



Impact of meal fatty acid composition on postprandial lipaemia, vascular function and blood pressure in postmenopausal women

Kumari M. Rathnayake^{1,2}, Michelle Weech¹, Kim G. Jackson¹ and Julie A. Lovegrove^{1*}

¹Hugh Sinclair Unit of Human Nutrition, Department of Food & Nutritional Sciences and Institute for Cardiovascular and Metabolic Research, University of Reading, Reading RG6 6AP, UK

²Department of Applied Nutrition, Faculty of Livestock, Fisheries and Nutrition, Wayamba University of Sri Lanka, Makandura, 60170, Sri Lanka

Abstract

CVD are the leading cause of death in women globally, with ageing associated with progressive endothelial dysfunction and increased CVD risk. Natural menopause is characterised by raised non-fasting TAG concentrations and impairment of vascular function compared with premenopausal women. However, the mechanisms underlying the increased CVD risk after women have transitioned through the menopause are unclear. Dietary fat is an important modifiable risk factor relating to both postprandial lipaemia and vascular reactivity. Meals rich in SFA and MUFA are often associated with greater postprandial TAG responses compared with those containing *n*-6 PUFA, but studies comparing their effects on vascular function during the postprandial phase are limited, particularly in postmenopausal women. The present review aimed to evaluate the acute effects of test meals rich in SFA, MUFA and *n*-6 PUFA on postprandial lipaemia, vascular reactivity and other CVD risk factors in postmenopausal women. The systematic search of the literature identified 778 publications. The impact of fat-rich meals on postprandial lipaemia was reported in seven relevant studies, of which meal fat composition was compared in one study described in three papers. An additional study determined the impact of a high-fat meal on vascular reactivity. Although moderately consistent evidence suggests detrimental effects of high-fat meals on postprandial lipaemia in postmenopausal (than premenopausal) women, there is insufficient evidence to establish the impact of meals of differing fat composition. Furthermore, there is no robust evidence to conclude the effect of meal fatty acids on vascular function or blood pressure. In conclusion, there is an urgent requirement for suitably powered robust randomised controlled trials to investigate the impact of meal fat composition on postprandial novel and established CVD risk markers in postmenopausal women, an understudied population at increased cardiometabolic risk.

Key words: Fatty acids: Postprandial lipaemia: Vascular function: Blood pressure: Postmenopausal women

Introduction

CVD, which include CHD (myocardial infarction and angina), stroke and peripheral vascular disease⁽¹⁾, are a key contributor to the burden of disease globally⁽²⁾. Over the past 50 years, the prevalence of CVD has fallen in Western populations; however, CVD are currently the major cause of death in women in the UK, accounting for 32% of all deaths⁽³⁾. Furthermore, the prevalence of CVD is dramatically increasing in other areas, including Eastern Europe, Asia and the Indian subcontinent⁽⁴⁾.

The aetiology for CVD is multifactorial and includes several modifiable risk factors, such as cigarette smoking, a sedentary lifestyle, obesity, elevated blood pressure, dyslipidaemia, type 2 diabetes mellitus, and non-modifiable factors, such as advancing age, sex, family history of heart disease and ethnicity^(5,6). Among the non-modifiable risk factors, ageing is associated with progressive endothelial dysfunction (characterised by a loss of vascular wall homeostasis leading to a decrease in vascular reactivity and raised blood pressure) in both sexes,

although it appears to occur earlier in men than women⁽⁷⁾. The most prominent sex-related difference in physiological ageing is the menopause (cessation of menstruation) in women, which usually occurs between the ages of 45 and 55 years, with 51 years being the average age of menopause in the UK⁽⁸⁾. This natural part of ageing in women contributes a significant cardiovascular milestone in terms of both physiology and pathology since oestrogen deficiency is known to impair lipid metabolism and endothelial function, and the menopause is a recognised risk factor for CVD⁽⁹⁾. It has further been shown by van der Schouw *et al.*⁽¹⁰⁾ that for each year of delay in the age of onset of the natural menopause, CVD risk falls by 2%.

Diet is one of the most important modifiable risk factors in relation to CVD⁽¹¹⁾. As a strategy to reduce the incidence of CVD, public health policy makers recommend that intakes of dietary SFA are reduced to <10% of total energy in the UK⁽¹²⁾. Substituting SFA with MUFA or PUFA may provide additional benefits in relation to CVD risk factors, including reductions in

Abbreviations: ACE, angiotensin-converting enzyme; CM, chylomicron; FMD, flow-mediated dilatation; HDL-C, HDL-cholesterol; LDL-C, LDL-cholesterol; LPL, lipoprotein lipase; RAS, renin-angiotensin system; RCT, randomised controlled trial; TRL, TAG-rich lipoprotein.

* **Corresponding author:** Professor J. A. Lovegrove, fax +44 118 3787708, email j.a.lovegrove@reading.ac.uk

the fasting lipid profile and improvements in endothelial function. A systematic review proposed that lowering dietary SFA intake by modifying dietary fat composition, rather than reducing total fat intake, may reduce cardiovascular events by 14%⁽¹³⁾. Since individuals spend a large proportion of the day in the fed (postprandial) state, modifications to the fatty acid composition of our meals that are repeated on a daily basis may have a significant impact on postprandial lipaemia and vascular health, which over time could affect CVD risk. Therefore, the aim was to systematically review and critically evaluate the existing evidence from acute studies comparing meals rich in SFA, MUFA and *n*-6 PUFA on postprandial lipaemia, vascular reactivity, blood pressure and biomarkers of vascular function and inflammation. We chose to specifically focus on postmenopausal women since they represent an understudied group within the population at increased CVD risk.

Before presentation of the methodology and results of the literature review, we provide a general overview of postprandial lipaemia and vascular function, and describe the individual impact of the menopause and dietary fat composition on these CVD risk factors.

Postprandial lipaemia

With the pattern of meal ingestion in Western societies, a greater part of the day is spent in the postprandial than fasting state. Kolovou *et al.*⁽¹⁴⁾ defined postprandial lipaemia as a complex syndrome characterised by non-fasting hypertriglycerolaemia and its augmentation is associated with an increased risk of cardiovascular events. Following a fat-containing meal, there is a transient rise in circulating TAG-rich lipoproteins (TRL), such as chylomicrons (CM) and VLDL. After entering the circulation, the CM-TAG is hydrolysed into NEFA by lipoprotein lipase (LPL), forming cholesteryl ester-rich CM remnants, which are cleared by the liver via receptor-mediated uptake. VLDL follows a similar route of metabolism in the circulation as CM particles, although VLDL are hydrolysed at a slower rate as the larger CM are the preferential substrate for LPL. VLDL TAG depletion produces smaller VLDL (intermediate-density lipoprotein or VLDL remnants), a proportion of which are metabolised to form LDL before they are cleared via hepatic LDL receptors using apo B-100 as a ligand. During the postprandial period, there is an accumulation of TRL in the circulation due to competition between intestinal and hepatic TRL for the same lipolytic and receptor-mediated uptake⁽¹⁵⁾. A delayed clearance of TRL in the circulation enhances the accumulation of TRL particles carrying acceptor sites for the cholesteryl ester transfer protein, which transfers TAG from TRL (CM and VLDL) and exchanges it with cholesteryl esters from HDL and LDL. Remodelling of the lipid content of the LDL and HDL particles make them suitable substrates for LPL and hepatic lipase, leading to the formation of smaller and denser LDL (LDL₃) and HDL (HDL₃) particles⁽¹⁶⁾. HDL₃ is rapidly removed from the circulation, decreasing circulating HDL-cholesterol (HDL-C) concentrations, which is one proposed mechanism for the inverse association between exaggerated postprandial lipaemia and CVD risk⁽¹⁷⁾. Another possible mechanism is that LDL₃ has a lower binding affinity for the LDL receptor, reducing

their rate of clearance from the circulation and enabling them to infiltrate the arterial wall⁽¹⁶⁾.

Since atherosclerosis is now also considered to be a postprandial phenomenon, three large prospective cohort studies aimed to determine the link between cardiovascular events and non-fasting TAG^(18–20). In the Norwegian Counties Study, hazard ratios of 1.2 and 1.03 for deaths from CVD per 1 mmol/l increase in non-fasting TAG were reported in women and men, respectively, after 27 years of follow-up in a total of 86 261 participants⁽²⁰⁾. Furthermore, the Copenhagen City Heart Study that followed 7581 women and 6391 men for 31 years showed that relative to women with non-fasting TAG of <1 mmol/l, hazard ratios for myocardial infarction ranged from 1.5 for women with TAG between 1.0 and 1.99 mmol/l rising to 4.2 for those with TAG \geq 5 mmol/l⁽¹⁹⁾. However, the corresponding hazard ratios for men were lower at 1.3 and 2.1, respectively. In the Women's Health Study, fasting (*n* 20 118) and non-fasting (*n* 6391) TAG predicted cardiovascular events after 11.4 years of follow-up after adjusting for age, blood pressure, smoking status and hormone therapy. The authors also reported that the strongest association between cardiovascular events and non-fasting TAG occurred 2–4 h after the last meal, with the association declining as the fasting time increased⁽¹⁸⁾. These studies have demonstrated the greater importance of non-fasting than fasting TAG concentrations as a predictor of CVD risk in women compared with men.

The impact of menopausal status on the variability of the postprandial lipaemic responses has been reported in a number of studies^(21–24) (online Supplementary Table S1). In general, premenopausal women have lower postprandial TAG responses than men^(25–28), which is in contrast to the higher reported responses observed in postmenopausal women compared with men of a similar age⁽²⁹⁾. In response to a single oral vitamin A fat loading test, van Beek *et al.*⁽²¹⁾ investigated whether a natural menopause was associated with reduced protection from exaggerated postprandial lipaemia. Higher concentrations of postprandial plasma TAG and retinyl palmitate (an indirect marker of CM) were observed in postmenopausal compared with premenopausal women of similar age, BMI, daily energy and fat intake, *APOE* genotype, LPL activity, and HDL-C concentration, even after adjusting for the confounding effect of fasting TAG. Relative to premenopausal women, Masding *et al.*⁽²³⁾, Schoppen *et al.*⁽²²⁾ and Jackson *et al.*⁽²⁴⁾ also reported significantly higher postprandial TAG responses after single and sequential fat-rich test meals in healthy postmenopausal women. Although raised LDL-cholesterol (LDL-C) is an established risk factor for CVD, large prospective studies have shown non-fasting TAG to be a better predictor of CVD risk in women than fasting LDL-C^(30–32). *Post hoc* analysis of the Dietary Studies: Reading Unilever Postprandial Trials (DISRUPT) menopausal groups according to age also revealed a greater increase in non-fasting TAG than fasting LDL-C concentrations during the late premenopausal period, suggesting that age and the menopause have a differential impact on these two lipid CVD risk biomarkers⁽²⁴⁾.

A major biochemical change that occurs in women after the menopause is a reduction in the secretion of endogenous oestrogen and progesterone⁽³³⁾. These hormones not only play a major role in sexual physiology, but are also involved in

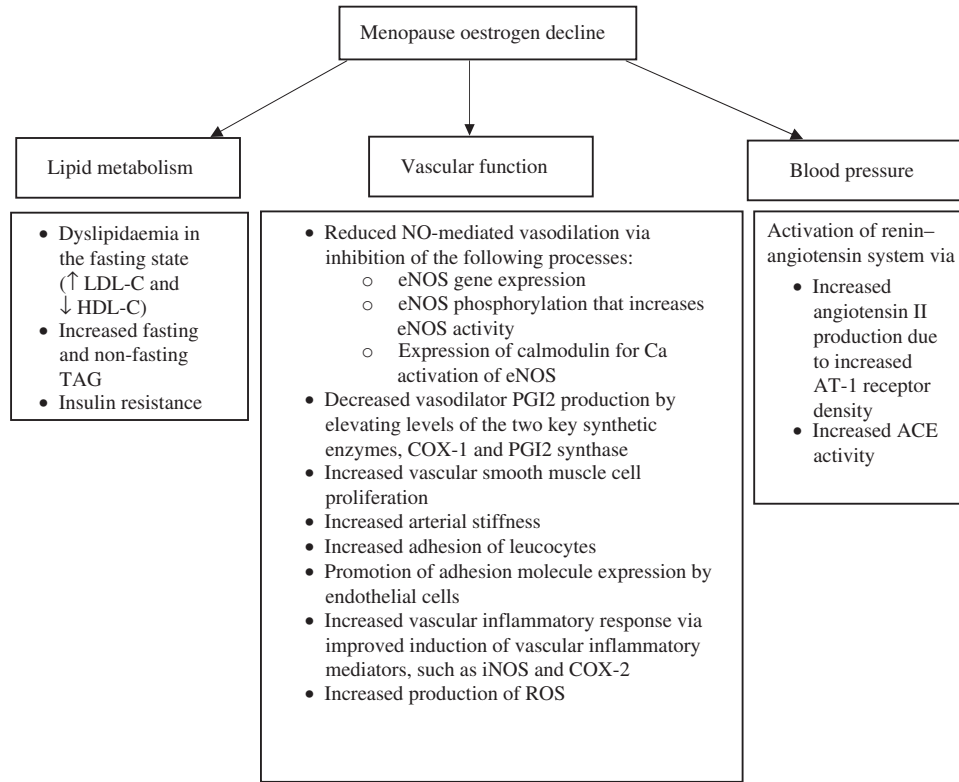


Fig. 1. Consequences of the decline in oestrogen during the menopause on lipid metabolism, vascular function and blood pressure. Adapted from Davis *et al.*⁽¹⁰⁶⁾. ACE, angiotensin converting enzyme; AT-1; angiotensin 1 receptor; COX, cyclo-oxygenase; eNOS, endothelial NO synthase; HDL-C, HDL-cholesterol; iNOS, inducible NO synthase; LDL-C, LDL-cholesterol; ROS, reactive oxygen species.

various physiological processes associated with the vasculature and lipid metabolism. A reduction in oestrogen following the menopause is associated with a detrimental impact on lipoprotein metabolism, vascular reactivity and blood pressure (Fig. 1). For example, there is much evidence to suggest that oestrogen (endogenous and exogenous) lowers fasting plasma concentrations of total and LDL-C, lipoprotein(a) and apo B, whilst elevating HDL-C, apo AI and apo AII^(34–36). The effects of oestradiol (the predominant type of oestrogen) on lipid metabolism is reported to contribute 25% of its protective effects on the fasting lipid profile⁽³⁷⁾. One possible mechanism to explain this effect, that was identified in *in vitro* animal studies, was an increase in the number of high-affinity LDL-receptors on liver cell membranes that enhance LDL uptake by the liver⁽³⁶⁾. Furthermore, the administration of even short-term (2–6 weeks) oestradiol therapy reduces the menopause-related rise in postprandial TAG in postmenopausal women^(38,39). These findings indicate that 17 β -oestradiol may accelerate the postprandial clearance of TRL and have a beneficial effect on postprandial lipaemia.

Vascular function and blood pressure

Vascular function is a measure of cardiovascular health. The components of impaired vascular function, including hypertension^(40,41), arterial stiffness⁽⁴²⁾ and impaired endothelial dependent vasodilation (endothelial dysfunction)^(43,44), are all associated with cardiovascular mortality. In a healthy blood vessel, the

endothelium, which is comprised of a monolayer of endothelial cells that lines the blood vessel walls, regulates vascular wall homeostasis by immediately responding to blood-borne and locally produced stimuli to regulate blood flow, blood pressure and vascular tone. It does so by maintaining a precise balance between the release of endothelium-derived vasodilators (such as NO) and vasoconstrictors (such as endothelin-I), which actively regulates vascular permeability to plasma constituents, platelets and leucocyte adhesion molecules⁽⁴⁵⁾ as well as aggregation and thrombosis⁽⁴⁶⁾. However, when the synthesis or bioavailability of NO is reduced, the resulting imbalance of these vasoactive substances disrupts vascular homeostasis. This 'endothelial dysfunction' is characterised by vasoconstriction, increased expression of adhesion molecules and pro-inflammatory cytokines, platelet activation and increased oxidative stress⁽⁴⁷⁾, and is becoming increasingly recognised as an important step for the initiation of coronary atherosclerosis⁽⁴⁸⁾ and increased CVD risk in postmenopausal women⁽⁴⁹⁾.

There are a number of non-invasive methods that are used to evaluate endothelial function⁽⁵⁰⁾. Flow-mediated dilatation (FMD) is the 'gold standard' technique that uses ultrasound to assess endothelium-dependent vasodilation in the conduit arteries in the peripheral circulation and is used as a surrogate measure of NO production⁽⁵¹⁾. It is now recognised as a screening tool to assess future CVD risk^(43,49,52,53). Rossi *et al.* reported that postmenopausal women in the lowest tertile of percentage FMD response (reflective of impaired vascular reactivity) had the greatest relative risk of cardiovascular events.

Furthermore, it has been shown that endothelial function is impaired across the stages of the menopause transition in healthy women with the highest percentage FMD response reported in premenopausal women, with a progressive decline in perimenopausal and postmenopausal women, respectively⁽⁵⁴⁾. These findings suggest that the perimenopausal stage (the transition towards the menopause where oestrogen production starts to fall) is a crucial turning point in women where changes in CVD risk commence.

Majmudar *et al.*⁽⁵⁵⁾ revealed that menopausal status is associated with reduced NO activity, which is restored with oestrogen replacement therapy and may be an important mechanism facilitating the detrimental effect of the menopause on CVD risk and mortality. Another study that acutely administered oestrogen (17 β -oestradiol) to postmenopausal women demonstrated protective effects on forearm microvascular responses to the endothelium-dependent vasodilator acetylcholine via improvements in NO activity⁽⁵⁶⁾. Impaired blood flow in the microcirculation has been proposed to be an indicator of initial endothelial damage in individuals at risk of CVD⁽⁵⁷⁾. Furthermore, it has been repeatedly shown that 17 β -oestradiol stimulates the production of vasodilatory prostaglandins, such as prostacyclin (PGI₂)^(58,59). These vascular effects are believed to be partly responsible for the long-term benefit of oestrogen therapy on cardiovascular risk in postmenopausal women. However, findings from the Women's Health Initiative study have questioned the benefits of oestrogen therapy. After a short (6–8 years) or longer-term (18 years) follow-up relative to a placebo, oestrogen therapy did not protect against myocardial infarction or coronary death, although the findings did show a lower CHD risk among the younger postmenopausal women (50–59 years)^(60,61). More recently, a systematic review involving 43 637 women reported the number of cardiovascular events to increase following the long-term (>1 year) use of oestrogen therapy⁽⁶²⁾, whereas a meta-analysis of peri- and post-menopausal women (*n* 40 048; aged 53–79 years) showed a risk reduction for fractures and diabetes but no significant impact on risk of CHD when oestrogen therapy was compared with a placebo⁽⁶³⁾. In contrast, there is much evidence to suggest that oestrogens (endogenous and exogenous) have several cardioprotective effects (Fig. 1)^(35,64,65). These include reductions in plasma markers of endothelial activation (E-selectin), fibrinogen, plasminogen activator inhibitor type 1 and tissue plasminogen activator antigen^(55,66). However, increases in plasma markers of inflammation (C-reactive protein) and hypercoagulability have also been reported^(35,64).

Hypertension (high blood pressure) is one of the main age-related disorders in postmenopausal women^(67,68), which has been identified as a leading risk factor for myocardial infarction and stroke in women⁽⁶⁹⁾. The renin–angiotensin system (RAS) is a hormonal cascade that plays a key role in the regulation of fluid and electrolyte balance and arterial blood pressure. Upon activation of the RAS cascade, angiotensin II is produced in the liver by angiotensin-converting enzyme (ACE) following conversion of angiotensin I to angiotensin II⁽⁷⁰⁾. Angiotensin II is a potent vasoconstrictor which degrades bradykinin (a vasodilator) causing arterioles to constrict, resulting in increased blood pressure⁽⁷¹⁾. It is well documented in the literature that

oestrogen acts on RAS at different points of the cascade, including the inhibition of ACE activity. *In vitro* and *in vivo* animal studies have also demonstrated the potential effects of oestrogen on the endothelial-dependent vasodilator response to acetylcholine in coronary and uterine arteries^(72–74). Loss of oestrogen-dependent cardiovascular protection induces endothelial dysfunction, and may also be involved in the activation of the RAS cascade. Evidence from both clinical and animal studies has shown an inverse association between oestrogen and the activation of RAS^(75–78). This has been proposed to occur due to oestrogen-induced down-regulation of angiotensin receptor I expression leading to an augmented level of angiotensin II⁽⁷⁶⁾. This is a major component of the RAS system and has several harmful effects on the vascular wall including vasoconstriction, vascular smooth muscle cell proliferation, reactive oxygen species generation and endothelial cell apoptosis^(79–81). Oestrogen deficiency has also been reported to lead to an up-regulation of ACE activity causing an accumulation of angiotensin II⁽⁸²⁾.

Impact of meal fat composition on postprandial lipaemia and vascular function

The chronic effects of substitution of SFA with PUFA on fasting lipid levels have been extensively studied⁽⁸³⁾, however, the acute effects are less well known. One systematic review and meta-analysis of randomised controlled trials (RCT) compared the effects of oral fat tolerance tests with differing fatty acid compositions on postprandial TAG responses in men and women⁽⁸⁴⁾. Relative to a single SFA-rich meal challenge, a PUFA-rich meal significantly reduced the postprandial lipaemic response over 8 h, whereas a trend for a reduced response was identified following a MUFA-rich meal. However, differences were not evident at 4 h, suggesting that a longer follow-up time after the test meal (i.e. 8 h) is preferable to observe the acute effects of meal fat composition on postprandial lipaemia. Of the eighteen studies included in the review by Monfort-Pires *et al.*⁽⁸⁴⁾, none included postmenopausal women, which reflects the paucity of postprandial data in this population subgroup.

With regards to vascular function, West⁽⁸⁵⁾ suggested that consumption of a single high-fat meal (50–105 g of fat) can impair postprandial FMD by 45 to 80 %, which appears to occur within 2 to 5 h after a high-fat meal^(86–89). Prolonged postprandial lipaemia is known to induce endothelial dysfunction by promoting the formation of free radicals by accelerating the rate of β -oxidation of NEFA (for example, superoxide radicals). Increased production of reactive oxygen species or free radicals reduces the amount of bioactive NO by chemical inactivation to form toxic peroxynitrite⁽⁹⁰⁾. In addition, it has been shown that persisting oxidative stress will render endothelial NO synthase dysfunctional, markedly reducing NO production⁽⁹¹⁾. Furthermore, high concentrations of TRL during the postprandial state enhance inflammation by inducing the secretion of pro-inflammatory cytokines⁽⁹²⁾ and expression of soluble cell adhesion molecules⁽⁹³⁾.

Reviews by Hall⁽⁹⁴⁾ and Vafeiadou *et al.*⁽⁹⁵⁾ stated that the acute effects of dietary fats on vascular function are less researched. The authors concluded that high-fat meals have a

detrimental effect on postprandial vascular function and that there is limited and inconclusive evidence for the comparative effects of test meals rich in MUFA or *n*-6 PUFA with SFA. Of note, the data derived from these reviews were mainly from studies where the effects of a single high-fat meal on postprandial vascular function in different subject groups were determined; however, none of the studies identified included postmenopausal women only.

Methods

A systematic approach was used to identify all relevant published literature according to the method used by Vafeiadou *et al.*⁽⁹⁵⁾. The PubMed (<http://www.ncbi.nlm.nih.gov/pubmed/>) database was used to perform the literature search, which included all studies published in English until October 2016. A protocol that included search terms to conduct the literature search was prepared by two authors (K. M. R. and M. W.), which was agreed by all authors. Three categories of search terms were identified: (i) study group search term (postmenopausal or postmenopausal or post-menopause or menopause or menopausal); (ii) exposure search terms (which included descriptors of SFA, MUFA and *n*-6 PUFA, and relevant food sources, for example, butter, safflower oil and olive oil); and (iii) outcomes (which included descriptors of vascular function, blood pressure, biomarkers of vascular function and inflammation, and plasma lipids) (Supplementary

information). The Medical Subject Heading Browser (<http://www.nlm.nih.gov/mesh/MBrowser.html>) was used to identify relevant exposures and outcomes. Two additional studies were identified through hand searching of original articles found using the PubMed search. The titles and abstracts of every paper identified from the search were assessed for relevance by one author (K. M. R.) and any uncertainties were discussed with other members of the review team until a consensus was reached. This review was restricted to epidemiological studies (cross-sectional, case-control and cohort) and RCT in postmenopausal women with respect to test meals rich in SFA, MUFA and/or *n*-6 PUFA. Only published peer-reviewed literature was considered (i.e. 'grey' literature, such as dissertations, conference proceedings, reports, letters to editors and other non-peer-reviewed research, was excluded). In the present review, we only considered acute studies as our objectives were to determine the impact of meal fatty acids on non-fasting TAG responses, vascular function, blood pressure, and biomarkers of vascular function and inflammation as important CVD risk factors in postmenopausal women. Fig. 2 presents a summary of the literature search and reasons for exclusion of the studies.

Results and discussion

This systematic search identified 778 publications in total. Of these, there were nine relevant articles describing seven

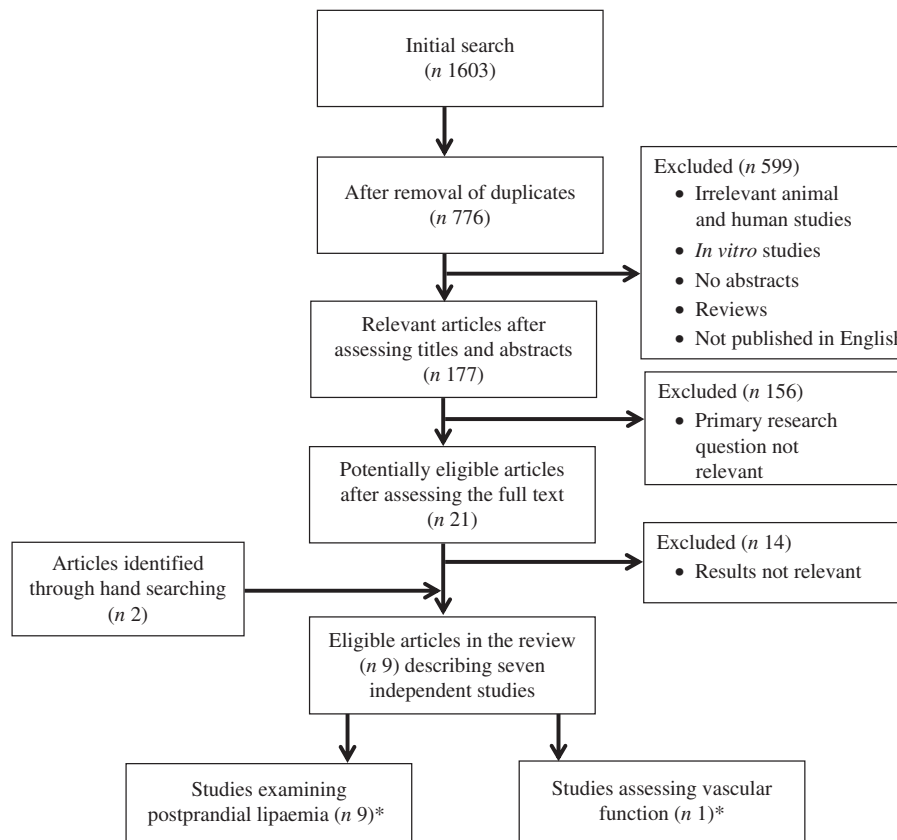


Fig. 2. Flow of information through the different phases of the review. * Of the studies included, one publication reported data on both postprandial lipaemia and vascular function.

independent studies in postmenopausal women that examined the acute effects of meals enriched in SFA and/or MUFA and/or *n*-6 PUFA on postprandial lipaemia^(96–104). One of these studies also determined the impact of a single high-fat meal with a PUFA:SFA ratio of 0.06 on vascular reactivity⁽¹⁰¹⁾ (Table 1). No studies were identified that reported the acute effects of meal fatty acids on postprandial blood pressure, or biomarkers of vascular function and inflammation in postmenopausal women. Only one single-blind RCT compared the effects of meal fat composition on postprandial lipaemia using a sequential meal protocol, the results of which were presented in three publications^(97,98,100). As opposed to a single-meal protocol, the use of a multiple-meal design by the researchers is considered superior because it more closely mimics the eating pattern of free-living individuals, particularly in Westernised societies, and provokes a sustained lipaemic response. Five publications described cross-sectional epidemiological studies, which were single-arm studies that did not include comparator meals, and whose fatty acid compositions varied^(96,101–104). Among these postprandial studies with blood samples collected between 6 to 10 h after the test meal, two studies^(96,102) used a sequential two-meal protocol, whereas the other three studies^(101,103,104) incorporated a single-meal approach. In addition, one case-control study was identified that considered the responses of normolipaemic, hypercholesterolaemic and mixed hyperlipidaemic postmenopausal women to a single high-fat meal⁽⁹⁹⁾.

Data on these human studies will be presented in two sections that address the effects of fatty acid composition on (i) postprandial lipaemia and (ii) postprandial vascular function in postmenopausal women.

Acute effects of meal fat composition on postprandial lipaemia

The five cross-sectional studies, investigating both single and sequential meals, provide consistent evidence that fat-rich loads, irrespective of fatty acid composition, augment postprandial TAG in postmenopausal women^(96,101–104) (Table 1). Furthermore, Pirro *et al.*⁽⁹⁹⁾ reported a significantly greater postprandial TAG response at 4, 6 and 8 h after a standardised oral fat load (65 g of fat) in mixed hyperlipidaemic women compared with hypercholesterolaemic and normolipidaemic women, which may reflect their higher baseline TAG concentrations. As expected, other factors involved in lipid metabolism, including increases in apo B-48⁽¹⁰²⁾, glucose⁽¹⁰³⁾ and insulin⁽¹⁰³⁾ as well as reductions in HDL-C^(99,103,104), glutathione⁽¹⁰¹⁾ and NEFA⁽¹⁰³⁾ were also observed postprandially compared with fasting values. However, comparison of the findings from the different studies are challenging due to differences in the nature of the fats and oils used in the test meal, the amount and composition of fat, and postprandial follow-up times, as well as the use of both single and sequential test meal protocols. They are also limited in their cross-sectional design in that the lack of comparator meals prevents any conclusions from being made regarding the impact of meal fat composition on postprandial lipaemia. Among all nine articles (seven independent studies) reported in Table 1, only one study described in three publications compared the postprandial lipaemic

responses to test meals containing oils rich in SFA (palm oil), MUFA (olive oil), *n*-6 PUFA (safflower oil) and a mixture of *n*-6 PUFA and *n*-3 PUFA (safflower and fish oils)^(97,98,100). In this study by Jackson and colleagues^(97,98,100), ten postmenopausal women ingested a high-fat breakfast containing 40 g of the assigned test fat followed by a low-fat, high-carbohydrate lunch (5.4 g total fat) given 5 h later. The authors observed significantly higher concentrations of plasma NEFA and lower insulin sensitivity following the SFA meal compared with the other test oils. During the postprandial state it has been shown that up to 50% of the liberated NEFA is dietary-derived CM-TAG due to the action of LPL upon TAG to release NEFA⁽¹⁰⁰⁾. Although Robertson *et al.*⁽¹⁰⁰⁾ did not determine the specific fatty acid composition of the circulating NEFA after consumption of the meals, a similar study reported the postprandial change in the plasma NEFA profile to represent the fatty acid composition of the test meals⁽¹⁰⁵⁾. Based on the same sequential-meal study, Jackson *et al.*⁽⁹⁸⁾ further examined the postprandial TAG and apo B-48 (the apolipoprotein specifically associated with CM) responses, including the responses in three distinct TRL sub-fractions, and reported significant differences in the apo B-48 time-course profiles between the four different test oils⁽⁹⁸⁾. In particular, the MUFA meal resulted in the formation of a greater number of both large (S_v >400 fraction) and moderately (S_v 60–400 fraction) sized apo B-48 particles compared with the other three study meals. The findings from this study suggested that olive oil may enhance CM formation and Jackson *et al.*⁽⁹⁷⁾ hypothesised that MUFA may modify the activity or expression of intestinal microsomal TAG transfer protein, which is involved with TRL assembly.

Acute effects of meal fat composition on vascular function

Only one study examined the acute impact of total fat and/or SFA and/or MUFA and/or *n*-6 PUFA on vascular reactivity in postmenopausal women. A significant decrease in the percentage FMD response at 2 h (2.3 (SD 2.6) %) compared with baseline (7.7 (SD 2.8) %; $P < 0.05$) was observed in healthy women after a 65 g oral fat load with a PUFA:SFA ratio of 0.06⁽¹⁰¹⁾ (Table 1). The lack of comparator meal in this study makes it difficult to draw conclusions regarding the impact of fatty acid composition on vascular function in postmenopausal women.

Summary

A systematic approach was used to review the literature on the impact of meal fat composition (SFA, MUFA and *n*-6 PUFA) on postprandial lipaemia, blood pressure, vascular function and biomarkers of vascular function and inflammation in postmenopausal women. However, there is at present an extremely limited number of RCT that have investigated the effects of meal fatty acid composition on measures of postprandial lipaemia and vascular function in this population subgroup. Furthermore, differences in study designs (such as the absence of a comparator test meal, and differences in meal fat composition, study duration and outcome measures) prevent any firm conclusions being drawn from the present literature review.



Table 1. Acute test-meal studies investigating the effects of meal fat content and composition on postprandial lipaemia and vascular function in postmenopausal women

Reference	Subject group, age (mean) and subjects (n)	Study design	Meal type	Amount of fat (% meal fat if available)	Fatty acid composition	Time of postprandial data	Postprandial measurements (plasma/serum)	Significant outcomes compared with baseline, unless otherwise stated
Postprandial lipaemia								
Westerveld <i>et al.</i> (1996) ⁽¹⁰⁴⁾	59 years (n 16) normolipidaemic	Cross-sectional*	Single	50 g (40%)	PUFA:SFA 0.06	8 h	TAG, HDL-C and HDL-apo A-1	↓ HDL-C at 3 to 8 h ($P < 0.05$) ↓ HDL-apo A-1 at 3 and 6 h ($P < 0.05$) ↑ TAG at 8 h ($P < 0.05$)
Pirro <i>et al.</i> (2001) ⁽⁹⁹⁾	57 years (n 17) normolipidaemic, 54 years (n 17) hypercholesterolaemic and 55 years (n 16) mixed hyperlipidaemic	Case-control	Single	65 g (83%)	PUFA:SFA 0.06	8 h	TC, TAG, HDL-C, HDL ₂ HDL ₃ , LDL, LDL particle size and Lp(a)	↑ TAG at 4, 6 and 8 h, ↓ HDL-C at 6 h and ↓ Lp(a) at 4 and 6 h in normolipidaemic PoM ($P < 0.05$) ↑ TAG at 4, 6 and 8 h, ↓ HDL-C at 4 and 6 h, ↓ HDL ₂ at 4 h and ↓ Lp(a) at 4 h in hypercholesterolaemic women ($P < 0.05$) ↑ TAG at 4, 6 and 8 h, ↓ LDL size at 4 and 6 h, ↓ HDL-C at 4, 6 and 8 h, ↓ HDL ₂ at 6 h and ↓ Lp(a) at 4 and 6 h in mixed hyperlipidaemic PoM ($P < 0.05$)
Silva <i>et al.</i> (2005) ⁽¹⁰²⁾	52–76 years (62 years) (n 17)	Cross-sectional*	Sequential	Breakfast: 30 g (46%E) Lunch: 44 g (52%E)	Breakfast: 27%E SFA, 12%E MUFA, 5%E PUFA and 2%E <i>trans</i> -fatty acids Lunch: 27%E SFA, 18%E MUFA, 5%E PUFA and 2%E <i>trans</i> -fatty acids	10 h	TAG, and apo B-48	↑ TAG at 3.5 h after breakfast and 1 h after lunch ↑ apo B-48 at 2.5 h after breakfast and 1 h after lunch
Alssema <i>et al.</i> (2008) ⁽⁹⁶⁾	60.1 years (n 76)	Cross-sectional*	Sequential	Both breakfast and lunch compositions: Fat-rich meal: 50 g fat, 56 g CHO, 28 g protein CHO-rich meal: 4 g fat, 162 g CHO, 22 g protein	No information	8 h	TAG, HDL-C and CETP	↑ TAG at 8 h ($P < 0.05$), ↓ HDL-C at 8 h ($P < 0.05$) after the fat-rich meal ↑ TAG at 8 h ($P < 0.05$), ↓ HDL-C at 8 h and ↑ CETP after the CHO-rich meal ($P < 0.05$)
Wassef <i>et al.</i> (2012) ⁽¹⁰³⁾	58 years (45–74 years) (n 19) obese	Cross-sectional*	Single	¹³ C-labelled breakfast 80 g fat (68%) + 0.017 g ¹³ C-triolein/g fat	25%E SFA, 26%E MUFA, 10%E PUFA and 6%E other sources	6 h	TAG, glucose, NEFA and insulin	↓ NEFA between 1 and 2 h Plasma peak TAG at 4 h Plasma peak glucose at 1 h Serum peak insulin at 1 h
Robertson <i>et al.</i> (2002) ⁽¹⁰⁰⁾ Jackson <i>et al.</i> (2002) ⁽⁹⁸⁾ Jackson <i>et al.</i> (2002) ⁽⁹⁷⁾	50–63 years (56 years) (n 10)	Single-blind randomised cross-over	Sequential	Breakfast: 41 g† Lunch: 6 g	High SFA (g/100 g): 10 g <i>n</i> -6 PUFA, 0 g <i>n</i> -3 PUFA, 40 g MUFA and 50 g SFA High MUFA (g/100 g): 11 g <i>n</i> -6 PUFA, 0 g <i>n</i> -3 PUFA, 72 g MUFA and 17 g SFA High <i>n</i> -6 PUFA (g/100 g): 74 g <i>n</i> -6 PUFA, 0 g <i>n</i> -3 PUFA, 15 g MUFA and 11 g SFA High <i>n</i> -3/ <i>n</i> -6 PUFA (g/100 g): 39 g <i>n</i> -6 PUFA, 22 g <i>n</i> -3 PUFA, 22 g MUFA and 19 g SFA	8 h	Glucose, NEFA and insulin TAG and apo B-48 in plasma and three TAG-rich lipoprotein subfractions	Insulin response: SFA > <i>n</i> -6 PUFA = <i>n</i> -3 PUFA = MUFA ($P < 0.006$) Glucose: No significant effect ↑ NEFA at 5 h following high SFA breakfast Insulin sensitivity: <i>n</i> -6 PUFA = <i>n</i> -3 PUFA = MUFA > SFA ↑ Sf > 400 fraction apo B-48 response after MUFA than SFA, <i>n</i> -6 PUFA and <i>n</i> -3/ <i>n</i> -6 PUFA meals ($P < 0.002$) ↑ apo B-48 IAUC in the Sf > 400 fraction after the MUFA than SFA, <i>n</i> -6 PUFA and <i>n</i> -3/ <i>n</i> -6 PUFA meals ($P < 0.04$)
Postprandial lipaemia and vascular function								
Siepi <i>et al.</i> (2002) ⁽¹⁰¹⁾	57 years (n 10)	Cross-sectional*	Single	65 g	PUFA:SFA 0.06	6 h	TAG and GSH Brachial FMD	↑ TAG at 4 and 6 h ($P < 0.05$) ↓ GSH at 2 h ($P < 0.05$) ↓ FMD at 2 h ($P < 0.05$)

HDL-C, HDL-cholesterol; ↓, decrease over time relative to baseline (fasting) unless otherwise specified; ↑, increase over time relative to baseline (fasting) unless otherwise specified; TC, total cholesterol; Lp(a), lipoprotein(a); PoM, postmenopausal women; %E, energy percentage; CHO, carbohydrate; CETP, cholesteryl ester transfer protein; IAUC, incremental AUC; FMD, flow-mediated dilatation; GSH, glutathione.

* No comparator group.

† Fatty acid values given per 100 g of test oil, of which 41 g was included in the breakfast.

Conclusions

In conclusion, there is an urgent requirement for suitably powered RCT to investigate the effects of meal fat composition on postprandial lipaemia and vascular function in postmenopausal women. With the increased prevalence of non-communicable diseases in women, especially after the menopause, future studies should consider both healthy postmenopausal women and those at increased cardiometabolic risk using well-standardised measures of vascular function. Since non-fasting TAG is an important CVD risk factor for women, it would be preferable to use robust test-meal protocols that are more reflective of habitual eating patterns to gain a greater understanding of the day-long postprandial handling of different dietary fats.

Acknowledgements

K. M. R. was supported by the Commonwealth Scholarship Commission, UK. This research received no specific grant from any funding agency, commercial or non-profit sectors.

The authors' responsibilities were as follows: K. M. R., M. W., K. G. J. and J. A. L. contributed to the conception of the literature search strategy. K. M. R. undertook the literature search, extracted and interpreted the data from the literature and wrote the manuscript. M. W., K. G. J. and J. A. L. critically appraised the document at all stages. J. A. L. was responsible for the final content.

None of the authors has any conflicts of interest.

Supplementary material

To view supplementary material for this article, please visit <https://doi.org/10.1017/S0954422418000033>

References

- Schwenke DC (1998) Antioxidants and atherogenesis. *J Nutr Biochem* **9**, 424–445.
- World Health Organization (2017) Top 10 causes of death worldwide. <http://www.who.int/mediacentre/factsheets/fs310/en/> (accessed March 2017).
- British Heart Foundation (2014) *Cardiovascular Disease Statistics 2014*. London: BHF.
- World Health Organization (2008) *The Global Burden of Disease: 2004 Update*. Geneva: WHO.
- Libby P, Ridker PM & Maseri A (2002) Inflammation and atherosclerosis. *Circulation* **105**, 1135–1143.
- Smith SC (2007) Multiple risk factors for cardiovascular disease and diabetes mellitus. *Am J Med* **120**, S3–S11.
- Celermajer DS, Sorensen KE, Spiegelhalter DJ, *et al.* (1994) Aging is associated with endothelial dysfunction in healthy men years before the age-related decline in women. *J Am Coll Cardiol* **24**, 471–476.
- Pokoradi AJ, Iversen L & Hannaford PC (2011) Factors associated with age of onset and type of menopause in a cohort of UK women. *Am J Obstet Gynecol* **34**, e31–e34.
- Wenger NK, Speroff L & Packard B (1993) Cardiovascular health and disease in women. *N Engl J Med* **329**, 247–256.
- Van der Schouw Y, van der Graaf Y, Steyerberg E, *et al.* (1996) Age at menopause as a risk factor for cardiovascular mortality. *Lancet* **347**, 714–718.
- Yusuf S, Hawken S, Öunpuu S, *et al.* (2004) Effect of potentially modifiable risk factors associated with myocardial infarction in 52 countries (the INTERHEART study): case-control study. *Lancet* **364**, 937–952.
- Department of Health (1991) *Dietary Reference Values for Food Energy and Nutrients in the United Kingdom: Report on the Panel of Dietary Reference Values of the Committee on Medical Aspects of Food Policy. Report of Health and Social Subjects*, no. 41. London: TSO.
- Hooper L, Summerbell CD, Thompson R, *et al.* (2012) Reduced or modified dietary fat for preventing cardiovascular disease. *Cochrane Database Syst Rev*, issue 5, CD002137.
- Kolovou GD, Mikhailidis DP, Nordestgaard BG, *et al.* (2011) Definition of postprandial lipaemia. *Curr Vasc Pharmacol* **9**, 292–301.
- Björkegren J, Packard C, Hamsten A, *et al.* (1996) Accumulation of large very low density lipoprotein in plasma during intravenous infusion of a chylomicron-like triglyceride emulsion reflects competition for a common lipolytic pathway. *J Lipid Res* **37**, 76–86.
- Jackson KG, Poppitt SD & Minihane AM (2012) Postprandial lipemia and cardiovascular disease risk: interrelationships between dietary, physiological and genetic determinants. *Atherosclerosis* **220**, 22–33.
- Chapman MJ, Le Goff W, Guerin M, *et al.* (2010) Cholesteryl ester transfer protein: at the heart of the action of lipid-modulating therapy with statins, fibrates, niacin, and cholesteryl ester transfer protein inhibitors. *Eur Heart J* **31**, 149–164.
- Bansal S, Buring JE, Rifai N, *et al.* (2007) Fasting compared with nonfasting triglycerides and risk of cardiovascular events in women. *JAMA* **298**, 309–316.
- Langsted A, Freiberg J, Tybjaerg-Hansen A, *et al.* (2011) Nonfasting cholesterol and triglycerides and association with risk of myocardial infarction and total mortality: The Copenhagen City Heart Study with 31 years of follow-up. *J Intern Med* **270**, 65–75.
- Lindman AS, Veierød M, Tverdal A, *et al.* (2010) Nonfasting triglycerides and risk of cardiovascular death in men and women from the Norwegian Counties Study. *Eur J Epidemiol* **25**, 789–798.
- van Beek AP, de Ruijter-Heijstek FC, Erkelens DW, *et al.* (1999) Menopause is associated with reduced protection from postprandial lipemia. *Arterioscler Thromb Vasc Biol* **19**, 2737–2741.
- Schoppen S, Perez-Granados AM, Navas-Carretero S, *et al.* (2010) Postprandial lipaemia and endothelial adhesion molecules in pre- and postmenopausal Spanish women. *Nutr Hosp* **25**, 256–261.
- Masding MG, Stears AJ, Burdge GC, *et al.* (2006) The benefits of oestrogens on postprandial lipid metabolism are lost in post-menopausal women with type 2 diabetes. *Diabet Med* **23**, 768–774.
- Jackson KG, Abraham EC, Smith AM, *et al.* (2010) Impact of age and menopausal status on the postprandial triacylglycerol response in healthy women. *Atherosclerosis* **208**, 246–252.
- Cohn JS, McNamara J, Cohn S, *et al.* (1988) Postprandial plasma lipoprotein changes in human subjects of different ages. *J Lipid Res* **29**, 469–479.
- Jackson KG, Clarke DT, Murray P, *et al.* (2010) Introduction to the DISRUPT postprandial database: subjects, studies and methodologies. *Genes Nutr* **5**, 39–48.

27. Koutsari C, Zagana A, Tzoras I, *et al.* (2004) Gender influence on plasma triacylglycerol response to meals with different monounsaturated and saturated fatty acid content. *Eur J Clin Nutr* **58**, 495–502.
28. Tentor J, Harada LM, Nakamura RT, *et al.* (2006) Sex-dependent variables in the modulation of postalimentary lipemia. *Nutr J* **22**, 9–15.
29. Burdge GC, Powell J & Calder PC (2006) Lack of effect of meal fatty acid composition on postprandial lipid, glucose and insulin responses in men and women aged 50–65 years consuming their habitual diets. *Br J Nutr* **96**, 489–500.
30. Bass KM, Newschaffer CJ, Klag MJ, *et al.* (1993) Plasma lipoprotein levels as predictors of cardiovascular death in women. *Arch Intern Med* **153**, 2209–2216.
31. Kannel WB (1987) Metabolic risk factors for coronary heart disease in women: perspective from the Framingham Study. *Am Heart J* **114**, 413–419.
32. Nordestgaard BG, Benn M, Schnohr P, *et al.* (2007) Nonfasting triglycerides and risk of myocardial infarction, ischemic heart disease, and death in men and women. *JAMA* **298**, 299–308.
33. Burger HG, Dudley EC, Robertson DM, *et al.* (2002) Hormonal changes in the menopause transition. *Recent Prog Horm Res* **57**, 257–276.
34. Žegura B & Gužič-Salobir B (2009) Hormone replacement therapy in postmenopause and cardiovascular diseases: facts and dilemmas. *Zdrav Vestn* **78**, 1165–1168.
35. Vehkavaara S, Silveira A, Hakala-Ala-Pietilä T, *et al.* (2001) Effects of oral and transdermal estrogen replacement therapy on markers of coagulation, fibrinolysis, inflammation and serum lipids and lipoproteins in postmenopausal women. *Thromb Haemost* **85**, 619–625.
36. Windler E, Kovanen PT, Chao Y-S, *et al.* (1980) The estradiol-stimulated lipoprotein receptor of rat liver-A binding site that membrane mediates the uptake of rat lipoproteins containing apoproteins B and E. *J Biol Chem* **255**, 10464–10471.
37. Gordon DJ, Probstfield JL, Garrison RJ, *et al.* (1989) High-density lipoprotein cholesterol and cardiovascular disease. Four prospective American studies. *Circulation* **79**, 8–15.
38. Bessesen DH, Cox-York KA, Hernandez TL, *et al.* (2015) Postprandial triglycerides and adipose tissue storage of dietary fatty acids: Impact of menopause and estradiol. *Obesity* **23**, 145–153.
39. Westerveld HT, Kock L, Van Rijn H, *et al.* (1995) 17β-Estradiol improves postprandial lipid metabolism in postmenopausal women. *J Clin Endocrinol Metab* **80**, 249–253.
40. Huang Y, Wang S, Cai X, *et al.* (2013) Prehypertension and incidence of cardiovascular disease: a meta-analysis. *BMC Med* **11**, 177.
41. Etehad D, Emdin CA, Kiran A, *et al.* (2016) Blood pressure lowering for prevention of cardiovascular disease and death: a systematic review and meta-analysis. *Lancet* **387**, 957–967.
42. Vlachopoulos C, Aznaouridis K & Stefanadis C (2010) Prediction of cardiovascular events and all-cause mortality with arterial stiffness: a systematic review and meta-analysis. *J Am Coll Cardiol* **55**, 1318–1327.
43. Ras RT, Streppel MT, Draaijer R, *et al.* (2013) Flow-mediated dilation and cardiovascular risk prediction: a systematic review with meta-analysis. *Int J Cardiol* **168**, 344–351.
44. Matsuzawa Y, Kwon TG, Lennon RJ, *et al.* (2015) Prognostic value of flow-mediated vasodilation in brachial artery and fingertip artery for cardiovascular events: a systematic review and meta-analysis. *J Am Heart Assoc* **4**, e002270.
45. Yao S-K, Ober JC, Krishnaswami A, *et al.* (1992) Endogenous nitric oxide protects against platelet aggregation and cyclic flow variations in stenosed and endothelium-injured arteries. *Circulation* **86**, 1302–1309.
46. Cohen RA (1995) The role of nitric oxide and other endothelium-derived vasoactive substances in vascular disease. *Prog Cardiovasc Dis* **38**, 105–128.
47. Verma S & Anderson TJ (2002) Fundamentals of endothelial function for the clinical cardiologist. *Circulation* **105**, 546–549.
48. Widlansky ME, Gokce N, Keaney JF, *et al.* (2003) The clinical implications of endothelial dysfunction. *J Am Coll Cardiol* **42**, 1149–1160.
49. Rossi R, Nuzzo A, Origliani G, *et al.* (2008) Prognostic role of flow-mediated dilation and cardiac risk factors in postmenopausal women. *J Am Coll Cardiol* **51**, 997–1002.
50. Fichtlscherer S, Rosenberger G, Walter DH, *et al.* (2000) Elevated C-reactive protein levels and impaired endothelial vasoreactivity in patients with coronary artery disease. *Circulation* **102**, 1000–1006.
51. Moens AL, Goovaerts I, Claeys MJ, *et al.* (2005) Flow-mediated vasodilation: a diagnostic instrument, or an experimental tool? *Chest* **127**, 2254–2263.
52. Inaba Y, Chen JA & Bergmann SR (2010) Prediction of future cardiovascular outcomes by flow-mediated vasodilation of brachial artery: a meta-analysis. *Int J Cardiovasc Imaging* **26**, 631–640.
53. Schächinger V, Britten MB & Zeiher AM (2000) Prognostic impact of coronary vasodilator dysfunction on adverse long-term outcome of coronary heart disease. *Circulation* **101**, 1899–1906.
54. Moreau KL, Hildreth KL, Meditz AL, *et al.* (2012) Endothelial function is impaired across the stages of the menopause transition in healthy women. *J Clin Endocrinol Metab* **97**, 4692–4700.
55. Majmudar N, Robson S & Ford G (2000) Effects of the menopause, gender, and estrogen replacement therapy on vascular nitric oxide activity. *J Clin Endocrinol Metab* **85**, 1577–1583.
56. Gilligan DM, Badar DM, Panza JA, *et al.* (1994) Acute vascular effects of estrogen in postmenopausal women. *Circulation* **90**, 786–791.
57. Brodsky SV, Gealekman O, Chen J, *et al.* (2004) Prevention and reversal of premature endothelial cell senescence and vasculopathy in obesity-induced diabetes by ebselen. *Circ Res* **94**, 377–384.
58. Ospina JA, Duckles SP & Krause DN (2003) 17β-Estradiol decreases vascular tone in cerebral arteries by shifting COX-dependent vasoconstriction to vasodilation. *Am J Physiol Heart Circ Physiol* **285**, H241–H250.
59. Ospina JA, Krause DN & Duckles SP (2002) 17β-Estradiol increases rat cerebrovascular prostacyclin synthesis by elevating cyclooxygenase-1 and prostacyclin synthase. *Stroke* **33**, 600–605.
60. Manson JE, Aragaki AK, Rossouw JE, *et al.* (2017) Menopausal hormone therapy and long-term all-cause and cause-specific mortality: The Women’s Health Initiative randomized trials. *JAMA* **318**, 927–938.
61. Hsia J, Langer RD, Manson JE, *et al.* (2006) Conjugated equine estrogens and coronary heart disease: The Women’s Health Initiative. *Arch Intern Med* **166**, 357–365.
62. Marjoribanks J, Farquhar C, Roberts H, *et al.* (2017) Long-term hormone therapy for perimenopausal and postmenopausal women. *Cochrane Database Syst Rev*, issue 1, CD004143.

63. Gartlehner G, Patel SV, Feltner C, *et al.* (2017) Hormone therapy for the primary prevention of chronic conditions in postmenopausal women: evidence report and systematic review for the US Preventive Services Task Force. *JAMA* **318**, 2234–2249.
64. Žegura B, Gužic-Salobir B, Šebešćten M, *et al.* (2006) The effect of various menopausal hormone therapies on markers of inflammation, coagulation, fibrinolysis, lipids, and lipoproteins in healthy postmenopausal women. *Menopause* **13**, 643–650.
65. Knopp RH, Zhu X & Bonet B (1994) Effects of estrogens on lipoprotein metabolism and cardiovascular disease in women. *Atherosclerosis* **110**, S83–S91.
66. Vigen C, Hodis H, Chandler W, *et al.* (2007) Postmenopausal oral estrogen therapy affects hemostatic factors, but does not account for reduction in the progression of subclinical atherosclerosis. *J Thromb Haemost* **5**, 1201–1208.
67. Wassertheil-Smoller S, Anderson G, Psaty BM, *et al.* (2000) Hypertension and its treatment in postmenopausal women. *Hypertension* **36**, 780–789.
68. Nash D, Magder L, Lustberg M, *et al.* (2003) Blood lead, blood pressure, and hypertension in perimenopausal and postmenopausal women. *JAMA* **289**, 1523–1532.
69. Abramson BL & Melvin RG (2014) Cardiovascular risk in women: focus on hypertension. *Can J Cardiol* **30**, 553–559.
70. Donoghue M, Hsieh F, Baronas E, *et al.* (2000) A novel angiotensin-converting enzyme-related carboxypeptidase (ACE2) converts angiotensin I to angiotensin 1–9. *Circ Res* **87**, e1–e9.
71. Leung PS (2004) The peptide hormone angiotensin II: its new functions in tissues and organs. *Curr Protein Pept Sci* **5**, 267–273.
72. Bell C (1973) Oestrogen-induced sensitization of the uterine artery of the guinea-pig to acetylcholine. *Br J Pharmacol* **49**, 595–601.
73. Bell C & Coffey C (1982) Factors influencing oestrogen-induced sensitization to acetylcholine of guinea-pig uterine artery. *J Reprod Fertil* **66**, 133–137.
74. Williams JK, Adams MR, Herrington DM, *et al.* (1992) Short-term administration of estrogen and vascular responses of atherosclerotic coronary arteries. *J Am Coll Cardiol* **20**, 452–457.
75. Hinojosa-Laborde C, Craig T, Zheng W, *et al.* (2004) Ovariectomy augments hypertension in aging female Dahl salt-sensitive rats. *Hypertension* **44**, 405–409.
76. Nickenig G, Bäumer AT, Grohè C, *et al.* (1998) Estrogen modulates AT1 receptor gene expression *in vitro* and *in vivo*. *Circulation* **97**, 2197–2201.
77. Nogawa N, Sumino H, Ichikawa S, *et al.* (2001) Effect of long-term hormone replacement therapy on angiotensin-converting enzyme activity and bradykinin in postmenopausal women with essential hypertension and normotensive postmenopausal women. *Menopause* **8**, 210–215.
78. Schunkert H, Danser AJ, Hense H-W, *et al.* (1997) Effects of estrogen replacement therapy on the renin-angiotensin system in postmenopausal women. *Circulation* **95**, 39–45.
79. Ginnan R, Guikema BJ, Halligan KE, *et al.* (2008) Regulation of smooth muscle by inducible nitric oxide synthase and NADPH oxidase in vascular proliferative diseases. *Free Radic Biol Med* **44**, 1232–1245.
80. Ono H, Minatoguchi S, Watanabe K, *et al.* (2008) Candesartan decreases carotid intima-media thickness by enhancing nitric oxide and decreasing oxidative stress in patients with hypertension. *Hypertens Res* **31**, 271–279.
81. Wassmann S, Wassmann K & Nickenig G (2004) Modulation of oxidant and antioxidant enzyme expression and function in vascular cells. *Hypertension* **44**, 381–386.
82. Fischer M, Baessler A & Schunkert H (2002) Renin angiotensin system and gender differences in the cardiovascular system. *Cardiovasc Res* **53**, 672–677.
83. Mozaffarian D, Micha R & Wallace S (2010) Effects on coronary heart disease of increasing polyunsaturated fat in place of saturated fat: a systematic review and meta-analysis of randomized controlled trials. *PLoS Med* **23**, e1000252.
84. Monfort-Pires M, Delgado-Lista J, Gomez-Delgado F, *et al.* (2016) Impact of the content of fatty acids of oral fat tolerance tests on postprandial triglyceridemia: systematic review and meta-analysis. *Nutrients* **8**, E580.
85. West SG (2001) Effect of diet on vascular reactivity: an emerging marker for vascular risk. *Curr Atheroscler Rep* **3**, 446–455.
86. Ong PJ, Dean TS, Hayward CS, *et al.* (1999) Effect of fat and carbohydrate consumption on endothelial function. *Lancet* **354**, 2134.
87. Plotnick GD, Corretti MC & Vogel RA (1997) Effect of antioxidant vitamins on the transient impairment of endothelium-dependent brachial artery vasoactivity following a single high-fat meal. *JAMA* **278**, 1682–1686.
88. Vogel RA, Corretti MC & Plotnick GD (1997) Effect of a single high-fat meal on endothelial function in healthy subjects. *Am J Cardiol* **79**, 350–354.
89. Vogel RA, Corretti MC & Plotnick GD (2000) The postprandial effect of components of the Mediterranean diet on endothelial function. *J Am Coll Cardiol* **36**, 1455–1460.
90. Pacher P, Beckman JS & Liaudet L (2007) Nitric oxide and peroxynitrite in health and disease. *Physiol Rev* **87**, 315–424.
91. Förstermann U (2010) Nitric oxide and oxidative stress in vascular disease. *Pflugers Arch* **459**, 923–939.
92. Margioris AN (2009) Fatty acids and postprandial inflammation. *Curr Opin Clin Nutr Metab Care* **12**, 129–137.
93. Rubin D, Claas S, Pfeuffer M, *et al.* (2008) s-ICAM-1 and s-VCAM-1 in healthy men are strongly associated with traits of the metabolic syndrome, becoming evident in the postprandial response to a lipid-rich meal. *Lipids Health Dis* **7**, 32.
94. Hall WL (2009) Dietary saturated and unsaturated fats as determinants of blood pressure and vascular function. *Nutr Res Rev* **22**, 18–38.
95. Vafeiadou K, Weech M, Sharma V, *et al.* (2012) A review of the evidence for the effects of total dietary fat, saturated, monounsaturated and *n*-6 polyunsaturated fatty acids on vascular function, endothelial progenitor cells and microparticles. *Br J Nutr* **107**, 303–324.
96. Alssema M, Schindhelm RK, Dekker JM, *et al.* (2008) Determinants of postprandial triglyceride and glucose responses after two consecutive fat-rich or carbohydrate-rich meals in normoglycemic women and in women with type 2 diabetes mellitus: The Hoorn Prandial Study. *Metabolism* **57**, 1262–1269.
97. Jackson KG, Robertson MD, Fielding BA, *et al.* (2002) Olive oil increases the number of triacylglycerol-rich chylomicron particles compared with other oils: an effect retained when a second standard meal is fed. *Am J Clin Nutr* **76**, 942–949.
98. Jackson KG, Robertson MD, Fielding BA, *et al.* (2002) Measurement of apolipoprotein B-48 in the Svedberg flotation rate (Sf) > 400, Sf 60–400 and Sf 20–60 lipoprotein fractions reveals novel findings with respect to the effects of dietary fatty acids on triacylglycerol-rich lipoproteins in postmenopausal women. *Clin Sci* **103**, 227–237.



99. Pirro M, Lupattelli G, Siepi D, *et al.* (2001) Postprandial lipemia and associated metabolic disturbances in healthy and hyperlipemic postmenopausal women. *Metabolism* **50**, 330–334.
100. Robertson M, Jackson KG, Fielding B, *et al.* (2002) Acute effects of meal fatty acid composition on insulin sensitivity in healthy post-menopausal women. *Br J Nutr* **88**, 635–640.
101. Siepi D, Marchesi S, Lupattelli G, *et al.* (2002) Postprandial endothelial impairment and reduced glutathione levels in postmenopausal women. *Ann Nutr Metab* **46**, 32–37.
102. Silva K, Wright JW, Williams CM, *et al.* (2005) Meal ingestion provokes entry of lipoproteins containing fat from the previous meal: possible metabolic implications. *Eur J Clin Nutr* **44**, 377–383.
103. Wassef H, Salem H, Bissonnette S, *et al.* (2012) White adipose tissue apolipoprotein C-I secretion in relation to delayed plasma clearance of dietary fat in humans. *Arterioscler Thromb Vasc Biol* **32**, 2785–2793.
104. Westerveld HT, Meijer E, Erkelens DW, *et al.* (1996) Postprandial reduction in high-density lipoprotein cholesterol concentrations in postmenopausal women: improvement by 17 β -estradiol. *Metabolism* **45**, 827–832.
105. Fielding BA, Callow J, Owen RM, *et al.* (1996) Postprandial lipemia: the origin of an early peak studied by specific dietary fatty acid intake during sequential meals. *Am J Clin Nutr* **63**, 36–41.
106. Davis SR, Lambrinou I, Lumsden M, *et al.* (2015) Menopause. *Nat Rev Dis Primers* **1**, 15004.