

Circadian Rhythms, Feeding Patterns, and Metabolic Regulation: Implications for Critical Care

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Abstract

Endogenous biological rhythms synchronise human physiology with daily cycles of light-dark, wake-sleep, feeding-fasting. Proper circadian alignment is crucial for physiological function, reflected in the rhythmic expression of molecular clock genes in various tissues, especially in skeletal muscle. Circadian disruption, such as misaligned feeding, dysregulates metabolism and increases the risk of metabolic disorders like type 2 diabetes. Such disturbances are common in critically ill patients, especially those who rely on enteral nutrition. Whilst continuous provision of enteral nutrition is currently the most common practice in critical care, this is largely dictated by convenience rather than evidence. Conversely, some findings indicate that intermittent provision of enteral nutrition aligned with daylight may better support physiological functions and improve clinical/metabolic outcomes. However, there is a critical need for studies of skeletal muscle responses to acutely divergent feeding patterns, in addition to complementary translational research to map tissue-level physiology to whole-body and clinical outcomes.

Introduction

Endogenous biological rhythms synchronise human physiology with the daily cycles of light and dark, wakefulness and sleep, as well as feeding and fasting. This synchronisation typically aligns human behaviours such as wakefulness, activity, and feeding with the daylight hours - while sleep, rest, and fasting are aligned with nighttime ^(1; 2; 3; 4; 5; 6).

In the context of these daily fluctuations in physiological regulation, temporal eating patterns (i.e. chrononutrition) are a key consideration for metabolic health, such that asynchrony between these states (e.g. through nocturnal eating patterns) can misalign the circadian timing system, leading to impairment of physiological function, increasing the risk for developing chronic metabolic disorders ^(3; 7; 8; 9). This is a topic of growing interest in the context of critical care whereby the environmental conditions within the intensive care unit (ICU), drastically differ from free-living conditions. In particular, the current default approach of continuous provision of nutrients to patients unable to feed themselves may further exacerbate circadian misalignment in critically ill patients thereby impacting recovery and long term outcomes ⁽¹⁰⁾. Among critically ill patients who receive enteral nutrition, approximately 33% develop insulin resistance, which might be explained by endocrine

disruption and/or skeletal muscle wastage due to inappropriate or misaligned enteral nutrition delivery patterns^(11; 12; 13). Remarkably, this practice is largely driven by convenience and ease of administration, rather than being based on a robust understanding of its impact on patients' circadian rhythms and recovery.

The aim of this review is to summarise the current understanding around the importance of biological rhythmicity and feeding patterns in metabolic regulation, explore the existing evidence supporting an intermittent pattern of enteral feeding in a critical care setting, and to highlight the potential directions for future research to address the current gaps in our understanding. In doing so the review aims to set the stage for future work that can inform and optimize nutritional strategies in critical care settings.

Biological rhythms and their significance in muscle metabolism

Skeletal muscle in particular is a robustly rhythmic tissue, which may underpin the coordinated disposal, degradation and synthesis of metabolic substrates across the day^(14; 15; 16; 17; 18; 19). Skeletal muscle is responsible for a significant proportion (~40-85 %) of dietary glucose and lipid disposal and is an important reservoir of amino acids stored as protein^(20; 21; 22; 23; 24). Previous work has revealed diurnal rhythmicity in ~1000 genes in skeletal muscle including those related to glucose and lipid metabolism, as well as protein turnover^(15; 17; 25). Lipidomic analysis within the same cohort identified diurnal rhythms in lipid metabolites particularly major membrane-lipid species such as the sphingolipids that are involved in insulin signalling and insulin resistance⁽¹⁴⁾. Similarly, genes related to autophagy - a vital component of the skeletal muscle adaptive response to variable nutrient supply^(26; 27) - also exhibit a diurnal rhythm⁽¹⁷⁾.

The relative importance of such rhythms in skeletal muscle for health and function is apparent from studies utilising circadian disruption either through misalignment of environmental cues or through experimental *in vitro* and *in vivo* (i.e. animal models) disruption of the endogenous clock. Disturbance of typical rhythms in skeletal muscle compromises the lipidome and can reduce the uptake/transport, utilisation and non-oxidative storage of glucose (i.e. glycogen synthesis), thereby reducing insulin sensitivity in human skeletal muscle^(17; 28; 29; 30; 31; 32; 33). These effects heighten the risk of type 2 diabetes (T2D), which itself is characterised by blunted circadian oscillations, collectively suggesting that circadian disruption is a defining feature of the insulin resistant state^(34; 35). Loss of key clock proteins, such as *Bmal1*, leads to an accelerated sarcopenic phenotype with age in mice⁽³⁶⁾,

and impairs various aspects essential for proper muscle performance, including sarcomeric structure, mitochondrial morphology, and muscle contractile activities⁽³⁷⁾. Collectively, this evidence highlights the importance of rhythmicity within skeletal muscle for both metabolic health and function.

Clinical implications of skeletal muscle rhythms for critically ill patients

Maintaining typical circadian rhythmicity of skeletal muscle is especially important for critically ill individuals. Despite a worldwide increase in survival rates of critically ill patients, long-term outcomes for those who do survive remain poor. A significant proportion experience chronic impairment in metabolism, sleep, physical function, and cognitive and psychological health⁽³⁸⁾. These adverse outcomes can be attributed to a myriad of factors, such as muscle disuse and inflammation stemming from injury or illness, among others^(11; 12; 13). While these factors could theoretically vary based on the specific conditions and circumstances of each patient, one common element that could markedly impact all critically ill patients is circadian disruption due to the stark contrast between typical daily life and the 24-h schedule of a working ICU environment^(10; 39; 40; 41). Consequently, understanding and addressing circadian disruption could be a key aspect of improving outcomes in critically ill patients. One such practically feasible strategy for targeting circadian disruption is through the appropriate provision (e.g. amount and timing) of nutrition to critically ill patients.

Enteral feeding patterns in critically ill patients

Annually, critical care units in the UK admit approximately 200,000 patients⁽⁴²⁾ and it's estimated that between 30-50% of these patients are already malnourished at the time of their admission⁽⁴³⁾. Approximately half of admitted critically ill patients will be fed enterally, providing vital support for various conditions (e.g. palliative, post-surgical and intensive care)⁽⁴⁴⁾, because they are unable to feed themselves for a prolonged period⁽⁴⁵⁾. However, ~33% of enterally fed patients develop insulin resistance and a larger portion experience substantial muscle atrophy^(12; 46; 47). Both of these could be explained, in part, by inappropriate delivery of enteral nutrition – which may exacerbate circadian disruption, further impairing metabolism at the tissue level of skeletal muscle. There is evidently a need to consider the implications of enteral feeding patterns in critically ill patients to maintain daily rhythmicity and prevent further deterioration of metabolism.

The default and most prevalent pattern worldwide is to deliver nutrients continuously, yet this decision is based on convenience not evidence. Recent systematic reviews consistently call for research into whether an intermittent feeding pattern may be superior^(48; 49; 50). A number of studies have explored the effects of intermittent feeding in the Intensive Care Unit (ICU). These studies have not been successful in showing improvements in morbidity or mortality^(41; 51; 52). However, it is worth noting that intermittent feeding protocols often still include nighttime feeding or lack a sufficiently long fasting period, factors that could potentially undermine the potential benefits of this approach^(52; 53; 54; 55; 56; 57; 58).

Continuous feeding presents practical problems, with unscheduled interruptions for clinical procedures such that nutritional targets are unmet. Moreover, a permanently postprandial (fed) state extending throughout most, or all the sleeping phase is unlikely to be optimal for physiological function or circadian alignment. By contrast, regular bolus feedings specific to the daylight/waking phase are more aligned both with our natural eating patterns and with entrained biological rhythms in clinically relevant processes such as metabolic regulation/flexibility, protein turnover and autophagy^(41; 51; 59; 60). For instance, intermittent protein ingestion more effectively stimulates muscle protein synthesis than a continuous amino acid supply, which is an important outcome in critically ill patients to minimise the risk of muscle wastage^(61; 62; 63). Normal meal intake results in the pulsatile release of insulin and ghrelin⁽⁶⁴⁾, which is preserved with intermittent enteral feeding but lost with continuous feeding. Given that insulin is a potent modulator of clock gene and/or protein expression in multiple tissues, this pulsatile release may be necessary for maintaining rhythmicity in skeletal muscle,^(65; 66; 67; 68; 69). Interestingly, under controlled conditions the diurnal rhythm of skeletal muscle genes related to glucose, lipid, and protein metabolism are temporally related to the diurnal profile of insulin, highlighting the potential for feeding patterns to modulate and entrain skeletal muscle rhythmicity⁽²⁵⁾. Notably, shortening of the eating window through time restricted feeding (TRF) has been shown to increase the amplitude of oscillating muscle transcripts⁽⁷⁰⁾. However, neither the acute response (i.e. 24-h) or adaptation time of skeletal muscle to novel feeding patterns has been established^(41; 71).

Nonetheless, intermittent provision of enteral nutrition attenuates the progressive rise in plasma leptin and insulinemia seen with continuous feeding during bed rest⁽⁴⁷⁾ potentially enhancing splanchnic blood flow, and improve gastrointestinal tolerance of enteral nutrition while influencing skeletal muscle autophagy⁽⁷²⁾. While intermittent enteral nutrition may increase the risk of diarrhea, it can reduce the incidence of constipation, without affecting

other gastrointestinal outcomes⁽⁷³⁾. From a practical perspective, intermittent feeding offers several advantages. It imposes less limitation on patient mobility and necessitates fewer pauses for procedures or tests. It may also help achieve enteral calorie targets faster than continuous feeding in line with international guidelines emphasising the importance of providing early adequate enteral nutrition for critically ill patients^(52; 53; 54; 74; 75; 76).

In theory, intermittent feedings could sustain organ stress resistance and promote overall resilience, thus improving patient response to treatment and recovery from illness^(51; 59; 60). It is thus remarkable that the links between nutrient timing, chronobiological strain and human health outcomes remain to be established⁽⁷⁷⁾ and skeletal muscle metabolic responses have never been examined. However, in practice, improvements in morbidity and mortality are not yet observed.

Recommendations for future research

The existing body of research clearly indicates the existence of diurnal rhythms in skeletal muscle and the detrimental metabolic outcomes that can arise from disruption of these rhythms. However, a significant knowledge gap remains regarding how different feeding patterns influence these 24-hour profiles. In particular, it is now important to establish whether these rhythms occur independently from, of or are driven by, feeding pattern (i.e., whether they are driven by endogenous or exogenous clocks, respectively). Furthermore, disruption of circadian clocks as a result of enteral feeding pattern may also lead to insulin resistance, yet no studies to date have examined skeletal muscle clocks in response to divergent feeding patterns. Given the critical role of skeletal muscle in postprandial metabolic regulation^(20; 35), it is important to establish the temporal responses of this tissue to enteral nutrition delivery pattern.

In addition to furthering mechanistic understanding of divergent feeding patterns, it is important to recognise the need for translational studies to determine whether intermittent feeding with overnight fasting can produce improvements in physiological, hormonal, and metabolic responses in critically ill patients. Specifically, we need complementary studies that map tissue-level physiology onto whole-body and clinical outcomes. Given that existing studies of intermittent enteral nutrition still provide nutrition through the night studies aiming to establish the clinical feasibility, tolerability, and efficacy of intermittent *diurnal* feeding in critically ill adults would be particularly useful. Additional work should also seek

to establish the effects of intermittent enteral nutrition on long term outcomes (e.g. metabolism, sleep, physical function, and cognitive and psychological health).

Conclusion

Existing research highlights the significance of circadian rhythms in skeletal muscle metabolism and their relevance for critically ill patients. However the influence of feeding patterns (i.e. temporal variance in nutrient availability) on these rhythms remains unclear. Complementary mechanistic (i.e. in healthy adults) and clinical (i.e. in critically ill patients) studies contrasting the specific metabolic effects of intermittent and continuous nutrition are still required to improve our understanding and provide a more robust evidence base. In turn this will drive clinical practice in critically ill patients.

Author Contributions

HAS was responsible for the concept of the review. Both HAS and JAB contributed to the manuscript's design and writing. Both reviewed and approved the final version, agreeing to take responsibility for all aspects of the work, including addressing any questions regarding its accuracy or integrity. All authors meet the criteria for authorship, and only those who meet these criteria are listed as authors.

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References

1. Dibner C, Schibler U (2018) Body clocks: Time for the Nobel Prize. *Acta Physiol (Oxf)* **222**.
2. Ekmekcioglu C, Touitou Y (2011) Chronobiological aspects of food intake and metabolism and their relevance on energy balance and weight regulation. *Obesity reviews : an official journal of the International Association for the Study of Obesity* **12**, 14-25.
3. Johnston JD (2014) Physiological responses to food intake throughout the day. *Nutrition research reviews* **27**, 107-118.
4. McGinnis GR, Young ME (2016) Circadian regulation of metabolic homeostasis: causes and consequences. *Nat Sci Sleep* **8**, 163-180.
5. Longo VD, Panda S (2016) Fasting, Circadian Rhythms, and Time-Restricted Feeding in Healthy Lifespan. *Cell Metab* **23**, 1048-1059.
6. Gerhart-Hines Z, Lazar MA (2015) Circadian metabolism in the light of evolution. *Endocr Rev* **36**, 289-304.
7. Skene DJ, Skorniyakov E, Chowdhury NR *et al.* (2018) Separation of circadian- and behavior-driven metabolite rhythms in humans provides a window on peripheral oscillators and metabolism. *Proc Natl Acad Sci U S A* **115**, 7825-7830.
8. Lund J, Arendt J, Hampton SM *et al.* (2001) Postprandial hormone and metabolic responses amongst shift workers in Antarctica. *J Endocrinol* **171**, 557-564.
9. Flanagan A, Bechtold DA, Pot GK *et al.* (2021) Chrono-nutrition: From molecular and neuronal mechanisms to human epidemiology and timed feeding patterns. *J Neurochem* **157**, 53-72.

10. Kouw IWK, Heilbronn LK, van Zanten ARH (2022) Intermittent feeding and circadian rhythm in critical illness. *Curr Opin Crit Care* **28**, 381-388.
11. Woolfson AM, Saour JN, Ricketts CR *et al.* (1976) Prolonged nasogastric tube feeding in critically ill and surgical patients. *Postgrad Med J* **52**, 678-682.
12. Dirks ML, Wall BT, van de Valk B *et al.* (2016) One Week of Bed Rest Leads to Substantial Muscle Atrophy and Induces Whole-Body Insulin Resistance in the Absence of Skeletal Muscle Lipid Accumulation. *Diabetes* **65**, 2862-2875.
13. Luttikhoud J, van Norren K, Rijna H *et al.* (2016) Jejunal feeding is followed by a greater rise in plasma cholecystokinin, peptide YY, glucagon-like peptide 1, and glucagon-like peptide 2 concentrations compared with gastric feeding in vivo in humans: a randomized trial. *Am J Clin Nutr* **103**, 435-443.
14. Loizides-Mangold U, Perrin L, Vandereycken B *et al.* (2017) Lipidomics reveals diurnal lipid oscillations in human skeletal muscle persisting in cellular myotubes cultured in vitro. *Proc Natl Acad Sci U S A* **114**, E8565-E8574.
15. Perrin L, Loizides-Mangold U, Chanon S *et al.* (2018) Transcriptomic analyses reveal rhythmic and CLOCK-driven pathways in human skeletal muscle. *eLife* **7**.
16. Held NM, Wefers J, van Weeghel M *et al.* (2020) Skeletal muscle in healthy humans exhibits a day-night rhythm in lipid metabolism. *Molecular metabolism*, 100989.
17. Perrin L, Loizides-Mangold U, Chanon S *et al.* (2018) Transcriptomic analyses reveal rhythmic and CLOCK-driven pathways in human skeletal muscle. *Elife* **16**, 34114.
18. Hansen J, Timmers S, Moonen-Kornips E *et al.* (2016) Synchronized human skeletal myotubes of lean, obese and type 2 diabetic patients maintain circadian oscillation of clock genes. *Sci Rep* **6**, 35047.
19. Harmsen JF, van Weeghel M, Parsons R *et al.* (2022) Divergent remodeling of the skeletal muscle metabolome over 24 h between young, healthy men and older, metabolically compromised men. *Cell Rep* **41**, 111786.
20. DeFronzo RA, Jacot E, Jequier E *et al.* (1981) The effect of insulin on the disposal of intravenous glucose. Results from indirect calorimetry and hepatic and femoral venous catheterization. *Diabetes* **30**, 1000-1007.
21. Ferrannini E, Bjorkman O, Reichard GA, Jr. *et al.* (1985) The disposal of an oral glucose load in healthy subjects. A quantitative study. *Diabetes* **34**, 580-588.
22. Meyer C, Dostou JM, Welle SL *et al.* (2002) Role of human liver, kidney, and skeletal muscle in postprandial glucose homeostasis. *Am J Physiol* **282**, E419-427.

23. Ruge T, Hodson L, Cheeseman J *et al.* (2009) Fasted to fed trafficking of Fatty acids in human adipose tissue reveals a novel regulatory step for enhanced fat storage. *J Clin Endocrinol Metab* **94**, 1781-1788.
24. Argilés JM, Campos N, Lopez-Pedrosa JM *et al.* (2016) Skeletal Muscle Regulates Metabolism via Interorgan Crosstalk: Roles in Health and Disease. *J Am Med Dir Assoc* **17**, 789-796.
25. Smith HA, Templeman I, Davis M *et al.* (2024) Characterising 24-h skeletal muscle gene expression alongside metabolic & endocrine responses under diurnal conditions. *J Clin Endocrinol Metab*.
26. Kim KH, Lee MS (2014) Autophagy--a key player in cellular and body metabolism. *Nat Rev Endocrinol* **10**, 322-337.
27. Yin Z, Pascual C, Klionsky DJ (2016) Autophagy: machinery and regulation. *Microb Cell* **3**, 588-596.
28. Wefers J, van Moorsel D, Hansen J *et al.* (2018) Circadian misalignment induces fatty acid metabolism gene profiles and compromises insulin sensitivity in human skeletal muscle. *Proc Natl Acad Sci U S A* **115**, 7789-7794.
29. Harmsen JF, van Polanen N, van Weeghel M *et al.* (2021) Circadian misalignment disturbs the skeletal muscle lipidome in healthy young men. *Faseb j* **35**, e21611.
30. Bandet CL, Tan-Chen S, Bourron O *et al.* (2019) Sphingolipid Metabolism: New Insight into Ceramide-Induced Lipotoxicity in Muscle Cells. *Int J Mol Sci* **20**.
31. Morris CJ, Yang JN, Garcia JI *et al.* (2015) Endogenous circadian system and circadian misalignment impact glucose tolerance via separate mechanisms in humans. *Proc Natl Acad Sci U S A* **112**, E2225-2234.
32. Dyar KA, Ciciliot S, Wright LE *et al.* (2013) Muscle insulin sensitivity and glucose metabolism are controlled by the intrinsic muscle clock. *Mol Metab* **3**, 29-41.
33. Harfmann BD, Schroder EA, Kachman MT *et al.* (2016) Muscle-specific loss of Bmal1 leads to disrupted tissue glucose metabolism and systemic glucose homeostasis. *Skelet Muscle* **6**, 016-0082.
34. Gabriel BM, Altıntaş A, Smith JAB *et al.* (2021) Disrupted circadian oscillations in type 2 diabetes are linked to altered rhythmic mitochondrial metabolism in skeletal muscle. *Sci Adv* **7**, eabi9654.
35. DeFronzo RA, Tripathy D (2009) Skeletal muscle insulin resistance is the primary defect in type 2 diabetes. *Diabetes Care* **32 Suppl 2**, S157-163.

36. Kondratov RV, Kondratova AA, Gorbacheva VY *et al.* (2006) Early aging and age-related pathologies in mice deficient in BMAL1, the core component of the circadian clock. *Genes Dev* **20**, 1868-1873.
37. Andrews JL, Zhang X, McCarthy JJ *et al.* (2010) CLOCK and BMAL1 regulate MyoD and are necessary for maintenance of skeletal muscle phenotype and function. *Proc Natl Acad Sci U S A* **107**, 19090-19095.
38. Rousseau AF, Prescott HC, Brett SJ *et al.* (2021) Long-term outcomes after critical illness: recent insights. *Crit Care* **25**, 108.
39. Regmi P, Heilbronn LK (2020) Time-Restricted Eating: Benefits, Mechanisms, and Challenges in Translation. *iScience* **23**, 101161.
40. Daou M, Teliás I, Younes M *et al.* (2020) Abnormal Sleep, Circadian Rhythm Disruption, and Delirium in the ICU: Are They Related? *Front Neurol* **11**, 549908.
41. Bear DE, Hart N, Puthuchery Z (2018) Continuous or intermittent feeding: pros and cons. *Curr Opin Crit Care* **24**, 256-261.
42. Centre ICNAR (2024). <https://www.icnarc.org/> (2024)
43. Lew CCH, Yandell R, Fraser RJL *et al.* (2017) Association Between Malnutrition and Clinical Outcomes in the Intensive Care Unit: A Systematic Review [Formula: see text]. *JPEN J Parenter Enteral Nutr* **41**, 744-758.
44. Wang K, McIlroy K, Plank LD *et al.* (2017) Prevalence, Outcomes, and Management of Enteral Tube Feeding Intolerance: A Retrospective Cohort Study in a Tertiary Center. *JPEN J Parenter Enteral Nutr* **41**, 959-967.
45. Binnekade JM, Tepaske R, Bruynzeel P *et al.* (2005) Daily enteral feeding practice on the ICU: attainment of goals and interfering factors. *Crit Care* **9**, R218-225.
46. Dirks ML, Hansen D, Van Assche A *et al.* (2015) Neuromuscular electrical stimulation prevents muscle wasting in critically ill comatose patients. *Clin Sci (Lond)* **128**, 357-365.
47. Gonzalez JT, Dirks ML, Holwerda AM *et al.* (2020) Intermittent versus continuous enteral nutrition attenuates increases in insulin and leptin during short-term bed rest. *Eur J Appl Physiol* **120**, 2083-2094.
48. Ichimaru S (2018) Methods of Enteral Nutrition Administration in Critically Ill Patients: Continuous, Cyclic, Intermittent, and Bolus Feeding. *Nutr Clin Pract* **33**, 790-795.
49. Di Girolamo FG, Situlin R, Fiotti N *et al.* (2017) Intermittent vs. continuous enteral feeding to prevent catabolism in acutely ill adult and pediatric patients. *Curr Opin Clin Nutr Metab Care* **20**, 390-395.

50. Patel JJ, Rosenthal MD, Heyland DK (2018) Intermittent versus continuous feeding in critically ill adults. *Curr Opin Clin Nutr Metab Care* **21**, 116-120.
51. Van Dyck L, Casaer MP (2019) Intermittent or continuous feeding: any difference during the first week? *Curr Opin Crit Care* **25**, 356-362.
52. McNelly AS, Bear DE, Connolly BA *et al.* (2020) Effect of Intermittent or Continuous Feed on Muscle Wasting in Critical Illness: A Phase 2 Clinical Trial. *Chest* **158**, 183-194.
53. Kadamani I, Itani M, Zahran E *et al.* (2014) Incidence of aspiration and gastrointestinal complications in critically ill patients using continuous versus bolus infusion of enteral nutrition: a pseudo-randomised controlled trial. *Aust Crit Care* **27**, 188-193.
54. MacLeod JB, Lefton J, Houghton D *et al.* (2007) Prospective randomized control trial of intermittent versus continuous gastric feeds for critically ill trauma patients. *J Trauma* **63**, 57-61.
55. Nasiri M, Farsi Z, Ahangari M *et al.* (2017) Comparison of Intermittent and Bolus Enteral Feeding Methods on Enteral Feeding Intolerance of Patients with Sepsis: A Triple-blind Controlled Trial in Intensive Care Units. *Middle East J Dig Dis* **9**, 218-227.
56. Rhoney DH, Parker D, Jr., Formea CM *et al.* (2002) Tolerability of bolus versus continuous gastric feeding in brain-injured patients. *Neurol Res* **24**, 613-620.
57. Serpa LF, Kimura M, Faintuch J *et al.* (2003) Effects of continuous versus bolus infusion of enteral nutrition in critical patients. *Rev Hosp Clin Fac Med Sao Paulo* **58**, 9-14.
58. Steevens EC, Lipscomb AF, Poole GV *et al.* (2002) Comparison of continuous vs intermittent nasogastric enteral feeding in trauma patients: perceptions and practice. *Nutr Clin Pract* **17**, 118-122.
59. de Cabo R, Mattson MP (2019) Effects of Intermittent Fasting on Health, Aging, and Disease. *N Engl J Med* **381**, 2541-2551.
60. Thorburn A (2018) Autophagy and disease. *J Biol Chem* **293**, 5425-5430.
61. Atherton PJ, Etheridge T, Watt PW *et al.* (2010) Muscle full effect after oral protein: time-dependent concordance and discordance between human muscle protein synthesis and mTORC1 signaling. *Am J Clin Nutr* **92**, 1080-1088.
62. Davis TA, Fiorotto ML, Suryawan A (2015) Bolus vs. continuous feeding to optimize anabolism in neonates. *Curr Opin Clin Nutr Metab Care* **18**, 102-108.
63. Zaromskyte G, Prokopidis K, Ioannidis T *et al.* (2021) Evaluating the Leucine Trigger Hypothesis to Explain the Post-prandial Regulation of Muscle Protein Synthesis in Young and Older Adults: A Systematic Review. *Front Nutr* **8**, 685165.

64. Cummings DE, Purnell JQ, Frayo RS *et al.* (2001) A preprandial rise in plasma ghrelin levels suggests a role in meal initiation in humans. *Diabetes* **50**, 1714-1719.
65. Asher G, Gatfield D, Stratmann M *et al.* (2008) SIRT1 regulates circadian clock gene expression through PER2 deacetylation. *Cell* **134**, 317-328.
66. Crosby P, Hamnett R, Putker M *et al.* (2019) Insulin/IGF-1 Drives PERIOD Synthesis to Entrain Circadian Rhythms with Feeding Time. *Cell* **177**, 896-909.e820.
67. Mukherji A, Kobiita A, Chambon P (2015) Shifting the feeding of mice to the rest phase creates metabolic alterations, which, on their own, shift the peripheral circadian clocks by 12 hours. *Proc Natl Acad Sci U S A* **112**, E6683-6690.
68. Sun X, Dang F, Zhang D *et al.* (2015) Glucagon-CREB/CRTC2 signaling cascade regulates hepatic BMAL1 protein. *J Biol Chem* **290**, 2189-2197.
69. Tuvia N, Pivovarova-Ramich O, Murahovschi V *et al.* (2021) Insulin Directly Regulates the Circadian Clock in Adipose Tissue. *Diabetes* **70**, 1985-1999.
70. Lundell LS, Parr EB, Devlin BL *et al.* (2020) Time-restricted feeding alters lipid and amino acid metabolite rhythmicity without perturbing clock gene expression. *Nat Commun* **11**, 4643.
71. Johnston JD, Ordovás JM, Scheer FA *et al.* (2016) Circadian Rhythms, Metabolism, and Chrononutrition in Rodents and Humans. *Adv Nutr* **7**, 399-406.
72. Chowdhury AH, Murray K, Hoad CL *et al.* (2016) Effects of Bolus and Continuous Nasogastric Feeding on Gastric Emptying, Small Bowel Water Content, Superior Mesenteric Artery Blood Flow, and Plasma Hormone Concentrations in Healthy Adults: A Randomized Crossover Study. *Ann Surg* **263**, 450-457.
73. Heffernan AJ, Talekar C, Henain M *et al.* (2022) Comparison of continuous versus intermittent enteral feeding in critically ill patients: a systematic review and meta-analysis. *Crit Care* **26**, 325.
74. Arabi YM, Casaer MP, Chapman M *et al.* (2017) The intensive care medicine research agenda in nutrition and metabolism. *Intensive Care Med* **43**, 1239-1256.
75. Compher C, Bingham AL, McCall M *et al.* (2022) Guidelines for the provision of nutrition support therapy in the adult critically ill patient: The American Society for Parenteral and Enteral Nutrition. *JPEN J Parenter Enteral Nutr* **46**, 12-41.
76. Singer P, Blaser AR, Berger MM *et al.* (2019) ESPEN guideline on clinical nutrition in the intensive care unit. *Clin Nutr* **38**, 48-79.
77. Lewis P, Oster H, Korf HW *et al.* (2020) Food as a circadian time cue - evidence from human studies. *Nat Rev Endocrinol* **16**, 213-223.