

# Mucosal microbial parasites/symbionts in health and disease: an integrative overview

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## Editorial

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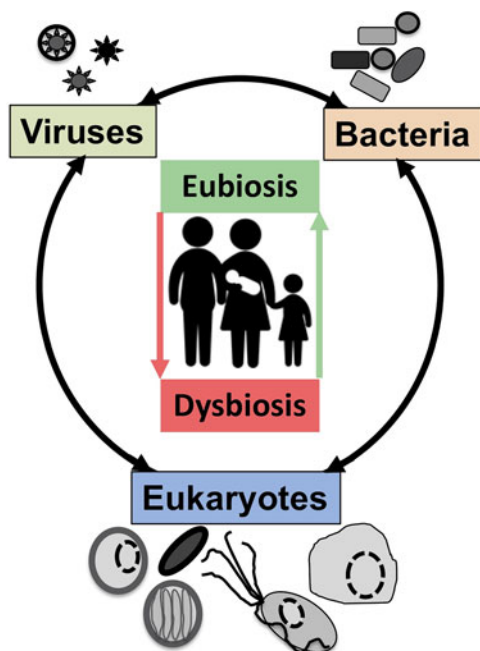
## Abstract

Microbial parasites adapted to thrive at mammalian mucosal surfaces have evolved multiple times from phylogenetically distant lineages into various extracellular and intracellular life styles. Their symbiotic relationships can range from commensalism to parasitism and more recently some host–parasites interactions are thought to have evolved into mutualistic associations too. It is increasingly appreciated that this diversity of symbiotic outcomes is the product of a complex network of parasites–microbiota–host interactions. Refinement and broader use of DNA based detection techniques are providing increasing evidence of how common some mucosal microbial parasites are and their host range, with some species being able to swap hosts, including from farm and pet animals to humans. A selection of examples will illustrate the zoonotic potential for a number of microbial parasites and how some species can be either disruptive or beneficial nodes in the complex networks of host–microbe interactions disrupting or maintaining mucosal homeostasis. It will be argued that mucosal microbial parasitic diversity will represent an important resource to help us dissect through comparative studies the role of host–microbe interactions in both human health and disease.

## Introduction

Intimately associated with the human mucosa functions and defences are the complex microbial communities, called the microbiota, which are increasingly understood to play key roles in myriads of aspects in human health and disease (Clemente *et al.*, 2012; Belkaid and Hand, 2014). Millions of years of refinements have ensured that mammalian mucosa in a state of homeostasis are effective at mediating simultaneously two conflicting and essential functions: (i) facilitate exchanges between the outside and the inside of the body to allow optimal breathing, nutrient and water uptake and reproduction and (ii) mediate protection against physical, chemical and biological insults, with the latter being mainly microbial in nature. Many of the molecules of the mucosal innate defence system are key for mediating interactions with microbes, including receptors sensing microbes and the central components of mucus (mucins and antimicrobial peptides) can be traced back to early phases of metazoans evolution (Schroder and Bosch, 2016; Bakshani *et al.*, 2018). In contrast, some of the effector molecules and cells characteristic of the human adaptive mucosal immune system represent more recent additions to the mucosal armoury against microbes with secretory IgAs, the archetypal antibody in human mucosal secretions, being only shared with reptiles and birds (Smith *et al.*, 2013). The mucosal microbiota form extraordinarily complex microbial ecosystems where bacteria, archaea, microbial eukaryotes and viruses form an intricate network of microbe–microbe and host–microbe interactions that can be broadly defined as eubiotic, associated with health, or dysbiotic, associated with disease (Petersen and Round, 2014; Levy *et al.*, 2017) (Fig. 1). Eubiotic relationships at mucosal surfaces are dependent on the functional characteristics of the microbiota community and corresponding finely tuned mucosal innate and adaptive immune responses to microbes, that together are required for harmonious, highly dynamic and continuous host–microbes interactions at mucosal surfaces (Clemente *et al.*, 2012; Belkaid and Hand, 2014; Levy *et al.*, 2017). Finely choreographed host–microbiota interactions are essential to maintain mucosal homeostasis in the broadest possible range of conditions experienced by humans, including variations in diet, exposures to various environmental microbes including pathogens and an increasing range of xenobiotics (Levy *et al.*, 2017; Ferreira *et al.*, 2018).

Mucosal microbial parasites (also referred to as parasitic protozoa or parasitic protists) are phylogenetically highly diverse and heterogenous that can be broadly distributed across human populations and can contribute to important pathologies but that are also often associated with asymptomatic interactions (Lukes *et al.*, 2015; Chabe *et al.*, 2017). Thus human–microbial parasite symbiotic relationships can range from commensalism to parasitism and more recently some host–parasites interactions are suggested to have evolved into mutualistic associations too (Lukes *et al.*, 2015; Loke and Lim, 2016; Chabe *et al.*, 2017; Stensvold, 2019). Hence these mucosa residents will be referred to here as ‘microbial eukaryote symbionts’ to better capture the diversity of symbiotic interactions mediated by organisms historically typically referred to as parasites (Lukes *et al.*, 2015; Stensvold, 2019). Notably it is increasingly appreciated that this diversity of symbiotic outcomes is the product of



**Fig. 1.** The now generally accepted new paradigm of microbe–microbe/host–microbe complex network of interactions that can contribute to health (maintaining homeostasis) or disease (inducing excessive inflammation through time and space) status of the animal/human host. The terms eubiosis and dysbiosis relate to the microbiota functional activities associated with respectively health through maintaining mucosal homeostasis (ensuring optimal mucosal functionality) or pathologies due to excess inflammation leading to damage mucosal surfaces and that can also contribute to disrupt systemic physiology and sub-optimal cognitive capacities. See main text and cited references for the conceptual limitations on the use of eubiosis and dysbiosis.

parasites–microbiota–host complex network of interactions (Clemente *et al.*, 2012; Burgess *et al.*, 2017; Rowan-Nash *et al.*, 2019), further highlighting the importance of contextuality for the phenotypic outcome of human–microbe interactions (Clemente *et al.*, 2012; Belkaid and Hand, 2014; Levy *et al.*, 2017). In this editorial, a selection of examples will illustrate how mucosal microbial parasites/symbionts (MMPS) can represent disruptive nodes of the host–microbes complex network of interactions underlying mucosal homeostasis and thus contribute directly or indirectly to mucosal dysbiosis. In contrast, other examples will illustrate the potential of microbial eukaryote symbionts to contribute to eubiosis (Stensvold and van der Giezen, 2018). With these seemingly contradictory considerations in mind, it will be argued that MMPS diversity will represent an important resource to help researchers to dissect the potential causal link between eubiosis and health and dysbiosis and disease through comparative studies. This is a research topic not without controversies and important difficulties and that will require a dramatic increase in the physiological functional characterization of members of the mucosal microbial communities (Hooks and O'Malley, 2017) including microbial eukaryotes (Chabe *et al.*, 2017; Stensvold and van der Giezen, 2018).

Several papers associated with this Special Issue are derived from talks that were delivered at the EMBO Conference ‘Anaerobic protists: Integrating parasitology with mucosal microbiota and immunology’ (<http://meetings.embo.org/event/17-anaerobic-protists>) (Labruyere *et al.*, 2017; Leitsch, 2017; Dessi *et al.*, 2019; Miranda-Ozuna *et al.*, 2019; Stensvold, 2019). These are complemented by articles providing broader perspectives on the study of the MMPS (Chabra *et al.*, 2019; Liu *et al.*, 2019; Midlej *et al.*, 2019; Bartley *et al.*, 2018; Deere *et al.*, 2018; van Gestel *et al.*, 2018; Vargas Rigo *et al.*, 2018; Chihi *et al.*,

2019; Rush *et al.*, 2019). For more in-depth coverage of MMPS biology including broader taxonomic coverage (e.g. Fungi and Helminths), mucosal sites and biology (e.g. lungs, mucus) and topics including parasite genomics, parasite diagnostics and mucosal vaccine, the reader is directed to the following reviews or original papers (Hupalo *et al.*, 2015; Serradell *et al.*, 2016; Baker *et al.*, 2017; Lemieux *et al.*, 2017; Ryan *et al.*, 2017; Collins and Belkaid, 2018; Corfield, 2018; Rowan-Nash *et al.*, 2019).

### Mucosal microbial eukaryote diversity, host range and zoonoses

What range of microbial eukaryote symbionts can thrive at our different mucosa, how broadly are they distributed across human populations and what are their host range beyond humans, how genetic diverse are they, what species/genetic lineage are associated with disease and how do these influence the mucosal microbiota and *vice versa*? These are some of the most basic questions for which we still have relatively limited knowledge for most species. This important knowledge gap currently limits us to properly assess the role in health and disease of microbial eukaryote symbionts and reflects the difficulty of studying mucosal associated organisms and viruses more generally through reductive approaches. A few examples will illustrate the importance of new perspectives one can gain from working on answering these basic questions in humans and animal models. New diagnostic technologies (Ryan *et al.*, 2017) and the increasing number of microbial eukaryote symbionts genome sequence data (Hupalo *et al.*, 2015) are all contributing at providing a better picture of the natural history of MMPS, including non-pathogenic species (Chihi *et al.*, 2019). These in combination with metagenomics surveys (Lokmer *et al.*, 2019) will generate a more comprehensive knowledge on MMPS diversity and host range and their link with health and disease.

The relatively common gut MMPS *Blastocystis* spp., *Dientamoeba fragilis* are reviewed by Stensvold (2019) (both species) and van Gestel *et al.* (2018) (*D. fragilis*). These species are thought to be common in some populations but there are a number of contradictory datasets in relation to their potential role in both disease and health and issues with the apparent important prevalence variations between populations (van Gestel *et al.*, 2018). Although potentially misleading detection tools can explain some variation between studies (van Gestel *et al.*, 2018; Gough *et al.*, 2019), a combination of environmental and biological explanations are also likely to play a role. An intriguing possibility suggested for *D. fragilis* higher prevalence in some countries is pig farming, which could potentially play a role in its higher prevalence in Denmark and the Netherlands where both humans and pigs cohabit in relatively higher densities (van Gestel *et al.*, 2018). This highlights the importance of considering both human and animal prevalence and study in detail the genetic diversity and phylogeny of the microbial eukaryote symbionts to establish their origins among humans and their potential association with animal reservoirs. This is also relevant for the relatively better known species such as *Giardia*, including in developed countries such as the UK (Horton *et al.*, 2019). A recent survey for *Giardia duodenalis* among cattle in Scotland further illustrates the importance of studying animal populations, where this species was shown to be common across surveyed beef and dairy cattle (~32%) and included genetic lineages associated with human symptomatic infections (Bartley *et al.*, 2018). In another example, vaccination to protect dogs and cats from *G. duodenalis* infections (100% prevalence) in a peri-urban disadvantaged community in Argentina, using an elegant vaccination strategy (Rivero *et al.*, 2010), was shown to reduce dog and cat

infections with the concomitant reduction of children infections in the community associated with the vaccinated pets (Serradell *et al.*, 2016). This example illustrates the importance of both the knowledge of the epidemiology of a potential pathogen and the molecular mechanisms underlying surface antigen variation to develop an effective vaccine for relevant hosts to eventually also control infections among humans. Similarly, the prevalence of *Entamoeba* spp., including *Entamoeba histolytica*, among humans, chimpanzees and baboon in the Greater Gombe Ecosystem in Tanzania, where the human and nonhuman primate populations overlap, demonstrated a high level of prevalence (~60% for all *Entamoeba* spp. and ~10% of *E. histolytica*) among all three species highlighting the potential for zoonotic transmission of *Entamoeba* species (Deere *et al.*, 2018). Notably the presence of *E. histolytica* in chimpanzees was apparently never associated with symptoms in the tested population, in contrast to human infections (Deere *et al.*, 2018).

Beyond the gut, an interesting set of data for *Trichomonas vaginalis* and *Trichomonas tenax*, infecting respectively the urogenital tract (Hirt and Sherrard, 2015) and oral cavities (Marty *et al.*, 2017) also highlight the importance of specific and sensitive diagnostics and the knowledge of their distributions beyond humans (Maritz *et al.*, 2014). Through carefully testing the specificity of a molecular diagnostic tool used for *T. vaginalis* it was discovered that some infections of the urogenital tract (three male urine samples) could be due to *T. tenax* rather than *T. vaginalis* (Brosh-Nissimov *et al.*, 2019). A screening across dogs and cats for oral trichomonads also indicated a potential zoonotic source for *T. tenax* from pets (Kellerova and Tachezy, 2017). Genotyping *T. tenax* clinical isolates from humans also established that a subset of genetic lineages are significantly associated with periodontal patients, in addition of being common among the tested population in an affluent setting (35% among patients with periodontitis and 19% among healthy controls in the studied French cohort) (Benabdelkader *et al.*, 2019). Notably both *T. vaginalis* and *T. tenax* are likely derived from species infecting birds (Maritz *et al.*, 2014) as these two species are respectively more closely related phylogenetically to a distinct set of species infecting birds including *Trichomonas gallinae*, common among pigeons, and *Trichomonas gypaetini* isolated from vultures among other *Trichomonas* spp. isolated from various bird species (Martinez-Diaz *et al.*, 2015). Transfer of *T. gallinae* from columbiform to passerines has led to important mortality rates for some passerine species dramatically illustrating the potential for a *Trichomonas* species to jump host and spread rapidly through populations and to become a virulent parasite in some contexts (wild finches such as the common chaffinch) whereas it is often a commensal in others (the columbiform rock pigeon) (Amin *et al.*, 2014). The comparative study of the molecular basis of the interactions between these various *Trichomonas* species and mucosal landmarks required to initiate and sustain the colonization of various hosts and mucosa will be of great interest and represent a fascinating model system to study MMPS transfers between birds and from birds to mammals, including humans (Maritz *et al.*, 2014).

### Symbiosis: from parasitism to commensalism to mutualism

Although a number of MMPS are known to be associated with pathologies, leading to important morbidities and mortality rates in some contexts (Bar *et al.*, 2015; Burgess *et al.*, 2017), many infections by the same species are asymptomatic (Lukes *et al.*, 2015; Chabe *et al.*, 2017; Stensvold, 2019). The outcome of host–microbial eukaryote symbiont interactions is dependent on the combination of the characteristics of the host, the microbial eukaryote and the mucosa microbiota, with increasing

evidence for an important role played by cross kingdoms interactions (Fig. 1) (Burgess *et al.*, 2017; Rowan-Nash *et al.*, 2019). Inter-kingdom interactions can modulate the inflammatory tone of the mucosa through multiple possible direct and indirect interactions between mucosal microbes, microbes and epithelial cells and microbes and immunocytes (Fig. 2). Notably the epithelial cells play key roles in both sensing microbes and orchestrating the mucosal immunological innate and adaptive responses mediated by the combination of epithelial cells and immunocytes (Fig. 2) (Petersen and Round, 2014; Levy *et al.*, 2017). Primary immunodeficiencies, due to specific genetic background interfering with epithelial cells and/or immunocytes–microbes interactions, or secondary immunodeficiencies due to infections (e.g. HIV/AIDS) or malnutrition, can dramatically increase the susceptibility of the host to numerous infections including by those of MMPS. This is particularly marked for intracellular parasites such as *Cryptosporidium* and Microsporidia, with the HIV/AIDS pandemic highlighting both the importance of the adaptive immune response in controlling these parasites and the high level of human exposure to these opportunistic intracellular pathogens from diverse zoonotic reservoirs (Stentiford *et al.*, 2016; Khan *et al.*, 2018).

In other contexts, MMPS could provide benefit to their mammalian carrier. Mice carrying the recently described gut trichomonad *Tritrichomonas musculus* were shown to be more resistant to challenges by the bacterial pathogen *Salmonella typhimurium* through enhancing mucosal defences by increasing intestinal inflammation via inflammasome activation and increase of the proinflammatory IL-18 production leading to a  $T_H1/T_H17$  immune response (Chudnovskiy *et al.*, 2016). This higher protection level to *Salmonella* was however associated with a cost as *T. musculus* colonization was also associated with a higher rate of colorectal cancer (Chudnovskiy *et al.*, 2016). This contrasts with helminths infections that tend to inhibit gut inflammation through stimulating  $T_H2/Treg$  responses (Cortes *et al.*, 2018).

These contrasting examples illustrate the importance, and potential great value, of increasing our knowledge of the natural history of mammal–MMPS interactions and the importance of studying various microbial eukaryote symbiont species in humans and animal models to dissect the complex host–MMPS–microbiota interactions to illuminate their influence in both health and disease (Loke and Lim, 2016). Additional examples of potentially beneficial microbial eukaryote symbionts, including *Entamoeba* spp. and *Blastocystis*, are discussed in this Special Issue (Stensvold, 2019) and in other contexts (Lukes *et al.*, 2015; Chabe *et al.*, 2017; Stensvold and van der Giezen, 2018) and in the next section.

### Microbial eukaryote symbionts/parasite–bacteria–virus interactions

The complex interplay between MMPS, bacteria, archaea and viruses and mammalian host health and disease status is increasingly being uncovered through the study of various microbial cellular species, bacteriophages and eukaryote infecting viruses, different mucosal surfaces and mammalian species, including humans (Clemente *et al.*, 2012; Burgess *et al.*, 2017; Chabe *et al.*, 2017; Rowan-Nash *et al.*, 2019). Here a few examples illustrating the link between these interactions and health and disease are covered with MMPS potentially contributing to either eubiosis or dysbiosis depending on the context of the hosts and their associated microbiota and environmental factors such as diet and xenobiotics (e.g. antibiotics) (Fig. 3).

Arguably one of the most fascinating and complex examples includes *Trichomonas vaginalis* that infect the urogenital tracts (UGT) of humans (Hirt and Sherrard, 2015). A complex set of



endosymbionts, the bacterial members of the UGT microbiota and how these interactions can influence the parasite pathobiology including increasing human infecting virus transmission rates. These considerations will be important to complement more traditional investigations focusing on the study of specific aspects of host–parasite interactions, such as the potential role of environmental glucose concentration variation (Miranda-Ozuna *et al.*, 2019) and cell surface and secreted factors such as exosomes (Mercer and Johnson, 2018), in modulating the virulence of the parasite. These examples illustrate dramatically the importance to investigate host–MMPS–microbiota–virus interactions in an integrative manner to develop more refined diagnostics and novel prophylactic and therapeutic strategies to eventually promote reproductive and sexual health more efficiently. It will also be of interest to investigate the possibility that related endosymbionts (to TVV and *Mycoplasma*) are also present in other *Trichomonas* species including bird infecting species and *T. tenax* associated with periodontitis (described in the previous section).

The MMPS *Giardia*, *Entamoeba* and *Cryptosporidium* are also known to be infected by RNA viruses (Gomez-Arreaza *et al.*, 2017). *Cryptosporidium*-infected virus is associated with a higher rate of the parasite propagation capacity; however, it is not clear if this increases the virulence of such infections. Similarly there is currently no evidence for *Giardia* and *Entamoeba* that their RNA viruses can contribute to boosting the pathobiology of these parasites (Gomez-Arreaza *et al.*, 2017). Complex interplay between *Giardia*, *Entamoeba* and *Cryptosporidium* with bacteria members of the microbiota have also been shown to influence the virulence of these parasites in both negative (e.g. inhibiting infections) and positive ways (e.g. promoting virulence) (Burgess *et al.*, 2017; Rowan-Nash *et al.*, 2019). A remarkable example illustrating the importance of the microbiota in playing a role in reducing the impact of *Cryptosporidium* infection was uncovered when investigating two candidate drugs to treat the parasite. Two novel drugs that had promising properties in initial *in vitro* tests had an opposite effect on *Cryptosporidium* infections in a mouse model (Gorla *et al.*, 2014). Although one of the drugs was potent in controlling the parasite, the other drug was shown to actually boost infection levels, which was associated with a significant change in the bacterial taxonomic composition of the gut microbiota, with in particular a dramatic increase of the population of the mucin loving gut bacteria *Akkermansia muciniphila* (2800-fold increase compared to the pre-treatment state), suggesting a dysbiotic microbiota (Gorla *et al.*, 2014). This was rationalized as an off-target impact of the drug on members of the gut microbiota. Although *A. muciniphila* is considered to be an important mutualist associated with human health (Cani and de Vos, 2017), the significant boost in *Cryptosporidium* infection level could be explained by an excessive degradation by *A. muciniphila* of the mucus protective layer in the gut facilitating access to, and eventual infection of, epithelial cells by *Cryptosporidium*. An apparently similar outcome was observed in a mouse model with a humanized gut microbiota fed with a diet depleted from plant fibers, which led to the depletion of the mucus protective layers by the microbiota and higher susceptibility to pathogens (Desai *et al.*, 2016). These examples illustrate how environmental factors, including xenobiotics (an antibiotic in the example above) and diet, can influence the mucosal microbial ecology and by doing so modulate the host susceptible to infections by potential pathogens, including MMPS.

### Antibiotics and vaccines for mucosal parasites/symbionts

In contrast to the availability of a broad range of antibiotic treatment regimens for bacteria, there are far less efficient options to


treat with drugs symptomatic infections due to microbial parasites (Farthing, 2006; Leitsch, 2017). As for bacteria, there is also the issue of microbial parasites developing resistance to existing drugs regimens and for off-target effects on the microbiota (Wypych and Marsland, 2018). Furthermore, some patients can develop strong reactions to some drugs including to the commonly used metronidazole targeting anaerobic parasites (Leitsch, 2017). These considerations stimulate continuous research efforts to identify new drugs to treat microbial parasites, either based on modifying existing well established drugs such as 5-nitroimidazole (Leitsch, 2017), or new drugs such as plant derived phenanthrenes (Vargas Rigo *et al.*, 2018). Irrespective of the drug, it is increasingly appreciated necessary to consider their broad impact on the host microbiota, with increasing evidence that antibiotic treatments are being associated with dysbiosis favouring opportunistic pathogens, including pathobionts, and/or leading to a difunctional immune response to microbial and other antigens that can lead to debilitating conditions such as allergies and asthma (Wypych and Marsland, 2018). In the case of the treatment of anaerobic mucosal parasites (such as *Trichomonas* and *Giardia*) with metronidazole/imidazole, the anaerobic members of the microbiota will also be affected (Leitsch, 2017). This can contribute to dysbiosis in the gut microbiota in particular where anaerobes are known to play important roles (Wypych and Marsland, 2018) (Fig. 3).

In comparison to drug treatments options, vaccines for MMPS are even less well developed. This is due to the combination of the inherent difficulties in developing effective mucosal vaccines (Lycke, 2012) and the complex biology of MMPS, including their capacity to mediate cell surface antigen variation (Deitsch *et al.*, 2009; Gargantini *et al.*, 2016) and the little knowledge we have on the nature of the host immune response to eradicate MMPS (Farthing, 2006; Chapwanya *et al.*, 2016). One promising strategy that takes advantage of the properties of VSP proteins from *Giardia* (Gargantini *et al.*, 2016) and viral-like particles has great potential to develop novel oral vaccines for various pathogens (Serradell *et al.*, 2019), including a broad range of MMPS in addition to *Giardia* (Serradell *et al.*, 2016).

### Conclusion and some speculations

From the examples covered here and in cited publications one can conclude that it might be more appropriate to refer to many extracellular microbial eukaryotic symbionts with various pathogenic potential as pathobionts that is, they are members of the mucosal microbial ecosystems that can become pathogenic in some contexts where host genetic, environment and properties of the microbial community as a whole all play a role (Chow *et al.*, 2011). Acquired immunodeficiencies or transfer of MMPS between different hosts species can lead to sub-optimal interactions with some species becoming pathogenic (Farthing, 2006; Price *et al.*, 2017). In contrast intracellular parasites, including the Apicomplexa *Cryptosporidium* and the Microsporidia, are typically thought to be primarily gut pathogens (Farthing, 2006), as they must directly exploit their host cell energy and metabolites to proceed through their life cycle and in the process compromise the integrity of the epithelial monolayer of the gut (Farthing, 2006; Dean *et al.*, 2016). One aim of this editorial was to illustrate specific aspects of the intricate and complex interactions taking place between MMPS, the other members of the microbiota and their animal or human hosts. These highlight the importance of collaborative research projects integrating parasitology, microbiology, virology, pharmacology and mucosal immunology in the context of both basic and medical and veterinary research on the factors influencing mucosa health and disease. Generating more comprehensive knowledge on the link

between these microbial interactions and mucosal and systemic health and disease is undoubtedly one of 'the most difficult and challenging scientific endeavour of our time' (Birchough and Hansson, 2017), as it will need to identify and characterize key aspects of thousands of highly dynamic interactions mediated by a complex cocktail of metabolites, cell–virus and cell–cell interactions involving complex microbial communities, epithelial cells and immunocytes. The knowledge derived from the study of these complex network of interactions will be required to eventually develop much needed novel prophylactic, including mucosal vaccines for overt pathogens, and therapeutic strategies (including highly specific drugs, prebiotics, fecal transplants), to regenerate, maintain and promote human and animal health at mucosal surfaces. It is also suggested that considering microbial eukaryote symbionts/parasites will provide important opportunities for much required comparative studies to delicately dissect key nodes orchestrating mucosal–microbes interactions and how these are causally linked to the specific phenotypic outcomes in their human and animal hosts. Contextualization of the diversity of both MMPS, the microbiota at large (bacteria, archaea and viruses) and their host within an evolutionary and ecological framework will also likely be important at helping building a more predictive theoretical framework for the outcome of host–microbes interactions (Amato, 2016; Davenport *et al.*, 2017; Rook *et al.*, 2017; Ferreiro *et al.*, 2018).

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## References

- Amato KR (2016) Incorporating the gut microbiota into models of human and non-human primate ecology and evolution. *American Journal of Physical Anthropology* **159**, S196–S215.
- Amin A, Bilic I, Liebhart D and Hess M (2014) Trichomonads in birds – a review. *Parasitology* **141**, 733–747.
- Baker JL, Bor B, Agnello M, Shi W and He X (2017) Ecology of the oral microbiome: beyond bacteria. *Trends in Microbiology* **25**, 362–374.
- Bakshani CR, Morales-Garcia AL, Althaus M, Wilcox MD, Pearson JP, Bythell JC and Burgess JG (2018) Evolutionary conservation of the antimicrobial function of mucus: a first defence against infection. *Npj Biofilms and Microbiomes* **4**, 14.
- Bar AK, Phukan N, Pinheiro J and Simoes-Barbosa A (2015) The interplay of host microbiota and parasitic protozoans at mucosal interfaces: implications for the outcomes of infections and diseases. *PLoS Neglected Tropical Diseases* **9**, e0004176.
- Bartley PM, Roehle BK, Thomson S, Shaw HJ, Peto F, Innes EA and Katzer F (2018) Detection of potentially human infectious assemblages of *Giardia duodenalis* in fecal samples from beef and dairy cattle in Scotland. *Parasitology*, 1–8. doi: 10.1017/S0031182018001117.
- Belkaid Y and Hand TW (2014) Role of the microbiota in immunity and inflammation. *Cell* **157**, 121–141.
- Benabdelkader S, Andreani J, Gillet A, Terrer E, Pignoly M, Chaudet H, Aboudharam G and La Scola B (2019) Specific clones of *Trichomonas tenax* are associated with periodontitis. *PLoS ONE* **14**, e0213338.
- Birchough G and Hansson GC (2017) Bacteria tell us how to protect our intestine. *Cell Host & Microbe* **22**, 3–4.
- Brosh-Nissimov T, Hindiyeh M, Azar R, Smollan G, Belausov N, Mandelboim M, Rahav G, Keller N and Gefen-Halevi S (2019) A false-positive *Trichomonas vaginalis* result due to *Trichomonas tenax* presence in clinical specimens may reveal a possible *T. tenax* urogenital infection. *Clinical Microbiology and Infection* **25**, 123–124.
- Burgess SL, Gilchrist CA, Lynn TC and Petri Jr WA (2017) Parasitic protozoa and interactions with the host intestinal microbiota. *Infection and Immunity* **85**, e00101-17.
- Canli PD and de Vos WM (2017) Next-generation beneficial microbes: the case of *Akkermansia muciniphila*. *Frontiers in Microbiology* **8**, 1765.
- Chabe M, Lokmer A and Segurel L (2017) Gut protozoa: friends or foes of the human gut microbiota? *Trends in Parasitology* **33**, 925–934.
- Chabra A, Rahimi-Esboei B, Habibi E, Monadi T, Azadbakht M, Elmi T, Keshavarz valian H, Akhtari J, Fakhar M and Naghshvar F (2019) Effects of some natural products from fungal and herbal sources on *Giardia lamblia* in vivo. *Parasitology* **146**, 1188–1198.
- Chapwanya A, Usman AY and Irons PC (2016) Comparative aspects of immunity and vaccination in human and bovine trichomoniasis: a review. *Tropical Animal Health and Production* **48**, 1–7.
- Chihl A, Stensvold CR, Ben-Abda I, Ben-Romdhane R, Aoun K, Siala E and Bouratbine A (2019) Development and evaluation of molecular tools for detecting and differentiating intestinal amoebae in healthy individuals. *Parasitology*, 1–7. doi: 10.1017/S0031182018002196.
- Chow J, Tang H and Mazmanian SK (2011) Pathobionts of the gastrointestinal microbiota and inflammatory disease. *Current Opinion in Immunology* **23**, 473–480.
- Chudnovskiy A, Mortha A, Kana V, Kennard A, Ramirez JD, Rahman A, Remark R, Mogno I, Ng R, Gnjjatic S, Amir ED, Solovoyov A, Greenbaum B, Clemente J, Faith J, Belkaid Y, Grigg ME and Merad M (2016) Host-protozoan interactions protect from mucosal infections through activation of the inflammasome. *Cell*, **167**, 444–456. e414.
- Clemente JC, Ursell LK, Parfrey LW and Knight R (2012) The impact of the gut microbiota on human health: an integrative view. *Cell* **148**, 1258–1270.
- Collins N and Belkaid Y (2018) Do the microbiota influence vaccines and protective immunity to pathogens? Engaging our endogenous adjuvants. *Cold Spring Harbor Perspectives in Biology* **10**, a028860.
- Corfield AP (2018) The interaction of the gut microbiota with the mucus barrier in health and disease in human. *Microorganisms* **6**, E78.
- Cortes A, Toledo R and Cantacessi C (2018) Classic models for New perspectives: delving into Helminth-microbiota-immune system interactions. *Trends in Parasitology* **34**, 640–654.
- Davenport ER, Sanders JG, Song SJ, Amato KR, Clark AG and Knight R (2017) The human microbiome in evolution. *BMC Biology* **15**, 127.
- Dean P, Hirt RP and Embley TM (2016) Microsporidia: why make nucleotides if you can steal them? *PLoS Pathogens* **12**, e1005870.
- Deere JR, Parsons MB, Lonsdorf EV, Lipende I, Kamenya S, Collins DA, Travis DA and Gillespie TR (2018) *Entamoeba histolytica* infection in humans, chimpanzees and baboons in the Greater Gorongosa Ecosystem, Tanzania. *Parasitology*, 1–7. doi: 10.1017/S0031182018001397.
- Deitsch KW, Lukehart SA and Stringer JR (2009) Common strategies for antigenic variation by bacterial, fungal and protozoan pathogens. *Nature Reviews Microbiology* **7**, 493–503.
- Desai MS, Seekatz AM, Koropatkin NM, Kamada N, Hickey CA, Wolter M, Pudlo NA, Kitamoto S, Terrapon N, Muller A, Young VB, Henrissat B, Wilmes P, Stappenbeck TS, Nunez G and Martens EC (2016) A dietary fiber-depleted gut microbiota degrades the colonic mucus barrier and enhances pathogen susceptibility. *Cell* **167**, 1339–1353. e1321.
- Dessi D, Margarita V, Cocco AR, Marongiu A, Fiori PL and Rappelli P (2019) *Trichomonas vaginalis* and *Mycoplasma hominis*: new tales of two old friends. *Parasitology*, 1–6. doi: 10.1017/S0031182018002135.
- Farthing MJ (2006) Treatment options for the eradication of intestinal protozoa. *Nature Clinical Practice. Gastroenterology & Hepatology* **3**, 436–445.
- Ferreiro A, Crook N, Gasparrini AJ and Dantas G (2018) Multiscale evolutionary dynamics of host-associated microbiomes. *Cell* **172**, 1216–1227.
- Fichorova R, Fraga J, Rappelli P and Fiori PL (2017) *Trichomonas vaginalis* infection in symbiosis with Trichomonasvirus and Mycoplasma. *Research in Microbiology* **168**, 882–891.
- Gargantini PR, Serradell MDC, Rios DN, Tenaglia AH and Lujan HD (2016) Antigenic variation in the intestinal parasite *Giardia lamblia*. *Current Opinion in Microbiology* **32**, 52–58.
- Gomez-Arreaza A, Haenni AL, Dunia I and Avilan L (2017) Viruses of parasites as actors in the parasite-host relationship: a 'menage a trois'. *Acta Tropica* **166**, 126–132.
- Gorla SK, McNair NN, Yang G, Gao S, Hu M, Jala VR, Haribabu B, Striepen B, Cuny GD, Mead JR and Hedstrom L (2014) Validation of IMP dehydrogenase inhibitors in a mouse model of cryptosporidiosis. *Antimicrobial Agents and Chemotherapy* **58**, 1603–1614.

- Gough R, Ellis J and Stark D (2019) Comparison and recommendations for the use of *Dientamoeba fragilis* real-time PCR assays. *Journal of Clinical Microbiology* **57**, e01466-18.
- Hinderfeld AS, Phukan N, Bar AK, Robertson AM and Simoes-Barbosa A (2019) Cooperative interactions between *Trichomonas vaginalis* and associated bacteria enhance paracellular permeability of the cervicovaginal epithelium by dysregulating tight junctions. *Infection and Immunity* **87**, e00141-19.
- Hirt RP and Sherrard J (2015) *Trichomonas vaginalis* origins, molecular pathobiology and clinical considerations. *Current Opinion in Infectious Diseases* **28**, 72–79.
- Hooks KB and O'Malley MA (2017) Dysbiosis and its discontents. *MBio* **8**, e01492-17.
- Horton B, Bridle H, Alexander CL and Katzer F (2019) *Giardia duodenalis* in the UK: current knowledge of risk factors and public health implications. *Parasitology* **146**, 413–424.
- Hupalo DN, Bradic M and Carlton JM (2015) The impact of genomics on population genetics of parasitic diseases. *Current Opinion in Microbiology* **23**, 49–54.
- Kellerova P and Tachezy J (2017) Zoonotic *Trichomonas tenax* and a new trichomonad species, *Trichomonas brixi* n. sp., from the oral cavities of dogs and cats. *International Journal for Parasitology* **47**, 247–255.
- Khan A, Shaik JS and Grigg ME (2018) Genomics and molecular epidemiology of *Cryptosporidium* species. *Acta Tropica* **184**, 1–14.
- Kissinger P (2015) *Trichomonas vaginalis*: a review of epidemiologic, clinical and treatment issues. *BMC Infectious Diseases* **15**, 307.
- Labruyere E, Thibeaux R, Olivo-Marin JC and Guillen N (2017) Crosstalk between *Entamoeba histolytica* and the human intestinal tract during amoebiasis. *Parasitology*, 1–10. doi: 10.1017/S0031182017002190.
- Leitsch D (2017) A review on metronidazole: an old warhorse in antimicrobial chemotherapy. *Parasitology*, 1–12. doi: 10.1017/S0031182017002025.
- Lemieux MW, Sonzogni-Desautels K and Ndao M (2017) Lessons learned from protective immune responses to optimize vaccines against cryptosporidiosis. *Pathogens (Basel, Switzerland)* **7**, E2.
- Levy M, Kolodziejczyk AA, Thaiss CA and Elinav E (2017) Dysbiosis and the immune system. *Nature Reviews Immunology* **17**, 219–232.
- Liu H, Xu N, Yin J, Yuan Z, Shen Y and Cao J (2019) Prevalence and multi-locus genotyping of potentially zoonotic *Giardia duodenalis* in pigs in Shanghai, China. *Parasitology* **146**, 1199–1205.
- Loke P and Lim YAL (2016) A commensal protozoan strikes a balance in the gut. *Cell Host & Microbe* **20**, 417–419.
- Lokmer A, Cian A, Froment A, Gantois N, Viscogliosi E, Chabe M and Segurel L (2019) Use of shotgun metagenomics for the identification of protozoa in the gut microbiota of healthy individuals from worldwide populations with various industrialization levels. *PLoS ONE* **14**, e0211139.
- Lukes J, Stensvold CR, Jirku-Pomajbikova K and Wegener Parfrey L (2015) Are human intestinal eukaryotes beneficial or commensals? *PLoS Pathogens* **11**, e1005039.
- Lycke N (2012) Recent progress in mucosal vaccine development: potential and limitations. *Nature Reviews Immunology* **12**, 592–605.
- Maritz JM, Land KM, Carlton JM and Hirt RP (2014) What is the importance of zoonotic trichomonads for human health? *Trends in Parasitology* **30**, 333–341.
- Martinez-Diaz RA, Ponce-Gordo F, Rodriguez-Arce I, del Martinez-Herrero MC, Gonzalez FG, Molina-Lopez RA and Gomez-Munoz MT (2015) *Trichomonas gypaetini* n. sp., a new trichomonad from the upper gastrointestinal tract of scavenging birds of prey. *Parasitology Research* **114**, 101–112.
- Marty M, Lemaitre M, Kemoun P, Morrier JJ and Monsarrat P (2017) *Trichomonas tenax* and periodontal diseases: a concise review. *Parasitology* **144**, 1417–1425.
- Mercer F and Johnson PJ (2018) *Trichomonas vaginalis*: pathogenesis, symbiotic interactions, and host cell immune responses. *Trends in Parasitology* **34**, 683–693.
- Miranda-Ozuna JFT, Rivera-Rivas LA, Cardenas-Guerra RE, Hernandez-Garcia MS, Rodriguez-Cruz S, Gonzalez-Robles A, Chavez-Munguia B and Arroyo R (2019) Glucose-restriction increases *Trichomonas vaginalis* cellular damage towards HeLa cells and proteolytic activity of cysteine proteinases (CPs), such as TvCP2. *Parasitology*, 1–11. doi: 10.1017/S0031182019000209.
- Petersen C and Rowan AD (2014) Defining dysbiosis and its influence on host immunity and disease. *Cellular Microbiology* **16**, 1024–1033.
- Pindak FF, Mora de Pindak M, Hyde BM and Gardner Jr WA (1989). Acquisition and retention of viruses by *Trichomonas vaginalis*. *Genitourinary Medicine* **65**, 366–371.
- Pinheiro J, Biboy J, Vollmer W, Hirt RP, Keown JR, Artuyants A, Black MM, Goldstone DC and Simoes-Barbosa A (2018) The protozoan *Trichomonas vaginalis* targets bacteria with laterally acquired NlpC/P60 peptidoglycan hydrolases. *MBio* **9**, e01784-18.
- Price LB, Hungate BA, Koch BJ, Davis GS and Liu CM (2017) Colonizing opportunistic pathogens (COPs): the beasts in all of us. *PLoS Pathogens* **13**, e1006369.
- Rendon-Maldonado J, Espinosa-Cantellano M, Soler C, Torres JV and Martinez-Palomo A (2003) *Trichomonas vaginalis*: in vitro attachment and internalization of HIV-1 and HIV-1-infected lymphocytes. *Journal of Eukaryotic Microbiology* **50**, 43–48.
- Rivero FD, Saura A, Pucca CG, Carranza PG, Torri A and Lujan HD (2010) Disruption of antigenic variation is crucial for effective parasite vaccine. *Nature Medicine* **16**, 551–557, 551p following 557.
- Rook G, Backhed F, Levin BR, McFall-Ngai MJ and McLean AR (2017) Evolution, human-microbe interactions, and life history plasticity. *Lancet* **390**, 521–530.
- Rowan-Nash AD, Korry BJ, Mylonakis E and Belenky P (2019) Cross-domain and viral interactions in the microbiome. *Microbiology and Molecular Biology Reviews* **83**, e00044-18.
- Rush G, Reynolds MW, Calvani NED and Slapeta J (2019) Addressing the constraints of *Trichomonas* foetus sample collection in remote areas: lyophilized modified Diamond's media as a substitute for liquid medium. *Parasitology*, 1–4. doi: 10.1017/S0031182019000258.
- Ryan U, Papparini A and Oskam C (2017) New technologies for detection of enteric parasites. *Trends in Parasitology* **33**, 532–546.
- Schroder K and Bosch TC (2016) The origin of mucosal immunity: lessons from the holobiont hydra. *MBio* **7**, e01184-16.
- Serradell MC, Saura A, Rupil LL, Gargantini PR, Faya MI, Furlan PJ and Lujan HD (2016) Vaccination of domestic animals with a novel oral vaccine prevents *Giardia* infections, alleviates signs of giardiasis and reduces transmission to humans. *NPJ Vaccines* **1**, 16018.
- Serradell MC, Rupil LL, Martino RA, Pucca CG, Carranza PG, Saura A, Fernandez EA, Gargantini PR, Tenaglia AH, Petiti JP, Tonelli RR, Reinoso-Vizcaino N, Echenique J, Berod L, Piaggio E, Bellier B, Sparwasser T, Klatzmann D and Lujan HD (2019) Efficient oral vaccination by bioengineering virus-like particles with protozoan surface proteins. *Nature Communications* **10**, 361.
- Smith PD, MacDonald TT and Blumberg RS (2013) Phylogeny of the mucosal immune system. In Smith PD, MacDonald TT and Blumberg RS (eds), *Principles of Mucosal Immunology*. London: Garland Science/Taylor & Francis Group, pp. 19–26.
- Stensvold CR (2019) Pinning down the role of common luminal intestinal parasitic protists in human health and disease – status and challenges. *Parasitology*, 1–7. doi: 10.1017/S0031182019000039.
- Stensvold CR and van der Giezen M (2018) Associations between gut microbiota and common luminal intestinal parasites. *Trends in Parasitology* **34**, 369–377.
- Stentiford GD, Becnel J, Weiss LM, Keeling PJ, Didier ES, Williams BP, Bjornson S, Kent ML, Freeman MA, Brown MJF, Troemel ER, Roessel K, Sokolova Y, Snowden KF and Solter L (2016) Microsporidia – emergent pathogens in the global food chain. *Trends in Parasitology* **32**, 336–348.
- Thi Trung Thu T, Margarita V, Cocco AR, Marongiu A, Dessi D, Rappelli P and Fiori PL (2018) *Trichomonas vaginalis* transports virulent *Mycoplasma hominis* and transmits the infection to human cells after Metronidazole treatment: a potential role in bacterial Invasion of fetal membranes and amniotic fluid. *Journal of Pregnancy* **2018**, 5037181.
- van Gestel RS, Kusters JG and Monkelaan JF (2018) A clinical guideline on *Dientamoeba fragilis* infections. *Parasitology* 1–9. doi: 10.1017/S0031182018001385.
- Vargas Rigo G, Petro-Silveira B, Devereux M, McCann M, Souza Dos Santos AL and Tasca T (2018) Anti-*Trichomonas vaginalis* activity of 1,10-phenanthroline-5,6-dione-based metalloids and synergistic effect with metronidazole. *Parasitology*, 1–5. doi: 10.1017/S003118201800152X.
- Victor M, Felipe Rubim, Wilmer Villarreal, Érica S. Martins-Duarte, Maribel Navarro, Wanderley de Souza and Marlene Benchimol (2019) Zinc-clotrimazole complexes are effective against *Trichomonas vaginalis*. *Parasitology* **146**, 1206–1216.
- Wypych TP and Marsland BJ (2018) Antibiotics as instigators of microbial dysbiosis: implications for asthma and allergy. *Trends in Immunology* **39**, 697–711.