocytes (Lee *et al.*, 2003), and some similar evidence exists in periparturient dairy cattle (Silvestre *et al.*, 2011; Trevisi *et al.*, 2011).

Lastly, inflammasomes are cytoplasmic sensors, such as NLRP3, that are triggered by microbial molecules in addition to damage signals related to metabolic stress, including saturated fatty acids and ROS and inhibited by ketone bodies such as BHB. NLRP3 controls the expression of caspase-1, which is used for the maturation of pro-inflammatory cytokines IL-1 β and IL-18. More specifically, caspase-1 cleaves pro-IL-1 β and pro-IL-18, resulting in the activation of these cytokines. While little is known about inflammasomes in dairy cattle, a relationship has been established between excessive activation of NLRP3 and human diseases such as cardiovascular disease, type 2 diabetes and obesity (Haneklaus and O'Neill, 2015). Saturated fatty acids, such as palmitate, activate NLRP3 and induce insulin resistance, likely through IL-1 β (Wen *et al.*, 2011). Similarly, knocking out NLRP3 improved insulin signaling, reduced the amount of IFN- γ mRNA and protein and reduced the number of effector memory T cells in adipose tissue (Vandanmagsar *et al.*, 2011). In contrast to the activation by saturated fatty acids, BHB attenuates inflammatory responses associated with NLRP3 (Youm *et al.*, 2015; Goldberg *et al.*, 2017). Moreover, in dairy cattle, a continuous intravenous infusion of BHB decreased glucose concentrations in the blood of postpartum dairy cattle, but had no effect on insulin concentrations, which could be due to enhanced insulin sensitivity (Zarrin *et al.*, 2017). In human disease models, BHB suppressed the synthesis of TNF- α (Yamanashi *et al.*, 2017) and IL-1 β (Youm *et al.*, 2015; Goldberg *et al.*, 2017). Taken together, these data suggest a potential anti-inflammatory mechanism via BHB suppression of inflammasome activation, subsequently improving insulin sensitivity.

Another class of potent lipid mediators are the oxylipids, which have been reviewed in much greater detail elsewhere (Sordillo, 2018). Oxylipids (also referred to as eicosanoids) are rapidly synthesized from polyunsaturated fatty acids (PUFA) during inflammatory events. Polyunsaturated fatty acids can be oxygenated either enzymatically (cyclooxygenase, lipoxygenase or cytochrome P450 epoxygenase pathways) or through non-enzymatic pathways. Moreover, oxylipids can exert either pro- or anti-inflammatory responses, and this effect can be dependent upon the substrate. Simply put, n-3 PUFA typically yield more anti-inflammatory or resolving oxylipids, whereas n-6 PUFA yield more pro-inflammatory oxylipids (Contreras *et al.*, 2012; Raphael and

Sordillo, 2013). To add to the complexity of these pathways, pro-inflammatory oxylipids can be further metabolized into anti-inflammatory oxylipids, highlighting their role in both the onset and resolution of inflammation (Sordillo, 2018). As previously mentioned, metabolic inflammation is characterized as a chronic, low-grade inflammatory state. In early-lactation dairy cows, an increase in plasma pro-inflammatory oxylipids was associated with elevated concentrations of pro-inflammatory cytokines (Raphael *et al.*, 2014). Moreover, the ratio of pro- to anti-inflammatory oxylipids (such as 9-HODE to 9-oxoODE and 13-HODE to 13-oxoODE) was increased in early lactation, but this ratio decreased as cows approached peak lactation (Raphael *et al.*, 2014). The administration of exogenous TNF- α during the first week of lactation prevented an increase in plasma anti-inflammatory oxylipids during that week (Yuan *et al.*, 2013), demonstrating a cross-talk between cytokine and oxylipid signaling in transition cows. It should be noted that oxylipid profiles can vary between tissue, milk or plasma samples (Mavangira *et al.*, 2015; Contreras *et al.*, 2017), and thus immune responses in one organ could be affected differently than in another organ. This is particularly true for adipose tissue, as the ratio of pro- to anti-inflammatory oxylipids (i.e. 9-HODE:9-oxoODE and 13-HODE:13-oxoODE) is considerably smaller than in plasma (Contreras *et al.*, 2017). Considering all of these findings, changes in lipid metabolism in early-lactation dairy cows likely play a key role in determining the inflammatory tone, although this effect may vary depending on the organ.

Inflammatory tone is maintained in the face of exogenous agents

The evidence above points to an immune–metabolic cross-talk underlying inflammatory tone in early-lactation dairy cattle. However, if inflammatory tone is truly controlled in a homeostatic manner, then it should be possible to observe counter-regulatory responses to exogenous 'nudges' in one direction or another. In fact, we observed signs of such counter-regulatory mechanisms in three studies, described below.

In one study, we used oral administration of a non-steroidal anti-inflammatory drug (NSAID), sodium salicylate, to alter inflammatory signaling in early-lactation dairy cattle (Farney *et al.*, 2013). Although this treatment generally did not cause measurable changes in circulating inflammatory mediators during the 7-day treatment period, we observed a marked rise in pro-inflammatory oxylipids in blood plasma from salicylate-treated cows a week *after* the treatment ended. This surprising finding led us to suggest that a homeostatic mechanism maintained inflammatory tone in the face of exogenous salicylate by upregulating pro-inflammatory pathways, potentially explaining the limited effects during the treatment window. When the exogenous agent was removed, this shift in endogenous tone was observed as an inflammatory rebound (Farney *et al.*, 2013). Similar findings have been reported in humans (Pijak, 2006).

Another observation consistent with the results described above was an increase in immune signaling proteins in adipose tissue of cows treated with salicylate for the first 5 days of lactation (Takiya *et al.*, 2019). Altered proteins clearly pointed to enhanced complement activation as well as a likely increase in macrophage populations in the tissue in response to oral salicylate. Although other 'off-target' mechanisms cannot be ruled out, this enhancement of adipose tissue inflammatory mechanisms may provide the underpinning for homeostatic maintenance of inflammatory signals in the bloodstream of cows treated with this NSAID.

A third study, this time employing pro-inflammatory cytokine administration, provided further evidence of a regulated inflammatory tone. In this experiment, we utilized miniature osmotic pumps to continuously deliver a small amount of recombinant bovine TNF- α into a subcutaneous adipose tissue depot of lactating dairy cows (Martel *et al.*, 2014). Unlike studies where a daily bolus administration of an equal amount of TNF- α altered the metabolism of cows (Bradford *et al.*, 2009; Yuan *et al.*, 2013), this continuous administration had no measurable impact on metabolism. Furthermore, transcriptional responses in both liver and contralateral adipose tissue pointed to decreased inflammatory signaling, a surprising finding given the pro-inflammatory nature of the cytokine infused. The most convincing evidence of a counter-regulatory anti-inflammatory response to TNF- α administration was a highly significant increase in IL-10 protein in the contralateral adipose depot (Martel *et al.*, 2014), potentially due to a cross-talk between the immune system and the sympathetic nervous system (Bowers *et al.*, 2004) to inhibit TNF- α production (Elenkov *et al.*, 1995). Here again, a subtle and chronic exogenous 'nudge' was counteracted by an apparent homeostatic response in the opposite direction.

Altered set-points for an inflammatory tone: adaptive or pathological?

A hallmark of homeostasis is the use of negative feedback mechanisms to keep a regulated variable near its set-point. Inflammatory responses do not seem to align with this paradigm, because the cytokine storm (along with changes in other mediators) that accompanies an acute inflammatory event causes numerous deviations from typical set-points. The inflammatory response, though, is normally followed by a pre-programming resolution wave that does, in time, bring systems back into homeostasis (Serhan and Savill, 2005; Sugimoto *et al.*, 2019). From the vantage point of days, rather than minutes, perhaps the inflammatory network does not differ from endocrine regulatory systems so much.

In many homeostatic control systems, regulatory targets can be altered, either through pathology or homeorhesis. Considering the inflammatory tone as a regulated system, therefore, is not inconsistent with either acute inflammatory and anti-inflammatory cytokine waves in response to disease or with chronic elevation of inflammatory mediators in obesity. Temporary or chronic changes in inflammatory

set-points allow for the activation of various components of the immune system and for alterations in metabolism; whether these changes are considered homeorhetic or pathological ultimately comes down to whether we view them as adaptive or not.

A well-described example of homeorhetic regulation of inflammatory tone is endotoxin tolerance. In this phenomenon, the primary exposure to lipopolysaccharide (LPS) induces an acute inflammatory response, whereas chronic or repeated exposures lead to diminished responses in many animals (Elsasser *et al.*, 2005; Petzl *et al.*, 2012). The concept that a low dose of a potential toxin can protect against subsequent exposures that should be harmful is called hormesis (Mattson, 2008). Although endotoxin tolerance is still being studied, several contributing mechanisms have been described. Endotoxin exposure increases the shedding of TNF- α receptors on monocytes and increases the concentration of soluble 'decoy' TNF- α receptors in circulation (van der Poll *et al.*, 1995). Moreover, LPS signaling via the receptor CD14 results in TLR4 endocytosis, diminishing its ability to generate intracellular responses to LPS (Zanoni *et al.*, 2011). Similarly, the release of the acute-phase protein, lipopolysaccharide-binding protein (LBP), facilitates an inflammatory response to endotoxin at low concentrations, but at high concentrations, LBP neutralizes endotoxin in lipid membranes, resulting in the reduction of TNF- α synthesis (Gutsmann *et al.*, 2001). Each of these mechanisms enables a more tolerant response to LPS, noted by a reduction of pro-inflammatory cytokine synthesis in addition to mechanisms negating the effects of a potential cytokine storm, thus allowing for a more stable inflammatory tone in the face of chronic mild endotoxin exposure.

While the ability of the immune system to develop tolerance toward microbial stimuli has been well established, mechanisms underlying homeostasis in metabolic inflammation have not been clearly described. However, it is noteworthy that the anti-inflammatory effect of adiponectin (an adipocyte-derived cytokine) on macrophages is through the induction of immunological tolerance to pro-inflammatory stimuli (Tsatsanis *et al.*, 2005), in addition to stimulating the production of anti-inflammatory cytokines (Wolf *et al.*, 2004; Wulster-Radcliffe *et al.*, 2004). It seems probable that metabolic inflammation is enhanced in periparturient dairy cattle due, in part, to reductions in adiponectin (Kabara *et al.*, 2014).

Alterations in metabolic flux by inflammatory signaling in adipose tissue

Inflammatory mediators work on a variety of target organs to influence nutrient influx and partitioning. Although most postpartum dairy cows are in a catabolic state requiring the release of stored nutrient reserves, the magnitude of this catabolism varies greatly among cows. There is evidence suggesting that inflammatory mediators, working in part through altered insulin sensitivity, may underlie some of

the variation in the rate of tissue catabolism in early lactation (Zachut *et al.*, 2013; Zachut, 2015). This is consistent with the emerging roles of immune cells and signals in regulating adipose tissue metabolism.

Lipolysis is characterized by an orchestrated infiltration of a variety of immune cells, including monocytes, granulocytes and lymphocytes (Odegaard and Chawla, 2013). Pro-inflammatory activation of these cells results in the release of cytokines such as TNF- α . In turn, TNF- α promotes lipolysis through a variety of mechanisms; the injection of recombinant TNF- α in cattle reduced feed intake (Kushibiki *et al.*, 2003) and insulin sensitivity (Kushibiki *et al.*, 2001), and directly stimulated lipolysis (Kushibiki *et al.*, 2002). The pro-lipolytic effects of inflammatory mediators such as TNF- α are consistent with the typical catabolic response to disease.

In contrast, an elegant series of experiments illuminated an opposing immune mechanism in mice (Kosteli *et al.*, 2010). Caloric restriction in obese mice resulted in a greater number of M2 macrophages in adipose at the onset of negative energy balance (Kosteli *et al.*, 2010), which was also found in both diseased early-lactation dairy cattle and feed-restricted cows compared to cows in an anabolic state (Contreras *et al.*, 2015; 2016). Anti-inflammatory macrophages attenuated lipolysis in obese mice during fasting, despite the fact that TNF- α was greater in obese mice compared to lean mice fed a normal chow diet (Kosteli *et al.*, 2010). These data suggest that immunometabolic control of lipolysis is driven by the balance of M1 and M2 macrophages. The simultaneous activity of both M1 and M2 macrophages is consistent with the model of inflammatory homeostasis discussed previously.

One key shortcoming in much of the research conducted with inflammatory mediators has been the short-term nature of studies. We know a tremendous amount about cytokine impacts on metabolic flux via transcriptional or post-translational impacts on nutrient transporters and enzymes, but impacts on tissue remodeling have been largely ignored. This is not a trivial oversight; aside from driving immune responses, inflammatory signaling is probably used most widely in controlling proliferation and differentiation of cells (Cao *et al.*, 2001; Langen *et al.*, 2001; Baldassarre *et al.*, 2004). For an example of how this can lead to misinterpretation, consider a series of studies with mice engineered to limit inflammatory signaling in adipose tissue (Wernstedt Asterholm *et al.*, 2014). Inflammatory mediators generally have lipolytic effects, and these studies of two mouse models were well designed to determine whether eliminating cytokine signaling would enhance adipose triglyceride accretion. In fact, the opposite occurred. Suppressing cytokine signaling inhibited adipogenesis, constraining adipose tissue accretion in growing mice and ultimately resulting in more severe fatty liver disease (Wernstedt Asterholm *et al.*, 2014). In this case, removing cytokine stimulation of adipocyte differentiation had a greater impact on tissue mass than did the removal of the lipolytic stimulus. It is important to keep in mind the potential effects of inflammatory mediators on cellular dynamics in addition to metabolic enzymes and transporters.

Implications of non-steroidal anti-inflammatory therapy on dairy cow metabolism and productivity

A growing number of studies have evaluated the use of NSAID to modulate the transition to lactation in dairy cattle, and several have demonstrated interesting impacts on metabolism.

Treatment of dairy cows with oral sodium salicylate for the first 7 days of lactation caused a substantial decrease in blood glucose concentration, particularly among older cows (Farney *et al.*, 2013). This occurred in the absence of any measurable differences in energy balance compared to control cows during treatment, suggesting that a fuel-specific mechanism was responsible for this effect. We subsequently conducted a follow-up study where multiparous cows were treated with oral sodium salicylate, and plasma glucose turnover was determined with a stable isotope dilution method on day 7 of lactation. We found that glucose turnover rate was decreased by 25%, despite a significant decrease in insulin–glucagon ratio (Montgomery *et al.*, 2019). Given the steady-state assumptions used in metabolic flux studies, the decline in glucose turnover reflected both a decreased use of glucose and a decreased supply. Therefore, somewhat surprisingly, the evidence pointed to an inhibition of gluconeogenesis as the primary factor underlying salicylate-induced hypoglycemia in transition dairy cows, rather than enhanced peripheral utilization of glucose. It remains unclear exactly how sodium salicylate inhibited gluconeogenesis, but one plausible mechanism may be that it prevented inflammatory induction of gluconeogenesis (Mamedova *et al.*, 2013).

Similar peripartum NSAID treatment strategies have also resulted in substantial increases in milk secretion (Trevisi and Bertoni, 2008; Carpenter *et al.*, 2016; Swartz *et al.*, 2018). Treatment protocols that provided meaningful NSAID to the system for only a few days had rather astounding effects, with milk yield increases of 10% to 15% lasting 10 months. As in the discussion of potential impacts on adipose tissue metabolic flux, the net increase in milk yield could plausibly be due to an increase in either secretory epithelial cell abundance in the mammary gland or to a greater metabolic flux in each secretory cell. Further complicating the matter is the 6-week lag prior to milk responses (Carpenter *et al.*, 2016). Extra-mammary mechanisms may be the primary impact of treatment; for example, alterations in feeding behavior could possibly result in greater nutrient supply, allowing the mammary gland to better meet its potential to synthesize milk (Carpenter *et al.*, 2018). On the other hand, more direct effects are also possible. We and others have observed a reduction in early-lactation somatic cells in milk of cows treated with NSAID (Carpenter *et al.*, 2016; Shock *et al.*, 2018), and inflammatory signals from neutrophils can directly inhibit milk component synthesis both immediately (Connelly *et al.*, 2010) and over the longer term, through epigenetic mechanisms (Vanselow *et al.*, 2006). Although the mode of action remains to be clarified, the dramatic and sustained increase in milk secretion

following early-lactation NSAID treatment provides a potent example of just how powerful the immunometabolic complex is at the onset of lactation.

Burning questions

While numerous advances have clarified our view on homeostatic mechanisms related to metabolic inflammation, many questions are left unanswered, particularly in reference to periparturient dairy cattle. Elucidating these control mechanisms may better inform researchers searching for potential solutions to improve dairy cattle productivity and immunity. A few of these outstanding questions are outlined below.

- What is the biological purpose for maintaining inflammatory tone during NSAID therapy in periparturient dairy cattle?
- How does immune cell cross-talk between monocytes, lymphocytes and granulocytes impact inflammatory tone in bovine adipose tissue, and what are the subsequent effects on metabolic adaptations to lactation?
- Do monocytes develop immunological tolerance toward metabolite signals, and is this a reason for the low-grade inflammatory tone?
- Are there beneficial or deleterious effects from the rebound inflammatory response that occurs following NSAID administration? Does this effect contribute to long-term increases in milk yield, potentially through enhanced mammary gland development?


Conclusions

Resident immune cells are found in all metabolically important tissues, and they have a significant impact on nutrient flux in adipose tissue and perhaps other organs. These impacts are not unidirectional; in fact, adipocytes can alter immune cell phenotypes through both cytokine and nutrient release, whereas resident immune cells serve to clear out residual lipids and also modulate lipolysis. The cross-talk between cells in adipose tissue may have an outsized influence on whole-body inflammatory tone, and may provide for homeostatic regulation of the balance between immune protection and tolerance. Altering the inflammatory tone, even in subtle ways, can have surprisingly large impacts on whole-body nutrient use, including rates of lipolysis, gluconeogenesis and galactopoiesis. Peripartum NSAID administration has drawn attention not only for potential health benefits but also because of these metabolic responses. Although much remains to be understood about site and mode of action of these NSAID, recent findings nonetheless demonstrate the powerful impact of inflammatory signals in the transition period.

Acknowledgements

Contribution no. 19-277-J from the Kansas Agricultural Experiment Station. A brief preliminary version of this work has been published in abstract form (Bradford and Swartz,

2019). The work at Kansas State University described in this review was supported by the USDA National Institute of Food and Agriculture, Hatch project 1018048, and Agriculture and Food Research Initiative Competitive Grant no. 2013-01976, as well as by the National Science Foundation (grant #1456794).

 B. J. Bradford 0000-0002-6775-4961

Declaration of interest

Turner Swartz declares no conflict of interest. Barry Bradford has received speaking and/or consulting fees from Bayer AG, Merck Animal Health, Zoetis and Elanco.

Ethics statement

No primary research requiring ethics approvals are reported in this review paper.

Software and data repository resources

None of the findings were deposited in an official repository.

References

- Ahmed M and Gaffen SL (2010) IL-17 in obesity and adipogenesis. *Cytokine & Growth Factor Reviews* 21, 449–453.
- Akter SH, Häussler S, Germeroth D, Von Soosten D, Dänicke S, Südekum KH and Sauerwein H (2012) Immunohistochemical characterization of phagocytic immune cell infiltration into different adipose tissue depots of dairy cows during early lactation. *Journal of Dairy Science* 95, 3032–3044.
- Aylward BA, Clark ML, Galileo DS, Baernard AM, Wilson JR, Brannick E, Gressley T, Fecteau ME, Davis WC and Dyer RM (2019) Immune cell populations residing in mesenteric adipose depots and mesenteric lymph nodes of lean dairy cows. *Journal of Dairy Science* 102, 3452–3468.
- Bai Y and Sun Q (2015) Macrophage recruitment in obese adipose tissue. *Obesity Reviews* 16, 127–136.
- Baldassarre G, Nicoloso MS, Schiappacassi M, Chimienti E and Belletti B (2004) Linking inflammation to cell cycle progression. *Current Pharmaceutical Design* 10, 1653–1666.
- Bolus WR, Gutierrez DA, Kennedy AJ, Anderson-Baucum EK and Hasty AH (2015) CCR2 deficiency leads to increased eosinophils, alternative macrophage activation, and type 2 cytokine expression in adipose tissue. *Journal of Leukocyte Biology* 98, 467–477.
- Bowers RR, Festuccia WT, Song CK, Shi H, Migliorini RH and Bartness TJ (2004). Sympathetic innervation of white adipose tissue and its regulation of fat cell number. *American Journal of Physiology-Regulatory, Integrative and Comparative Physiology* 286, R1167–R1175.
- Bradford BJ, Mamedova LK, Minton JE, Drouillard JS and Johnson BJ (2009) Daily injection of tumor necrosis factor- α increases hepatic triglycerides and alters transcript abundance of metabolic genes in lactating dairy cattle. *Journal of Nutrition* 139, 1451–1456.
- Bradford BJ and Swartz TH (2019) Toward a homeostatic view of inflammation: the transition dairy cow example. In *Proceedings of the 17th International Conference on Production Diseases in Farm Animals*, 27–29 June 2019, Bern, Switzerland, pp. 88.
- Bradford BJ, Yuan K, Farney JK, Mamedova LK and Carpenter AJ (2015) Invited review: inflammation during the transition to lactation: new adventures with an old flame. *Journal of Dairy Science* 98, 6631–6650.
- Cao Y, Bonizzi G, Seagroves TN, Greten FR, Johnson R, Schmidt E V and Karin M (2001) IKK α provides an essential link between RANK signaling and cyclin D1 expression during mammary gland development. *Cell* 107, 763–775.
- Carpenter AJ, Ylloja CM, Mamedova LK, Olagaray KE and Bradford BJ (2018) Effects of early postpartum sodium salicylate treatment on long-term milk, intake, and blood parameters of dairy cows. *Journal of Dairy Science* 101, 1437–1447.

- Carpenter AJ, Ylloja CM, Vargas CF, Mamedova LK, Mendonca LG, Coetzee JF, Hollis LC, Gehring R and Bradford BJ (2016) Hot topic: early postpartum treatment of commercial dairy cows with nonsteroidal antiinflammatory drugs increases whole-lactation milk yield. *Journal of Dairy Science* 99, 672–679.
- Champagne E (2011) $\gamma\delta$ T cell receptor ligands and modes of antigen recognition. *Archivum Immunologiae Therapiae Experimentalis* 59, 117–137.
- Chatzigeorgiou A, KP Karalis, SR Bornstein and T Chavakis (2012) Lymphocytes in obesity-related adipose tissue inflammation. *Diabetologia* 55, 2583–2592.
- Chawla A, Nguyen KD and Goh YPS (2011) Macrophage-mediated inflammation in metabolic disease. *Nature Reviews Immunology* 11, 738–749.
- Chistiakov DA, Bobryshev YV, Kozarov E, Sobenin IA and Orekhov AN (2015) Intestinal mucosal tolerance and impact of gut microbiota to mucosal tolerance. *Frontiers in Microbiology* 5, 781.
- Connelly L, Barham W, Pigg R, Saint-Jean L, Sherrill T, Cheng DS, Chodosh LA, Blackwell TS and Yull FE (2010) Activation of nuclear factor kappa B in mammary epithelium promotes milk loss during mammary development and infection. *Journal of Cellular Physiology* 222, 73–81.
- Contreras GA, Kabara E, Brester J, Neuder L and Kiupel M (2015) Macrophage infiltration in the omental and subcutaneous adipose tissues of dairy cows with displaced abomasum. *Journal of Dairy Science* 98, 6176–6187.
- Contreras GA, Mattmiller SA, Raphael W, Gandy JC and Sordillo LM (2012) Enhanced n-3 phospholipid content reduces inflammatory responses in bovine endothelial cells. *Journal of Dairy Science* 95, 7137–7150.
- Contreras GA, Strieder-Barboza C and De Koster J (2018) Symposium review: modulating adipose tissue lipolysis and remodeling to improve immune function during the transition period and early lactation of dairy cows. *Journal of Dairy Science* 101, 2737–2752.
- Contreras GA, Strieder-Barboza C, De Souza J, Gandy J, Mavangira V, Lock AL and Sordillo LM (2017) Periparturient lipolysis and oxylipid biosynthesis in bovine adipose tissues. *PLoS ONE*, 12, e0188621.
- Contreras GA, Thelen K, Schmidt SE, Strieder-Barboza C, Preseault CL, Raphael W, Kiupel M, Caron J and Lock AL (2016) Adipose tissue remodeling in late-lactation dairy cows during feed-restriction-induced negative energy balance. *Journal of Dairy Science* 99, 10009–10021.
- Desruisseaux MS, Nagajothi ME, Trujillo HB, Tanowitz ME and Scherer PE (2007) Adipocyte, adipose tissue, and infectious disease. *Infection and Immunity* 75, 1066–1078.
- Elenkov IJ, Haskó G, Kovács KJ and Vizi ES (1995) Modulation of lipopolysaccharide-induced tumor necrosis factor- α production by selective α - and β -adrenergic drugs in mice. *Journal of Neuroimmunology* 61, 123–131.
- Elsasser TH, Blum JW and Kahl S (2005) Characterization of calves exhibiting a novel inheritable TNF-alpha hyperresponsiveness to endotoxin: associations with increased pathophysiological complications. *Journal of Applied Physiology* 98, 2045–2055.
- Farney JK, Mamedova LK, Coetzee JF, KuKanich B, Sordillo LM, Stoakes SK, Minton JE, Hollis LC and Bradford BJ (2013) Anti-inflammatory salicylate treatment alters the metabolic adaptations to lactation in dairy cattle. *American Journal of Physiology Regulatory, Integrative and Comparative Physiology* 305, R110–R117.
- Ferreira RC, Forsyth LE, Richman PI, Wells C, Spencer J and MacDonald TT (1990) Changes in the rate of crypt epithelial cell proliferation and mucosal morphology induced by a T-cell-mediated response in human small intestine. *Gastroenterology* 98, 1255–1263.
- Feuerer M, Herrero L, Cipolletta D, Naaz A, Wong J, Nayer A, Lee J, Goldfine AB, Benoist C, Shoelson S and Mathis D (2009) Lean, but not obese, fat is enriched for a unique population of regulatory T cells that affect metabolic parameters. *Nature Medicine* 15, 930–939.
- Galván-Peña S and O'Neill LAJ (2014) Metabolic reprogramming in macrophage polarization. *Frontiers in Immunology* 5, 1–6.
- García M, Bradford BJ and Nagaraja TG (2017) Invited review: ruminal microbes, microbial products, and systemic inflammation. *The Professional Animal Scientist* 33, 635–650.
- Goldberg EL, Asher JL, Molony RD, Shaw AC, Zeiss CJ, Wang C, Morozova-Roche LA, Herzog RI, Iwasaki A and Dixit VD (2017) β -Hydroxybutyrate deactivates neutrophil NLRP3 inflammasome to relieve gout flares. *Cell Reports* 18, 2077–2087.
- Gutsmann T, Müller M, Carroll SF, MacKenzie RC, Wiese A and Seydel U (2001) Dual role of Lipopolysaccharide (LPS)-binding protein in neutralization of LPS and enhancement of LPS-induced activation of mononuclear cells. *Infection and Immunity* 69, 6942–6950.
- Haneklaus M and O'Neill LAJ (2015) NLRP3 at the interface of metabolism and inflammation. *Immunological Reviews* 265, 53–62.
- Holtenius K, Waller KP, Essen-Gustavsson B, Holtenius P and Sandgren CHJTJV (2004) Metabolic parameters and blood leukocyte profiles in cows from herds with high or low mastitis incidence. *The Veterinary Journal* 168, 65–73.
- Hotamisligil GS (2006) Inflammation and metabolic disorders. *Nature* 444, 860–867.
- Hotamisligil GS, Shargill NS and Spiegelman BM (1993) Adipose expression of tumor necrosis factor- α : direct role in obesity-linked insulin resistance. *Science* 259, 87–91.
- Hoytema van Konijnenburg DP, Reis BS, Pedicord VA, Farache J, Victora GD and Mucida D (2017) Intestinal epithelial and intraepithelial T cell crosstalk mediates a dynamic response to infection. *Cell* 171, 783–794.e13.
- Iyer A, Brown L, Whitehead JP, Prins JB and Fairlie DP (2015) Nutrient and immune sensing are obligate pathways in metabolism, immunity, and disease. *The FASEB Journal* 29, 3612–3625.
- Kabara E, Sordillo LM, Holcombe S and Contreras GA (2014) Adiponectin links adipose tissue function and monocyte inflammatory responses during bovine metabolic stress. *Comparative Immunology, Microbiology and Infectious Diseases* 37, 49–58.
- Khan IM, Dai Perrard XY, Perrard JL, Mansoori A, Smith CW, Wu H and Ballantyne CM (2014) Attenuated adipose tissue and skeletal muscle inflammation in obese mice with combined CD4+ and CD8+ T cell deficiency. *Atherosclerosis* 233, 419–428.
- Khiaosa-Ard R and Zebeli Q (2018) Diet-induced inflammation: from gut to metabolic organs and the consequences for the health and longevity of ruminants. *Research in Veterinary Science* 120, 17–27. doi: [10.1016/j.rvsc.2018.08.005](https://doi.org/10.1016/j.rvsc.2018.08.005).
- Kosteli A, Sagar E, Haemmerle G, Martin JF, Lei J, Zechner R and Ferrante AW (2010) Weight loss and lipolysis promote a dynamic immune response in murine adipose tissue. *The Journal of Clinical Investigation* 120, 3466–3479.
- Kushibiki S, Hodate K, Shingu H, Hayashi T, Touno E, Shinoda M and Yokomizo Y (2002) Alterations in lipid metabolism induced by recombinant bovine tumor necrosis factor- α administration to dairy heifers. *Journal of Animal Science* 80, 2151–2157.
- Kushibiki S, Hodate K, Shingu H, Obara Y, Touno E, Shinoda M and Yokomizo Y (2003) Metabolic and lactational responses during recombinant bovine tumor necrosis factor- α treatment in lactating cows. *Journal of Dairy Science* 86, 819–827.
- Kushibiki S, Hodate K, Shingu H, Ueda Y, Shinoda M, Mori Y, Itoh T and Yokomizo Y (2001) Insulin resistance induced in dairy steers by tumor necrosis factor alpha is partially reversed by 2,4-thiazolidinedione. *Domestic Animal Endocrinology* 21, 25–37.
- Lacetera N, Scalia D, Bernabucci U, Ronchi B, Pirazzi D and Nardone A (2005) Lymphocyte functions in overconditioned cows around parturition. *Journal of Dairy Science* 88, 2010–2016.
- Lancaster GI, Langley KG, Berglund NA, Kammoun HL, Reibe S, Estevez E, Weir J, Mellett NA, Pernes G, Conway JRW, Lee MKS, Timpson P, Murphy AJ, Masters SL, Gerondakis S, Bartonicek N, Kaczorowski DC, Dinger ME, Meikle PJ, Bond PJ and Febbraio MA (2018) Evidence that TLR4 is not a receptor for saturated fatty acids but mediates lipid-induced inflammation by reprogramming macrophage metabolism. *Cell Metabolism* 27, 1096–1110.e5.
- Langen RCJ, Schols AMWJ, Kelders MCJM, Wouters EFM and Janssen-Heininger YMW (2001) Inflammatory cytokines inhibit myogenic differentiation through activation of nuclear factor- κ B. *The FASEB Journal* 15, 1169–1180.
- Laparra A, Tricot S, Le Van M, Damouche A, Gorwood J, Vaslin B, Favier B, Benoist S, Ho Tsong Fang R, Bosquet N, Le Grand R, Chapon C, Lambotte O and Bourgeois C (2019) The frequencies of immunosuppressive cells in adipose tissue differ in human, non-human primate, and mouse models. *Frontiers in Immunology* 10, 117–117.
- Lee JY, Plakidas A, Lee WH, Heikkinen A, Chanmugam P, Bray G and Hwang DH (2003) Differential modulation of toll-like receptors by fatty acids preferential inhibition by n-3 polyunsaturated fatty acids. *Journal of Lipid Research* 44, 479–486.
- Lee JY, Sohn KH, Rhee SH and Hwang DJ (2001) Saturated fatty acids, but not unsaturated fatty acids, induce the expression of cyclooxygenase-2 mediated through Toll-like receptor 4. *Journal of Biological Chemistry* 276, 16683–16689.
- Lopreato V, Hosseini A, Rosa F, Zhou Z, Alharthi A, Trevisi E and Looor JJ (2018) Dietary energy level affects adipose depot mass but does not impair in vitro

- subcutaneous adipose tissue response to short-term insulin and tumor necrosis factor- α challenge in nonlactating, nonpregnant Holstein cows. *Journal of Dairy Science* 101, 10206–10219.
- Lumeng CN, Bodzin JL and Saltiel AR (2007) Obesity induces a phenotypic switch in adipose tissue macrophage polarization. *The Journal of Clinical Investigation* 117, 175–184.
- Mamedova LK, Yuan K, Laudick AN, Fleming SD, Mashek DG and Bradford BJ (2013) Toll-like receptor 4 signaling is required for induction of gluconeogenic gene expression by palmitate in human hepatic carcinoma cells. *Journal of Nutritional Biochemistry* 24, 1499–1507.
- Man K, Kutayav VI and Chawla A (2017) Tissue immunometabolism: development, physiology, and pathobiology. *Cell Metabolism* 25, 11–26.
- Martel CA, Mamedova LK, Minton JE, Jones ML, Carroll JA and Bradford BJ (2014) Continuous low-dose infusion of tumor necrosis factor alpha in adipose tissue elevates adipose tissue interleukin 10 abundance and fails to alter metabolism in lactating dairy cows. *Journal of Dairy Science* 97, 4897–4906.
- Mattson MP (2008) Hormesis defined. *Ageing Research Reviews* 7, 1–7.
- Mavangira V, Gandy JC, Zhang C, Ryman VE, Jones AD and Sordillo LM (2015) Polyunsaturated fatty acids influence differential biosynthesis of oxylipids and other lipid mediators during bovine coliform mastitis. *Journal of Dairy Science* 98, 6202–6215.
- McCambridge G, Agrawal M, Keady A, Kern PA, Hasturk H, Nikolajczyk BS and Bharath LP (2019) Saturated fatty acid activates T cell inflammation through a Nicotinamide Nucleotide Transhydrogenase (NNT)-dependent mechanism. *Biomolecules* 9, 79.
- McGillicuddy FC, Chiquoine EH, Hinkle CC, Kim RJ, Shah R, Roche HM, Smyth EM and Reilly MP (2009) Interferon γ attenuates insulin signaling, lipid storage, and differentiation in human adipocytes via activation of the JAK/STAT pathway. *Journal of Biological Chemistry* 284, 31936–31944.
- McLaughlin T, Liu L-F, Lamendola C, Shen L, Morton J, Rivas H, Winer D, Tolentino L, Choi O, Zhang H, Hui Yen Chng M and Engleman E (2014) T-Cell profile in adipose tissue is associated with insulin resistance and systemic inflammation in humans. *Arteriosclerosis, Thrombosis, and Vascular Biology* 34, 2637–2643.
- Medzhitov R (2008) Origin and physiological roles of inflammation. *Nature* 454, 428–435.
- Mehta P, Nootio-Antar AM and Smith CW (2015) $\gamma\delta$ T cells promote inflammation and insulin resistance during high fat diet-induced obesity in mice. *Journal of Leukocyte Biology* 97, 121–134.
- Moisá SJ, Ji P, Drackley JK, Rodriguez-Zas SL and Looor JJ (2017) Transcriptional changes in mesenteric and subcutaneous adipose tissue from Holstein cows in response to plane of dietary energy. *Journal of Animal Science and Biotechnology* 8, 85.
- Molofsky AB, Nussbaum JC, Liang H-E, Van Dyken SJ, Cheng LE, Mohapatra A, Chawla A and Locksley RM (2013) Innate lymphoid type 2 cells sustain visceral adipose tissue eosinophils and alternatively activated macrophages. *Journal of Experimental Medicine* 210, 535–549.
- Montgomery SR, Mamedova LK, Zachut M, Kra G, Häussler S, Vaughn M, Gonzalez J and Bradford BJ (2019) Effects of sodium salicylate on glucose kinetics and insulin signaling in postpartum dairy cows. *Journal of Dairy Science* 102, 1617–1629.
- Morris DL, Singer K and Lumeng CN (2011) Adipose tissue macrophages: phenotypic plasticity and diversity in lean and obese states. *Current Opinion in Clinical Nutrition and Metabolic Care* 14, 341–346.
- Mukesh M, Bionaz M, Graugnard DE, Drackley JK and Looor JJ (2010) Adipose tissue depots of Holstein cows are immune responsive: inflammatory gene expression in vitro. *Domestic Animal Endocrinology* 38, 168–178.
- Newman AW, Miller A, Yepes FAL, Bitsko E, Nydam D and Mann S (2019) The effect of the transition period and postpartum body weight loss on macrophage infiltrates in bovine subcutaneous adipose tissue. *Journal of Dairy Science* 102, 1693–1701.
- Nishimura S, Manabe I, Nagasaki M, Eto K, Yamashita H, Ohsugi M, Otsu M, Hara K, Ueki K, Sugiura S, Yoshimura K, Kadowaki T and Nagai R (2009) CD8⁺ effector T cells contribute to macrophage recruitment and adipose tissue inflammation in obesity. *Nature Medicine* 15, 914–920.
- O'Rourke RW, White AE, Metcalf MD, Winters BR, Diggs BS, Zhu X and Marks DL (2012) Systemic inflammation and insulin sensitivity in obese IFN- γ knockout mice. *Metabolism* 61, 1152–1161.
- Odegaard JI and Chawla AJS (2013) Pleiotropic actions of insulin resistance and inflammation in metabolic homeostasis. *Science* 339, 172–177.
- Olefsky JM and Glass CK (2010) Macrophages, inflammation, and insulin resistance. *Annual Review of Physiology* 72, 219–246.
- Oliveira BM, Rasteiro AM, Correia A, Pinto A, Meireles P, Ferreira PG, Vilanova M and Teixeira L (2019) T cells in mesenteric and subcutaneous adipose tissue of Holstein-Friesian cows. *Scientific Reports* 9, 3413.
- Omenetti S and Pizarro TT (2015) The Treg/Th17 axis: a dynamic balance regulated by the gut microbiome. *Frontiers in Immunology* 6, 639.
- Petzl W, Günther J, Pfister T, Sauter-Louis C, Goetze L, von Aulock S, Hafner-Marx A, Schuberth H-JJ, Seyfert H-MM, and Zerbe H (2012) Lipopolysaccharide pretreatment of the udder protects against experimental *Escherichia coli* mastitis. *Innate Immunity* 18, 467–477.
- Pijak M (2006) Rebound inflammation and the risk of ischemic stroke after discontinuation of aspirin therapy. *Archives of Neurology* 63, 300–301.
- Raphael W, Halbert L, Contreras G and Sordillo L (2014) Association between polyunsaturated fatty acid-derived oxylipid biosynthesis and leukocyte inflammatory marker expression in periparturient dairy cows. *Journal of Dairy Science* 97, 3615–3625.
- Raphael W and Sordillo L (2013) Dietary polyunsaturated fatty acids and inflammation: the role of phospholipid biosynthesis. *International Journal of Molecular Sciences* 14, 21167–21188.
- Rocha VZ, Folco EJ, Sukhova G, Shimizu K, Gotsman I, Vernon AH and Libby P (2008) Interferon- γ , a Th1 cytokine, regulates fat inflammation: a role for adaptive immunity in obesity. *Circulation Research* 103, 467–476.
- Sadri H, Bruckmaier RM, Rahmani HR, Ghorbani GR, Morel I and Van Dorland HA (2010) Gene expression of tumour necrosis factor and insulin signalling-related factors in subcutaneous adipose tissue during the dry period and in early lactation in dairy cows. *Journal of Animal Physiology and Animal Nutrition* 94, e194–e202.
- Serhan CN and Savill J (2005) Resolution of inflammation: the beginning programs the end. *Nature Immunology* 6, 1191–1197.
- Shafer-Weaver KA, Corl CM and Sordillo LM (1999) Shifts in bovine CD4⁺ subpopulations increase T-helper-2 compared with T-helper-1 effector cells during the postpartum period. *Journal of Dairy Science* 82, 1696–1706.
- Shafer-Weaver KA and Sordillo LM (1997) Bovine CD8⁺ suppressor lymphocytes alter immune responsiveness during the postpartum period. *Veterinary Immunology and Immunopathology* 56, 53–64.
- Shock DA, Renaud DL, Roche SM, Poliquin R and Olson ME (2018) Evaluating the impact of meloxicam oral suspension administered at parturition on subsequent production, health, and culling in dairy cows: a randomized clinical field trial. *PLoS ONE* 13, e0209236.
- Silvestre F, Carvalho T, Crawford P, Santos J, Staples C, Jenkins T and Thatcher WJ (2011) Effects of differential supplementation of fatty acids during the peripartum and breeding periods of Holstein cows: II. Neutrophil fatty acids and function, and acute phase proteins. *Journal of Dairy Science* 94, 2285–2301.
- Sordillo LM (2018) Symposium review: oxylipids and the regulation of bovine mammary inflammatory responses. *Journal of Dairy Science* 101, 5629–5641.
- Sordillo LM and Raphael W (2013) Significance of metabolic stress, lipid mobilization, and inflammation on transition cow disorders. *Veterinary Clinics: Food Animal Practice* 29, 267–278.
- Steele MA, Penner GB, Chaucheyras-Durand F and Guan LL (2016) Development and physiology of the rumen and the lower gut: targets for improving gut health. *Journal of Dairy Science* 99, 4955–4966.
- Sugimoto MA, Vago JP, Perretti M and Teixeira MM (2019) Mediators of the resolution of the inflammatory response. *Trends in Immunology* 40, 212–227.
- Swartz TH, Schramm HH, Bewley JM, Wood CM, Leslie KE and Petersson-Wolfe CS (2018) Meloxicam administration either prior to or after parturition: Effects on behavior, health, and production in dairy cows. *Journal of Dairy Science* 101, 10151–10167.
- Takiya CS, Montgomery SR, Mamedova LK, Kra G, Nemes-Navon N, Levin Y, Fleming SD, Bradford BJ and Zachut M (2019) Proteomic analysis reveals greater abundance of complement and inflammatory proteins in subcutaneous adipose tissue from postpartum cows treated with sodium salicylate. *Journal of Proteomics* 204, 103399. doi: <https://doi.org/10.1016/j.jprot.2019.103399>.
- Tang Y, Bian Z, Zhao L, Liu Y, Liang S, Wang Q, Han X, Peng Y, Chen X, Shen L, Qiu D, Li Z and Ma X (2011) Interleukin-17 exacerbates hepatic steatosis and

- inflammation in non-alcoholic fatty liver disease. *Clinical and Experimental Immunology* 166, 281–290.
- Trevisi E and Bertoni G (2008) Attenuation with acetylsalicylate treatments of inflammatory conditions in periparturient dairy cows. In *Aspirin and Health Research Progress* (ed. PI Quinn), pp. 22–37. Nova Science Publishers, Inc., Hauppauge, NY, USA.
- Trevisi E, Grossi P, Cappelli FP, Cogrossi S and Bertoni G (2011) Attenuation of inflammatory response phenomena in periparturient dairy cows by the administration of an ω 3 rumen protected supplement containing Vitamin E. *Italian Journal of Animal Science* 10, 277–286.
- Tsatsanis C, Zacharioudaki V, Androulidaki A, Dermizaki E, Charalampopoulos I, Minas V, Gravanis A and Margioris AN (2005) Adiponectin induces TNF- α and IL-6 in macrophages and promotes tolerance to itself and other pro-inflammatory stimuli. *Biochemical and Biophysical Research Communications* 335, 1254–1263.
- van der Poll T, Calvano S, Kumar A, Braxton C, Coyle S, Barbosa K, Moldawer L and Lowry S (1995) Endotoxin induces downregulation of tumor necrosis factor receptors on circulating monocytes and granulocytes in humans. *Blood* 86, 2754–2759.
- Vandanmagsar B, Youm Y-H, Ravussin A, Galgani JE, Stadler K, Mynatt RL, Ravussin E, Stephens JM and Dixit VD (2011) The NLRP3 inflammasome instigates obesity-induced inflammation and insulin resistance. *Nature Medicine* 17, 179–188.
- Vanselow J, Yang W, Herrmann J, Zerbe H, Schuberth H-J, Petzl W, Tomek W and Seyfert H-M (2006) DNA-remethylation around a STAT5-binding enhancer in the α S1-casein promoter is associated with abrupt shutdown of α S1-casein synthesis during acute mastitis. *Journal of Molecular Endocrinology* 37, 463–477.
- Weisberg SP, McCann D, Desai M, Rosenbaum M, Leibel RL and Ferrante AW Jr (2003) Obesity is associated with macrophage accumulation in adipose tissue. *The Journal of Clinical Investigation* 112, 1796–1808.
- Wen H, Gris D, Lei Y, Jha S, Zhang L, Huang MT-H, Brickey WJ and Ting JPY (2011) Fatty acid-induced NLRP3-ASC inflammasome activation interferes with insulin signaling. *Nature Immunology* 12, 408–415.
- Wernstedt Asterholm I, C Tao, TS Morley, QA Wang, F Delgado-Lopez, ZV Wang and PE Scherer (2014) Adipocyte inflammation is essential for healthy adipose tissue expansion and remodeling. *Cell Metabolism* 20, 103–118.
- Winer S, Paltser G, Chan Y, Tsui H, Engleman E, Winer D and Dosch H-M (2009) Obesity predisposes to Th17 bias. *European Journal of Immunology* 39, 2629–2635.
- Wolf AM, Wolf D, Rumpold H, Enrich B and Tilg H (2004) Adiponectin induces the anti-inflammatory cytokines IL-10 and IL-1RA in human leukocytes. *Biochemical and Biophysical Research Communications* 323, 630–635.
- Wu D, Molofsky AB, Liang H-E, Ricardo-Gonzalez RR, Jouihan HA, Bando JK, Chawla A and Locksley RM (2011) Eosinophils sustain adipose alternatively activated macrophages associated with glucose homeostasis. *Science* 332, 243–247.
- Wulster-Radcliffe MC, Ajuwon KM, Wang J, Christian JA and Spurlock ME (2004) Adiponectin differentially regulates cytokines in porcine macrophages. *Biochemical and Biophysical Research Communications* 316, 924–929.
- Yamanashi T, Iwata M, Kamiya N, Tsunetomi K, Kajitani N, Wada N, Litsuka T, Yamauchi T, Miura A, Pu S, Shirayama Y, Watanabe K, Duman RS and Kaneko K (2017) Beta-hydroxybutyrate, an endogenous NLRP3 inflammasome inhibitor, attenuates stress-induced behavioral and inflammatory responses. *Scientific Reports* 7, 7677.
- Youm YH, Nguyen KY, Grant RW, Goldberg EL, Bodogai M, Kim D, D'Agostino D, Planavsky N, Lupfer C, Kanneganti TD, Kang S, Horvath TL, Fahmy TM, Crawford PA, Biragyn A, Alnemri E and Dixit VD (2015) The ketone metabolite beta-hydroxybutyrate blocks NLRP3 inflammasome-mediated inflammatory disease. *Nature Medicine* 21, 263–269.
- Yuan K, Farney JK, Mamedova LK, Sordillo LM and Bradford BJ (2013) TNF α altered inflammatory responses, impaired health and productivity, but did not affect glucose or lipid metabolism in early-lactation dairy cows. *PLoS ONE* 8, e80316.
- Zachut M (2015) Defining the adipose tissue proteome of dairy cows to reveal biomarkers related to peripartum insulin resistance and metabolic status. *Journal of Proteome Research* 14, 2863–2871.
- Zachut M, Honig H, Striem S, Zick Y, Boura-Halfon S and Moallem U (2013) Periparturient dairy cows do not exhibit hepatic insulin resistance, yet adipose-specific insulin resistance occurs in cows prone to high weight loss. *Journal of Dairy Science* 96, 5656–5669.
- Zanoni I, Ostuni R, Marek Lorri R, Barresi S, Barbalat R, Barton Gregory M, Granucci F and Kagan Jonathan C (2011) CD14 Controls the LPS-induced endocytosis of toll-like receptor 4. *Cell* 147, 868–880.
- Zarrin M, Gossen-Rösti L, Bruckmaier RM and Gross JJ (2017) Elevation of blood β -hydroxybutyrate concentration affects glucose metabolism in dairy cows before and after parturition. *Journal of Dairy Science* 100, 2323–2333.