

**Effect of the Mediterranean diet on the fecal long-chain fatty acid composition and intestinal barrier integrity: An exploratory analysis of the randomized controlled LIBRE trial**

Benjamin Seethaler<sup>1</sup>, Maryam Basrai<sup>1</sup>, Audrey M. Neyrinck<sup>2</sup>, Walter Vetter<sup>3</sup>, Nathalie M. Delzenne<sup>2</sup>, Marion Kiechle<sup>4</sup>, Stephan C. Bischoff<sup>1</sup>

<sup>1</sup>Institute of Nutritional Medicine, University of Hohenheim, Stuttgart, Germany

<sup>2</sup>Metabolism and Nutrition Research Group, Louvain Drug Research Institute, UCLouvain, Université catholique de Louvain, Brussels, Belgium

<sup>3</sup>Institute of Food Chemistry, University of Hohenheim, Stuttgart, Germany

<sup>4</sup>Department of Gynecology, Center for Hereditary Breast and Ovarian Cancer, Klinikum Rechts der Isar, Technical University Munich and Comprehensive Cancer Center Munich, Munich, Germany

**Corresponding author:** Stephan C. Bischoff, M.D., Professor of Medicine, Institute of Nutritional Medicine, University of Hohenheim, Fruwirthstr. 12, 70593 Stuttgart. Phone: +49 711 45924101. Fax: +49 711 45924343, Germany. E-mail: bischoff.stephan@uni-hohenheim.de

**Short title:** Mediterranean diet and fecal fatty acids

**Keywords:** Mediterranean diet; fecal fatty acids; LCFAs; gut barrier; intestinal barrier; gut permeability.



This peer-reviewed article has been accepted for publication but not yet copyedited or typeset, and so may be subject to change during the production process. The article is considered published and may be cited using its DOI

10.1017/S0007114524001788

The British Journal of Nutrition is published by Cambridge University Press on behalf of The Nutrition Society

**Abstract**

We recently showed that adherence to the Mediterranean diet increased the proportion of plasma omega-3 polyunsaturated fatty acids (*n*-3 PUFAs), which was associated with an improved intestinal barrier integrity. In the present exploratory analysis, we assessed fecal fatty acids in the same cohort, aiming to investigate possible associations with intestinal barrier integrity. Women from the LIBRE randomized controlled trial, characterized by an impaired intestinal barrier integrity, followed either a Mediterranean diet (intervention group, IG, *n*=33) or a standard diet (control group, CG, *n*=35). At baseline (BL), month 3 (V1), and month 12 (V2), plasma lipopolysaccharide binding protein (LBP), fecal zonulin, and fecal fatty acids were measured. In the IG, fecal proportions of palmitoleic acid (16:1, *n*-7) and arachidonic acid (20:4, *n*-6) decreased, while the proportion of linoleic acid (18:2, *n*-6) and alpha linoleic acid (18:3, *n*-3) increased (BL-V1 and BL-V2, all *P*<0.08). In the CG, fecal proportions of palmitic acid and arachidic acid increased while the proportion of linoleic acid decreased (BL-V1, all *P*<0.05). The decrease in the proportion of palmitoleic acid correlated with the decrease in plasma LBP ( $\Delta$ V1-BL *r*=0.72, *P*<0.001;  $\Delta$ V2-BL *r*=0.39, *P*<0.05) and correlated inversely with adherence to the Mediterranean diet (Mediterranean diet score;  $\Delta$ V1-BL *r*=-0.42, *P*=0.03;  $\Delta$ V2-BL *r*=-0.53, *P*=0.005) in the IG. Our data show that adherence to the Mediterranean diet induces distinct changes in the fecal fatty acid composition. Furthermore, our data indicate that the fecal proportion of palmitoleic acid, but not fecal *n*-3 PUFAs, are associated with intestinal barrier integrity in the IG.

## INTRODUCTION

The intestinal barrier protects the host against gut microbes, food antigens, and toxins present in the gastrointestinal tract. A functioning intestinal barrier is required for gut health, whereas intestinal barrier impairment is associated with a wide range of noncommunicable diseases, including cardiovascular disease, cancer, type 2 diabetes, and inflammatory bowel disease<sup>(1,2)</sup>.

Intestinal barrier integrity is affected by several endogenous and exogenous factors including diet, stress, excessive body weight, and low or extreme physical activity<sup>(1,3,4)</sup>. To assess intestinal barrier integrity in larger cohorts, we have recently validated plasma lipopolysaccharide binding protein (LBP) and fecal zonulin as suitable biomarkers<sup>(5)</sup>.

Even though it has been shown that the intestinal barrier plays a central role in health and disease, and the interest in this topic is increasing rapidly, the mechanisms by which the barrier function is regulated are not well known. We and others have shown that short-chain fatty acids, derived from bacterial fermentation of dietary fibers, improve intestinal barrier integrity<sup>(6-9)</sup>. Also, vitamins, minerals, amino acids and polyphenols might have an effect<sup>(10,11)</sup>. For the first time in a human trial, we have recently shown that diet-derived omega-3 polyunsaturated fatty acids (*n*-3 PUFAs), originated from increased adherence to the Mediterranean diet and assessed in plasma, are associated with an improvement in a previously impaired intestinal barrier integrity<sup>(12)</sup>. Data for this study derived from the randomized controlled LIBRE (Lifestyle Intervention Study in Women with Hereditary Breast and Ovarian Cancer) trial, which included women at high risk for breast and ovarian cancer due to a pathogenic germline mutation in the BRCA1 and/or BRCA2 genes<sup>(13)</sup>. These mutations have been shown to be associated with an altered intestinal barrier integrity<sup>(6,14)</sup>. In detail, LIBRE study participants showed a higher median concentration of fecal zonulin at baseline compared with a healthy cohort of 36 women with a similar range of age and BMI (178 ng/mg versus 110 ng/mg,  $P < 0.01$ , Mann–Whitney *U* test)<sup>(5)</sup>. It has been suggested that intestinal barrier impairment may be linked to breast cancer initiation and progression<sup>(15)</sup>, which is the reason why we are interested in investigating factors that alter intestinal barrier integrity.

We have previously shown that short-chain fatty acids (SCFAs) induced by dietary fibers from the Mediterranean diet stabilize intestinal barrier integrity<sup>(6)</sup>. Now we wondered if long-chain fatty acids (LCFAs) also regulate intestinal barrier function. We previously found that the proportion of plasma *n*-3 PUFAs, as well as plasma saturated fatty acids, were associated

with an improvement of intestinal barrier integrity<sup>(12)</sup>. Also, recent studies revealed that fiber supplementation alters the fecal proportions of LCFAs<sup>(16,17)</sup>. Since these previous findings are of clinical relevance, we have decided to investigate these in more detail in the LIBRE study. The aim of the present study is to investigate possible associations between the fiber-rich Mediterranean diet<sup>(18)</sup>, fecal proportions of LCFAs, and biomarkers of intestinal barrier integrity.

## MATERIALS & METHODS

### *Study design*

Data for the present exploratory study derived from the randomized controlled LIBRE 1 trial<sup>(13)</sup>. This study was a randomized (1:1 ratio; group allocation performed centrally using randomly permuted blocks of length 2–6; participating center and previous breast cancer were used as stratification factors), prospective, open-label, two-armed controlled multicenter trial, aiming to test the effect of a structured lifestyle intervention program focussing on the Mediterranean diet and increased physical activity on cancer-relevant outcomes. This study started in 2014 and included women at high risk for breast and ovarian cancer due to a pathogenic germline mutation in the BRCA1 and/or BRCA2 genes.

The primary endpoint was the number of participants who successfully completed the first 3 months of lifestyle intervention. A rate of 70% adherence or more was considered as success. Secondary endpoints comprised body mass index (BMI), physical fitness, the analyses of omega fatty acids, and fecal metabolites. The sample size in LIBRE-1 was adjusted to this goal but was not calculated based on statistical assumptions and tests, as described in the study protocol<sup>(13)</sup>. The LIBRE-2 confirmatory study, which aims to include 600 women, started in 2015, and recruitment is ongoing<sup>(19)</sup>.

In LIBRE, women with a history of breast cancer prior to study start as well as women without previous breast cancer were included. Inclusion criteria were female sex, age between 18 and 69 years, a pathogenic BRCA1/2 mutation and written informed consent. Exclusion criteria comprised, among others, a BMI below 15 kg/m<sup>2</sup>, and neoplastic diseases currently in treatment<sup>(13)</sup>.

Individuals from the intervention group (n = 33) received a structured lifestyle-intervention program, consisting of a three-month intensive phase with bi-weekly group classes on the Mediterranean diet as well as professionally guided sport training. The intensive phase was followed by a nine-month less intensive phase with monthly meetings. The control group (n =

35) were lectured once on the dietary recommendations of the German Nutrition Society (DGE) and once on the beneficial effects of regular physical activity on breast cancer incidence, prognosis, and recurrence at the beginning of the study. Study visits were at baseline, as well as 3 months (time point V1) and 12 months (time point V2) after baseline. Details on the enrolment, randomization, drop-outs, and available data for each time point are shown in the CONSORT flow chart in Supplementary Figure 1.

This study was conducted according to the guidelines laid down in the Declaration of Helsinki, and all procedures involving human subjects were approved by the ethics review board of the Klinikum Rechts der Isar of the Technical University of Munich (reference 5686/13). Written informed consent was obtained from all subjects. The trial was registered at [clinicaltrials.gov](https://clinicaltrials.gov) (reference: NCT02087592).

### ***Dietary measurements***

Two validated dietary questionnaires were used. The Mediterranean Diet Adherence Screener (MEDAS), developed in the Prevención con Dieta Mediterránea (PREDIMED) studies<sup>(20,21)</sup>. We calculated the MEDAS-Score as the percentage of the achieved score related to the achievable score (e.g. 7/14 = 50%; 7/13 = 54%).

In addition to the MEDAS, participants were asked to complete a 33-page long semi-quantitative Food Frequency Questionnaire (FFQ) established and validated by the European Prospective Investigation into Cancer and Nutrition (EPIC) consortium<sup>(22)</sup>. The EPIC-FFQ provides the daily intakes of food groups (e.g. fruits, vegetables, nuts) and nutrients (e.g. fats, carbohydrates, protein). Data from the EPIC-FFQ were adjusted for energy intake<sup>(23)</sup>. Since the EPIC-FFQ does not per se measure adherence to the Mediterranean diet, we calculated the Mediterranean Diet Score (MedD-Score), a commonly used score established by Trichopoulou et al.<sup>(24)</sup>.

Thus, in the present analyses, we included two independent scores which determine adherence to the Mediterranean diet, i.e. the MEDAS-Score and the MedD-Score. Dietary data for all variables and all time points is shown in Supplementary Table 1.

### ***Biological measurements***

The methodology to assess fecal fatty acids using gas chromatography<sup>(17)</sup> and plasma fatty acids using gas chromatography with mass spectrometry<sup>(12)</sup> has previously been described in detail. The proportion (%) of each fatty acid in the total fatty acid composition (= 100%) was

determined, and each fatty acid is shown as the proportion of the respective fatty acid. Fecal fatty acids were assessed in 2020 at the Université catholique de Louvain (Belgium).

The barrier biomarkers plasma LBP and fecal zonulin were assessed using enzyme-linked immunosorbent assays following the manufacturer's protocols (zonulin, REF K5600, Immundiagnostik AG, Bensheim, Germany; LBP, REFs DY870-05 & DY008, Bio-Techne GmbH, Wiesbaden, Germany).

### *Statistical analyses*

In this explorative analysis, we included all 68 participants from the completed LIBRE-1 study. The main focus of the present analysis was to assess possible changes in the fecal fatty acid composition, especially in the proportion of fecal long-chain *n*-3 PUFAs upon intervention. Considering the mean change in the summarized proportions of eicosapentaenoic acid (EPA), docosapentaenoic acid (DPA), and docosahexaenoic acid (DHA) in the fecal fatty acids between baseline and month 3, a post-hoc power calculation showed a power of 81% given an two-sided significance level of 5% (intervention group - 0.2% ± 0.7% [mean ± SD]; control group 0.2% ± 0.4%) (two-sided Wilcoxon-Mann-Whitney test; G\*Power software, Heinrich Heine University Duesseldorf, Germany).

As both gut barrier biomarkers and most fatty acids were not normally distributed (Shapiro-Wilk-Test), non-parametric tests were used. Differences between the intervention and control groups were tested using Fisher's exact test for categorical variables or Mann-Whitney U tests for quantitative data. Within-group differences over time were assessed using Wilcoxon matched-pairs signed rank tests. To assess changes over time, we calculated the shift for each parameter (BL values subtracted from the respective values at time points V1 and V2, shown as  $\Delta V1-BL$  and  $\Delta V2-BL$ ). Correlations were determined using Spearman's rank coefficient with shift values. A  $P < 0.08$  was considered as a trend, a  $P < 0.05$  was considered as statistically significant.

As these analyses are not confirmatory but to generate hypotheses to be further examined in larger studies, e.g. LIBRE-2, we did not perform post-hoc adjustment for multiple testing. To compensate for random findings in the correlation analyses, we would only draw conclusions based on results which can be found (i) consistently in both study arms and/or (ii) for both shifts  $\Delta V1-BL$  and  $\Delta V2-BL$ . Also, we only included those parameters in the correlation analyses which significantly changed during intervention. Statistical analyses were performed

using GraphPad Prism version 9.1.0 (GraphPad Software, San Diego, CA, USA). Data are shown as medians with interquartile ranges (25<sup>th</sup>;75<sup>th</sup> percentiles).

## RESULTS

### *Baseline characteristics*

At baseline, the intervention and control groups had similar numbers of women with previously diagnosed breast cancer, vegetarians, and smokers, and they did not differ in age, BMI (Table 1), and physical fitness (data not shown).

Both groups showed similar adherence to the Mediterranean diet according to the MedD-Score at baseline, yet the MEDAS-Score was slightly higher in the intervention group compared to the control group (50% versus 42%,  $P = 0.045$ ) (Supplementary Table 1). Besides the small difference in the MEDAS-Score, there was no baseline difference in diet. Fecal proportions of all fecal fatty acids assessed, as well as the levels of the intestinal barrier biomarkers LBP and zonulin did not differ between the groups at baseline (Table 2 and Supplementary Table 1).

### *Effect of the intervention on dietary habits and biomarkers of intestinal barrier integrity*

As described in detail before<sup>(12)</sup>, adherence to the Mediterranean diet increased markedly in the intervention group for at least one year ( $\Delta V1-BL$  and  $\Delta V2-BL$ , all  $P < 0.01$ ) (Supplementary Table 1). In the control group, there was a mild but significant increase in the adherence to the Mediterranean between baseline and V1 (all  $P < 0.01$ ), which was absent at V2.

Participants from the intervention group increased the intake of the typically Mediterranean food groups, especially the intake of nuts, fish and seafood, vegetables, legumes, fruits, olives, and vegetable oil ( $\Delta V1-BL$  and  $\Delta V2-BL$ , all  $P < 0.05$ ). At the same time, the intake of processed meat ( $P < 0.05$  for  $\Delta V1-BL$  and  $\Delta V2-BL$ ) and by trend red meat ( $P = 0.07$  for  $\Delta V1-BL$ ) decreased in the intervention group, but not in the control group.

Both gut barrier biomarkers decreased markedly in the intervention group (LBP and zonulin  $P < 0.01$  for  $\Delta V1-BL$  and  $\Delta V2-BL$ ), suggesting improved intestinal barrier integrity. In the control group, both biomarkers decreased in the first three months but returned to baseline levels after one year ( $P < 0.05$  for  $\Delta V1-BL$ ;  $P > 0.08$  for  $\Delta V2-BL$ ).

Diet data and data for the intestinal barrier biomarkers for all time points are shown in detail in Supplementary Table 1.



### ***Effect of the Mediterranean diet on the fecal fatty acid composition***

We observed several changes in the fecal fatty acid composition in both study arms. Interestingly, several fatty acids that changed during the study showed different directions in the two study arms. For example, the proportions of the saturated fatty acids palmitic acid (16:0, Figure 1a), stearic acid (18:0, Figure 1b) and arachidic acid (20:0) decreased in the intervention group while the proportions increased in the control group (Table 2). Also, the fecal proportion of the *n*-6 PUFA linoleic acid (18:2) increased in the intervention group and decreased in the control group (Figure 1d) (Table 2).

In the intervention group, but not the control group, there was a decrease in the proportion of the *cis n*-7 palmitoleic acid (16:1, Figure 1c, Table 2) and the proportion of the *n*-6 PUFA arachidonic acid (ARA, 20:4, Figure 1f, Table 2). Also, the proportion of the *n*-3 PUFA alpha-linoleic acid (ALA, 18:3) increased solely in the intervention group (Figure 1e, Table 2). There was no effect of the intervention on fecal proportions of the *n*-3 PUFAs EPA (20:5, *n*-3) and DHA (22:6, *n*-3) (Table 2).

Interestingly, there was a trend towards an increase in the fecal proportion of *trans* vaccenic acid (*t*18:1) in the intervention group ( $\Delta V1\text{-BL}$ ,  $P < 0.08$ ), a monounsaturated fatty derived from microbial metabolism of linoleic acids<sup>(17)</sup>.

### ***The fecal proportion of palmitoleic acid is associated with adherence to the Mediterranean diet and the gut barrier biomarker LBP***

To test for possible associations, we ran correlation analyses including dietary parameters, the proportion of fecal fatty acids, and the levels of the intestinal barrier biomarkers. For the correlation analyses, we used the shift values ( $\Delta V1\text{-BL}$  and  $\Delta V2\text{-BL}$ ) and included only parameters that were significantly altered during the study (see Table 2 and Supplementary Table 1).

In the intervention group, the decrease in the fecal proportion of the *n*-7 palmitoleic acid (*c*16:1) correlated with the decrease in the levels of plasma LBP (Figure 2a) and correlated inversely with adherence to the Mediterranean diet, assessed by the MedD-Score (Figure 2b) for both  $\Delta V1\text{-BL}$  and  $\Delta V2\text{-BL}$  (Table 3).

### ***The composition of fecal fatty acids is not associated with the composition of plasma fatty acids***



Lastly, to assess possible associations between the compositions of fecal fatty acids and plasma fatty acids, we performed correlation analyses. In the analyses we included the shift data ( $\Delta V1$ -BL and  $\Delta V2$ -BL) as well as the data for each time point (BL, V1, V2). We did not find any correlations between the proportion of fecal fatty acids and the proportion of plasma fatty acids that fit our selection criteria (correlation found in both groups and/or correlation found for both shifts and/or correlation found at two or more time points) (data not shown).

### ***Effect of the Mediterranean diet on the total amount of fecal fatty acids***

As recommended<sup>(25)</sup>, we report both the absolute and relative data of the fecal fatty acid composition. As shown in detail in Supplementary Table 2, the total amount of several fecal acids changed during intervention. Notably, the total amount of fecal fatty acids increased in the intervention group, but not the control group.

## **DISCUSSION**

In the present study we show that adherence to the Mediterranean diet induces several changes in the proportions of fecal fatty acids, especially in saturated fatty acids, but also in particular *n*-3 and *n*-6 PUFAs. Interestingly, even though we have recently confirmed findings from cell lines<sup>(26,27)</sup> and animals<sup>(28-30)</sup>, that diet-derived *n*-3 PUFAs, especially DHA, are associated with an improvement in biomarkers of intestinal barrier integrity<sup>(12)</sup>, we did not find an association between fecal *n*-3 PUFAs and biomarkers of intestinal barrier integrity in the present analyses.

### ***Adherence to the Mediterranean diet changed the fecal fatty acid composition***

Adherence to the Mediterranean diet induced several changes in the fecal proportions of long-chain fatty acids. Of interest, we did not see changes in the fecal proportions of the *n*-3 PUFAs DHA and EPA, commonly found in fish, even though the intake of fish increased in the intervention group. This finding is in line with data showing that after consuming different foods containing microencapsulated fish oil powder, only small amounts of *n*-3 PUFAs in ileal effluent<sup>(31)</sup>. Our study now confirms these findings, showing that the increased intake of fish and seafood observed in the LIBRE cohort did neither increase the fecal proportion of these *n*-3 PUFAs nor their total fecal amount. We assume that this finding is due to a very efficient intestinal absorption of *n*-3 PUFAs<sup>(32)</sup>.

We observed a decrease in the fecal proportion of the *n*-6 PUFA arachidonic acid (20:4). There is increasing evidence that eicosanoids derived from arachidonic acid have abilities to regulate the intestinal epithelial barrier, yet the underlying mechanisms and the physiological

relevance are not sufficiently understood<sup>(33)</sup>. Our data now indicate that the fecal proportion of arachidonic acid is not associated with impaired intestinal barrier integrity in humans. Though we base this finding on results from two validated intestinal barrier biomarkers, we suggest that future research in this field should assess intestinal barrier integrity by additional assessments, like the lactulose-mannitol-sucralose test.

We found a transient increase in the proportion of *trans* vaccenic acid (*t*18:1) in the intervention group in the first three months. This fatty acid, derived from microbial metabolism of linoleic acids, has previously been associated with the intake of specific dietary fibers, namely chitin-glucan, a branched  $\beta$ -1,3/1,6 glucan<sup>(17)</sup>. The findings from this previous study are in line with the present results, showing that the fecal proportion of *trans* vaccenic acid was also increasing in the intervention group in the first three months. Affirming this, we found an inverse correlation between the change in the fecal proportion of *trans* vaccenic acid and the *n*-3 PUFA alpha-linoleic acid (18:3) in the first three months ( $r = -0.44$ ,  $P = 0.01$ , Spearman correlation).

In the present study we focussed mainly on the fecal fatty acid composition expressed in percentage. This is due to the fact that fatty acid data expressed in percentage tends to exhibit lower inter-individual and intra-individual variability than absolute concentration and tends to be distributed normally, thereby increasing statistical power, especially when the total amount of fatty acids changes during the study course<sup>(25)</sup>. Furthermore, we chose to analyze our data using the fatty acid composition expressed in percentage to keep our results comparable with previous clinical trials investigating the effect of dietary fibers on fecal fatty acid patterns, which were expressed in percentage<sup>(16,17)</sup>.

### ***Fecal omega-3 fatty acids are not associated with biomarkers of intestinal barrier integrity***

Previously, *in vitro* studies<sup>(26,34,35)</sup> and studies in rodents<sup>(36–38)</sup> showed that *n*-3 PUFAs affect the tight junction proteins occludin and zonula occludens-1, which are essential for effective cell-cell connections, preventing uncontrolled paracellular permeability of e.g. lipopolysaccharides into the bloodstream, increasing the production of LBP<sup>(3)</sup>. Furthermore, it was shown that *n*-3 PUFAs induce the G-protein coupled receptor 120, which exerts anti-inflammatory effects and increases tight junction stability<sup>(39–41)</sup>, potentially decreasing uncontrolled paracellular permeability.

In contrast to plasma proportions<sup>(12)</sup>, the fecal proportion of *n*-3 PUFAs, were not associated with biomarkers of intestinal barrier integrity in the LIBRE cohort. We assume that this is due to the fact that *n*-3 PUFAs are absorbed throughout the passage via the digestive tract<sup>(32)</sup>.

Possibly, there might be beneficial effects of luminal *n*-3 PUFAs on intestinal barrier integrity in proximal parts of the intestinal tract. However, assessment of these fatty acids in fecal samples does not allow to assess this, as most of the *n*-3 PUFAs get absorbed along the digestive tract. Based on our findings, we propose that future studies should investigate the association between fecal fatty acids and intestinal barrier integrity using samples taken directly from the small intestine or proximal parts of the large intestine, e.g. by capsules<sup>(42)</sup>.

It has been recently shown *in vitro* that long-chain saturated fatty acids (esp. 16:0 and 18:0) increase intestinal permeability, presumably by inducing severe mitochondrial alterations in enterocytes, marked by a diminution of adenosine triphosphate production due to high proton leak, oxidative phosphorylation uncoupling, mitochondrial network remodeling, and reactive oxygen species generation<sup>(43)</sup>. Despite the finding that the fecal proportion of the two long-chain saturated fatty acids 16:0 and 18:0 decreased within the first three months in the present clinical study, there was no association between their fecal proportion and biomarkers of barrier function. This is in contrast to our previous findings, where we showed an association between the plasma proportion of long-chain saturated fatty acids and biomarkers of intestinal barrier integrity<sup>(12)</sup>. The reason for this difference is currently unknown.

### ***Fecal palmitoleic acid is associated with adherence to the Mediterranean diet and biomarkers of intestinal barrier integrity***

We observed a decrease in the fecal proportion of the *n*-7 monounsaturated *cis*-palmitoleic acid (*c*16:1). In humans, this fatty acid mainly originates from *de novo* lipogenesis mediated by stearoyl-CoA desaturase-1, the rate-limiting enzyme catalyzing the synthesis of monounsaturated fatty acids from saturated fatty acids. Consistent to findings from others<sup>(44,45)</sup>, we found an inverse association between palmitoleic acid and adherence to the Mediterranean diet. The role of palmitoleic acid in health and disease is not fully understood and is being discussed controversially, especially because previous results from studies in animals and humans partly showed opposing metabolic effects, as summarized by Frigolet and Gutiérrez-Aguilar<sup>(46)</sup>. In murine models, palmitoleic acid could effectively repair intestinal mucosal barrier<sup>(47)</sup>. In this study, the authors stated a potential role of the gut microbiota, declaring that palmitoleic acid rewired disrupted intestinal barrier and reprogrammed gut microbiota with selectively increasing the abundance of anti-inflammatory gut bacteria such as *Akkermansia muciniphila*<sup>(47)</sup>.

A study in high-fat diet-fed mice also showed that palmitoleic acid promotes intestinal tight junction integrity<sup>(48)</sup>. In detail, palmitoleic acid significantly increased the level of occludin

and claudin-1, two proteins that regulate the formation, maintenance, and function of tight junction in the intestine<sup>(48)</sup>. Furthermore, previous studies on metabolically active tissues have revealed that palmitoleic acid suppresses inflammatory cytokine production and attenuates inflammation<sup>(49,50)</sup>.

In contrast to these preclinical findings, our data indicate that a higher fecal proportion of palmitoleic acid is associated with higher levels of the gut barrier biomarker LBP, implying higher intestinal permeability. Here, further clinical studies are needed to investigate the association between this fatty acid and intestinal barrier integrity. These future studies should assess whether the opposing effects between preclinical studies (cell line, tissue, and animal studies) and clinical studies described by Frigolet and Gutiérrez-Aguilar<sup>(46)</sup> are also found in terms of intestinal barrier integrity in humans.

### ***Limitations and strengths***

A limitation of our study is that we only included women with BRCA mutations with associated mild intestinal barrier dysfunction. To what extent our finding will also hold true for other populations with more pronounced intestinal barrier dysfunction needs to be explored in future studies. In a validation study, both intestinal barrier biomarkers used in the present study were validated in healthy individuals without pathogenic BRCA germline mutations<sup>(5)</sup>. Also, fecal zonulin was validated as a well-suited barrier biomarker for overweight and obese individuals, but not for normal-weight individuals without barrier dysfunction<sup>(5)</sup>. These points need to be regarded, when interpreting the current findings, derived from normal weight women with impaired barrier function due to pathogenic BRCA germline mutations. Also, the approach of the present study is of an explorative nature, investigating outcome parameters which were not planned initially. The reasons why we still assessed these parameters are new developments in the field, which were not known at the time the LIBRE study was planned in 2013. For example, gut barrier biomarkers were validated for clinical use in 2021<sup>(5)</sup> and larger studies investigating the effect of dietary fiber on fecal LCFAs are from 2020 and 2021<sup>(16,17)</sup>. A strength of our study is the study design of a randomized controlled trial and a rigorous approach in the statistical analyses. To omit reporting random findings, we only show correlation results which were found consistently in the two study groups and/or found for more than one time point.

## **Conclusion**

To the best of our knowledge, this is the first clinical study investigating a potential association between fecal long-chain fatty acids and biomarkers of intestinal barrier integrity. Our findings show that there is only a small association between fecal long-chain fatty acids and biomarkers of intestinal barrier integrity, in contrast to what was previously shown for plasma fatty acids<sup>(12)</sup>.

In conclusion, our data show that adherence to the Mediterranean diet induces distinct changes in the fecal fatty acid composition. We showed that the fecal proportion of palmitoleic acid, a *n*-7 monounsaturated fatty acid, was associated with the intestinal barrier biomarker LBP. In contrast to *n*-3 PUFAs assessed in plasma, we found no association between fecal *n*-3 PUFAs and biomarkers of intestinal barrier integrity.

## **Acknowledgements**

We thank all participants and staff members involved in the LIBRE trial.

## **Financial support**

This study is part of the FiberTAG project ([www.fibertag.eu](http://www.fibertag.eu)) which was initiated from the European Joint Programming Initiative (JPI) “A Healthy Diet for a Healthy Life” ([www.healthydietforhealthylife.eu](http://www.healthydietforhealthylife.eu)). Funding for the team from the University of Hohenheim derived from the German Federal Ministry for Education and Research (BMBF; grant no. 01EA1701). Funding for the team from the Université catholique de Louvain derived from the Fonds de la Recherche Scientifique (FRS-FNRS) (PINT-MULTI R.8013.19, Belgium). NMD is a recipient of a grant from the Fonds de la Recherche Scientifique (FRS-FNRS) [PDR T.0068.19]. The LIBRE study was funded by the German Cancer Aid (Deutsche Krebshilfe; <http://www.krebshilfe.de>; grant no. 110013), Sphingotec GmbH, Senator Rösner Foundation, Marjan Miklus Foundation, and Waltraut Bergmann Foundation.

The funding sources had no role in the design, analysis or writing of this article.

## **Declarations of interests**

Declaration of interests: The authors declare none.

## Authorship

The authors' contributions were as follows. **MK, SCB**: designing the study and project administration (LIBRE); **NMD, SCB**: designing the study and project administration (FiberTAG); **BS, SCB**: formulating the research question; **BS, MB, AMN, WV**: carrying out the study; **BS**: data curation, formal analysis, analyzing the data; **BS, SCB**: interpreting the findings and writing the article; **SCB** had primary responsibility for the final content. **All authors** have read and approved the final manuscript.

## References

1. Martel J, Chang S-H, Ko Y-F, et al. (2022) Gut barrier disruption and chronic disease. *Trends Endocrinol. Metab.* **33**, 247–265.
2. Bischoff SC, Barbara G, Buurman W, et al. (2014) Intestinal permeability--a new target for disease prevention and therapy. *BMC Gastroenterol.* **14**, 189.
3. Bischoff SC, Kaden-Volynets V, Filipe Rosa L, et al. (2021) Regulation of the gut barrier by carbohydrates from diet - Underlying mechanisms and possible clinical implications. *Int. J. Med. Microbiol. IJMM* **311**, 151499.
4. Chantler S, Griffiths A, Matu J, et al. (2021) The Effects of Exercise on Indirect Markers of Gut Damage and Permeability: A Systematic Review and Meta-analysis. *Sports Med. Auckl. NZ* **51**, 113–124.
5. Seethaler B, Basrai M, Neyrinck AM, et al. (2021) Biomarkers for assessment of intestinal permeability in clinical practice. *Am. J. Physiol. Gastrointest. Liver Physiol.* **321**, G11–G17.
6. Seethaler B, Nguyen NK, Basrai M, et al. (2022) Short-chain fatty acids are key mediators of the favorable effects of the Mediterranean diet on intestinal barrier integrity: data from the randomized controlled LIBRE trial. *Am. J. Clin. Nutr.* **116**, 928–942.
7. Parada Venegas D, De la Fuente MK, Landskron G, et al. (2019) Short Chain Fatty Acids (SCFAs)-Mediated Gut Epithelial and Immune Regulation and Its Relevance for Inflammatory Bowel Diseases. *Front. Immunol.* **10**, 277.

8. Liu P, Wang Y, Yang G, et al. (2021) The role of short-chain fatty acids in intestinal barrier function, inflammation, oxidative stress, and colonic carcinogenesis. *Pharmacol. Res.* **165**, 105420.
9. Suzuki T, Yoshida S & Hara H (2008) Physiological concentrations of short-chain fatty acids immediately suppress colonic epithelial permeability. *Br. J. Nutr.* **100**, 297–305.
10. Bernardi S, Del Bo' C, Marino M, et al. (2020) Polyphenols and Intestinal Permeability: Rationale and Future Perspectives. *J. Agric. Food Chem.* **68**, 1816–1829.
11. Scott SA, Fu J & Chang PV (2020) Microbial tryptophan metabolites regulate gut barrier function via the aryl hydrocarbon receptor. *Proc. Natl. Acad. Sci. U. S. A.* **117**, 19376–19387.
12. Seethaler B, Lehnert K, Yahiaoui-Doktor M, et al. (2023) Omega-3 polyunsaturated fatty acids improve intestinal barrier integrity-albeit to a lesser degree than short-chain fatty acids: an exploratory analysis of the randomized controlled LIBRE trial. *Eur. J. Nutr.*
13. Kiechle M, Engel C, Berling A, et al. (2016) Lifestyle intervention in BRCA1/2 mutation carriers: study protocol for a prospective, randomized, controlled clinical feasibility trial (LIBRE-1 study). *Pilot Feasibility Stud.* **2:74**.
14. Voss MRH van, Diest PJ van, Smolders YHCM, et al. (2014) Distinct claudin expression characterizes BRCA1-related breast cancer. *Histopathology* **65**, 814–827.
15. Brennan K, Offiah G, McSherry EA, et al. (2010) Tight Junctions: A Barrier to the Initiation and Progression of Breast Cancer? *J. Biomed. Biotechnol.* **2010**, 460607.
16. Neyrinck AM, Rodriguez J, Zhang Z, et al. (2021) Prebiotic dietary fibre intervention improves fecal markers related to inflammation in obese patients: results from the Food4Gut randomized placebo-controlled trial. *Eur. J. Nutr.*
17. Rodriguez J, Neyrinck AM, Zhang Z, et al. (2020) Metabolite profiling reveals the interaction of chitin-glucan with the gut microbiota. *Gut Microbes* **12:1810530**, 1810530.

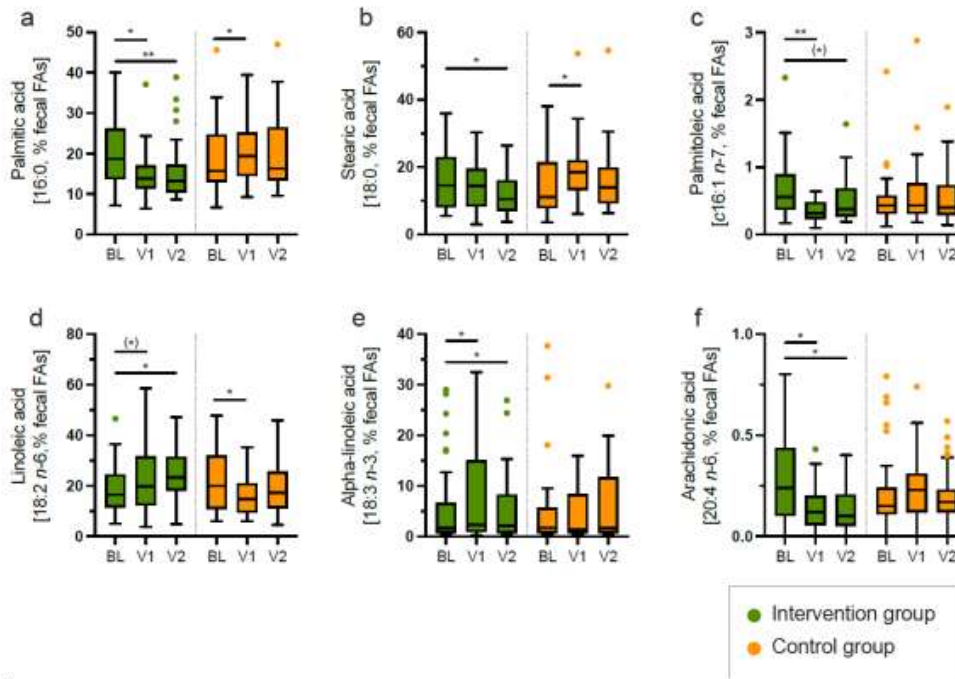


18. Donini LM, Serra-Majem L, Bulló M, et al. (2015) The Mediterranean diet: culture, health and science. *Br. J. Nutr.* **113**, S1–S3.
19. Kiechle M, Engel C, Berling A, et al. (2016) Effects of lifestyle intervention in BRCA1/2 mutation carriers on nutrition, BMI, and physical fitness (LIBRE study): study protocol for a randomized controlled trial. *Trials* **17**:368.
20. Schröder H, Fitó M, Estruch R, et al. (2011) A short screener is valid for assessing Mediterranean diet adherence among older Spanish men and women. *J. Nutr.* **141**, 1140–1145.
21. Hebestreit K, Yahiaoui-Doktor M, Engel C, et al. (2017) Validation of the German version of the Mediterranean Diet Adherence Screener (MEDAS) questionnaire. *BMC Cancer* **17**, 341.
22. Bohlscheid-Thomas S, Hoting I, Boeing H, et al. (1997) Reproducibility and relative validity of energy and macronutrient intake of a food frequency questionnaire developed for the German part of the EPIC project. European Prospective Investigation into Cancer and Nutrition. *Int. J. Epidemiol.* **26 Suppl 1**, S71-81.
23. Willett WC, Howe GR & Kushi LH (1997) Adjustment for total energy intake in epidemiologic studies. *Am. J. Clin. Nutr.* **65**, 1220S-1228S.
24. Trichopoulou A, Kouris-Blazos A, Wahlqvist ML, et al. (1995) Diet and overall survival in elderly people. *BMJ* **311**, 1457–1460.
25. Brenna JT, Plourde M, Stark KD, et al. (2018) Best practices for the design, laboratory analysis, and reporting of trials involving fatty acids. *Am. J. Clin. Nutr.* **108**, 211–227.
26. Xiao K, Liu C, Qin Q, et al. (2020) EPA and DHA attenuate deoxynivalenol-induced intestinal porcine epithelial cell injury and protect barrier function integrity by inhibiting necroptosis signaling pathway. *FASEB J. Off. Publ. Fed. Am. Soc. Exp. Biol.* **34**, 2483–2496.
27. Li E, Horn N & Ajuwon KM (2022) EPA and DHA inhibit endocytosis of claudin-4 and protect against deoxynivalenol-induced intestinal barrier dysfunction through PPAR $\gamma$

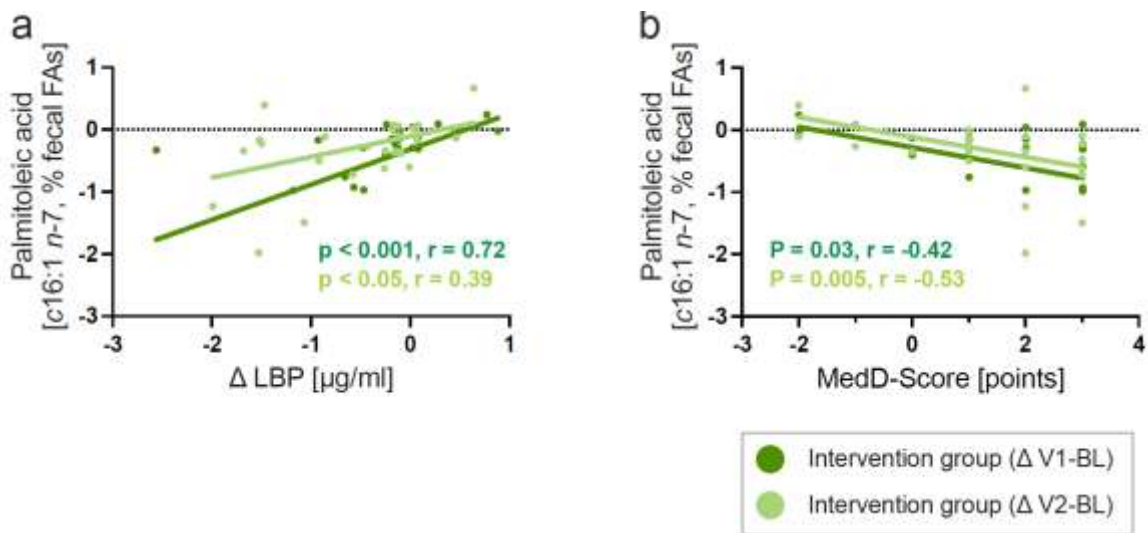
- dependent and independent pathways in jejunal IPEC-J2 cells. *Food Res. Int. Ott. Ont* **157**, 111420.
28. Fang J, Zhang Z, Cheng Y, et al. (2022) EPA and DHA differentially coordinate the crosstalk between host and gut microbiota and block DSS-induced colitis in mice by a reinforced colonic mucus barrier. *Food Funct.* **13**, 4399–4420.
  29. Du L, Hao Y-M, Yang Y-H, et al. (2022) DHA-Enriched Phospholipids and EPA-Enriched Phospholipids Alleviate Lipopolysaccharide-Induced Intestinal Barrier Injury in Mice via a Sirtuin 1-Dependent Mechanism. *J. Agric. Food Chem.* **70**, 2911–2922.
  30. Zhu H, Liu Y, Chen S, et al. (2016) Fish oil enhances intestinal barrier function and inhibits corticotropin-releasing hormone/corticotropin-releasing hormone receptor 1 signalling pathway in weaned pigs after lipopolysaccharide challenge. *Br. J. Nutr.* **115**, 1947–1957.
  31. Sanguansri L, Shen Z, Weerakkody R, et al. (2013) Omega-3 fatty acids in ileal effluent after consuming different foods containing microencapsulated fish oil powder - an ileostomy study. *Food Funct.* **4**, 74–82.
  32. Schuchardt JP & Hahn A (2013) Bioavailability of long-chain omega-3 fatty acids. *Prostaglandins Leukot. Essent. Fatty Acids* **89**, 1–8.
  33. Huang N, Wang M, Peng J, et al. (2021) Role of arachidonic acid-derived eicosanoids in intestinal innate immunity. *Crit. Rev. Food Sci. Nutr.* **61**, 2399–2410. Taylor & Francis.
  34. Li Q, Zhang Q, Wang M, et al. (2008) n-3 polyunsaturated fatty acids prevent disruption of epithelial barrier function induced by proinflammatory cytokines. *Mol. Immunol.* **45**, 1356–1365.
  35. Xiao G, Tang L, Yuan F, et al. (2013) Eicosapentaenoic acid enhances heat stress-impaired intestinal epithelial barrier function in Caco-2 cells. *PloS One* **8**, e73571.
  36. Hudert CA, Weylandt KH, Lu Y, et al. (2006) Transgenic mice rich in endogenous omega-3 fatty acids are protected from colitis. *Proc. Natl. Acad. Sci. U. S. A.* **103**, 11276–11281.

37. Chien Y-W, Peng H-C, Chen Y-L, et al. (2017) Different Dietary Proportions of Fish Oil Regulate Inflammatory Factors but Do Not Change Intestinal Tight Junction ZO-1 Expression in Ethanol-Fed Rats. *Mediators Inflamm.* **2017**, 5801768.
38. Xiao G, Yuan F, Geng Y, et al. (2015) EICOSAPENTAENOIC ACID ENHANCES HEATSTROKE-IMPAIRED INTESTINAL EPITHELIAL BARRIER FUNCTION IN RATS. *Shock Augusta Ga* **44**, 348–356.
39. Durkin LA, Childs CE & Calder PC (2021) Omega-3 Polyunsaturated Fatty Acids and the Intestinal Epithelium—A Review. *Foods* **10**, 199.
40. Rubbino F, Garlatti V, Garzarelli V, et al. (2022) GPR120 prevents colorectal adenocarcinoma progression by sustaining the mucosal barrier integrity. *Sci. Rep.* **12**, 381.
41. Mobraten K, Haug TM, Kleiveland CR, et al. (2013) Omega-3 and omega-6 PUFAs induce the same GPR120-mediated signalling events, but with different kinetics and intensity in Caco-2 cells. *Lipids Health Dis.* **12**, 101.
42. Rehan M, Al-Bahadly I, Thomas DG, et al. (2024) Smart capsules for sensing and sampling the gut: status, challenges and prospects. *Gut* **73**, 186–202.
43. Guerbette T, Rioux V, Bostoën M, et al. (2024) Saturated fatty acids differently affect mitochondrial function and the intestinal epithelial barrier depending on their chain length in the in vitro model of IPEC-J2 enterocytes. *Front. Cell Dev. Biol.* **12**, 1266842.
44. Giroli MG, Werba JP, Risé P, et al. (2021) Effects of Mediterranean Diet or Low-Fat Diet on Blood Fatty Acids in Patients with Coronary Heart Disease. A Randomized Intervention Study. *Nutrients* **13**, 2389.
45. Féart C, Torrès MJM, Samieri C, et al. (2011) Adherence to a Mediterranean diet and plasma fatty acids: data from the Bordeaux sample of the Three-City study. *Br. J. Nutr.* **106**, 149–158. Cambridge University Press.
46. Frigolet ME & Gutiérrez-Aguilar R (2017) The Role of the Novel Lipokine Palmitoleic Acid in Health and Disease. *Adv. Nutr.* **8**, 173S-181S.

47. Chen Y, Mai Q, Chen Z, et al. (2023) Dietary palmitoleic acid reprograms gut microbiota and improves biological therapy against colitis. *Gut Microbes* **15**, 2211501.
48. Liang Q, Zheng Y, Meng F, et al. (2024) Palmitoleic acid on top of HFD ameliorates insulin resistance independent of diacylglycerols and alters gut microbiota in C57BL/6J mice. *Food Sci. Hum. Wellness* **13**.
49. Cao H, Gerhold K, Mayers JR, et al. (2008) Identification of a Lipokine, a Lipid Hormone Linking Adipose Tissue to Systemic Metabolism. *Cell* **134**, 933–944.
50. Chan KL, Pilon NJ, Sivaloganathan DM, et al. (2015) Palmitoleate Reverses High Fat-induced Proinflammatory Macrophage Polarization via AMP-activated Protein Kinase (AMPK)\*. *J. Biol. Chem.* **290**, 16979–16988.



**Figure 1. Effect of the intervention on the fecal fatty acid composition (a-f).** Shown are data for baseline (BL), as well as after month 3 (V1) and month 12 (V2) for the proportion (%) of fecal fatty acids in the total fecal fatty acid composition (fecal FAs). Tukey boxplots with median, whiskers (1.5 x interquartile ranges), and outliers are shown in green (intervention group;  $n=33$ ) and orange (control group;  $n=35$ ). Within group difference to BL is indicated by asterisks ( $(^*)P < 0.08$ ;  $(^*)P < 0.05$ ;  $(^{**})P < 0.01$ ; Wilcoxon signed-rank test). This figure summarizes data shown in Table 2.



**Figure 2. Adherence to the Mediterranean diet as well as plasma levels of lipopolysaccharide binding protein (LBP) are associated with the fecal proportion of the *n*-7 monounsaturated palmitoleic acid.** Shown are the correlations between the proportion (%) of palmitoleic acid in the total fecal fatty acid composition (fecal FAs) and the adherence to the Mediterranean diet (assessed by the Mediterranean diet Score [MedD-Score]) (*a*) and the proportion of palmitoleic acid and the plasma levels of the gut permeability biomarker LBP (*b*). Spearman correlations were conducted using shift values (baseline [BL] values subtracted from the respective values after month 3 [V1] and month 12 [V2]). This figure summarizes the findings shown in Table 3.

**Table 1.** Patient characteristics at baseline.

Parameters	Intervention group	Control group	P value <sup>1</sup>
	<i>n</i> = 33	<i>n</i> = 35	
Diseased <sup>2</sup> [ <i>n</i> (%)]	23 (69.7)	23 (65.7)	0.80
Vegetarians [ <i>n</i> (%)]	2 (6.1)	4 (11.4)	0.67
Smokers [ <i>n</i> (%)]	4 (12.1)	4 (11.4)	0.99
Age [years]	42 (35;49)	41 (35;50)	0.84
BMI [kg/m <sup>2</sup> ]	23 (21;28)	24 (21;28)	0.49

<sup>1</sup>Between group difference. <sup>2</sup>Previously diagnosed with breast cancer. Total numbers and percentage (diseased, vegetarians, smokers) or median and interquartile ranges (age, BMI) are shown. Abbreviations: BMI, body mass index. Statistics: Fisher's exact test (categorical variables) and Mann-Whitney *U* test (numerical data).



**Table 2. Changes in the fecal fatty acid composition (%) during the study.** Shown are the data for baseline (BL), for the shift between BL and month 3 ( $\Delta$  V1-BL), and for the shift between BL and month 12 ( $\Delta$  V2-BL).

	Intervention group (n = 33)			Control group (n = 35)			Intervention vs. control groups (P)	
	BL	$\Delta$ V1-BL	$\Delta$ V2-BL	BL	$\Delta$ V1-BL	$\Delta$ V2-BL	$\Delta$ V1-BL	$\Delta$ V2-BL
<b>Fatty acids [%]<sup>1</sup></b>								
16:0 (Palmitic acid)	19 (14;26)	<b>-1.4 (-11;-0.7)*</b>	<b>-3.3 (-15;2.0)**</b>	16 (13;25)	<b>2.5 (-1.0;5.2)*</b>	0.6 (-3.1;4.6)	<b>&lt; 0.001</b>	<b>0.02</b>
16:1	0.2 (0.1;0.3)	<b>-0.1 (-0.1;0.0)**</b>	0.1 (-0.2;0.1)	0.2 (0.1;0.3)	<b>0.1 (0.1;0.2)*</b>	0.0 (-0.1;0.1)	<b>&lt; 0.001</b>	0.56
16:1 (Palmitoleic acid, <i>n</i> -7)	0.6 (0.4;0.9)	<b>-0.2 (-0.5;0.0)**</b>	<b>-0.1 (-0.4;0.1)*</b>	0.4 (0.3;0.6)	0.1 (-0.1;0.3)	0.0 (-0.1;0.1)	<b>&lt; 0.001</b>	<b>0.03</b>
18:0 (Stearic acid)	15 (8.1;23)	-0.9 (-6.3;2.3)	<b>-5.9 (-12;0.5)*</b>	11 (7.9;22)	<b>2.6 (-0.7;9.1)*</b>	1.5 (-2.5;4.8)	<b>0.01</b>	<b>0.01</b>
18:1 (Elaidic acid)	0.3 (0.2;0.5)	0.1 (-0.1;0.4)	0.1 (-0.2;0.3)	0.3 (0.2;0.6)	0.0 (-0.2;0.3)	0.0 (-0.3;0.1)	0.64	0.64
18:1 (Vaccenic acid)	3.0 (2.2;5.9)	<b>0.9 (-1.5;4.3)*</b>	-0.5 (-3.0;2.7)	4.0 (2.2;7.9)	0.2 (-1.5;2.4)	-0.3 (-2.4;3.6)	0.42	0.66
18:1 (Oleic acid, <i>n</i> -9)	25 (19;37)	-1.4 (-13;11)	3.4 (-5.9;15.8)	27 (18;36)	-1.8 (-9.8;4.8)	3.1 (-10;11.8)	0.53	0.35
18:1 <sup>C11</sup>	1.3 (1.1;1.7)	-0.2 (-0.5;0.3)	0.1 (-0.5;0.4)	1.2 (1.1;1.5)	0.0 (-0.3;0.3)	0.1 (-0.3;0.4)	0.43	0.62
18:2 (Linoleic acid, <i>n</i> -	16 (11;25)	<b>5.9 (-6.7;15)*</b>	<b>8.7 (-2.4;17)*</b>	20 (11;32)	<b>-4.2 (-15;2.5)*</b>	-0.3 (-13;4.8)	<b>0.01</b>	<

6)								<b>0.001</b>
18:2 (Rumenic acid)	0.6 (0.2;1.0)	0.3 (-0.2;0.8)	0.1 (-0.6;0.8)	0.6 (0.3;1.0)	0.1 (-0.4;0.7)	0.1 (-0.5;0.5)	0.77	0.9
18:2 <sup>T10C12</sup>	0.1 (0.1;0.1)	0.1 (-0.1;0.1)	0.1 (-0.1;0.1)	0.1 (0.0;0.1)	0.0 (-0.1;0.1)	0.0 (-0.1;0.1)	0.19	0.92
18:2 <sup>C9C11</sup>	0.1 (0.1;0.2)	0.1 (0.1;0.2)	0.1 (-0.1;0.1)	0.1 (0.0;0.1)	0.0 (-0.1;0.1)	0.0 (-0.1;0.1)	0.39	0.52
18:2 <sup>T11T13</sup>	0.1 (0.1;0.1)	-0.1 (-0.1;0.1)	<b>-0.1 (-0.1;0.0)*</b>	0.1 (0.0;0.1)	<b>0.1 (0.0;0.1)*</b>	<b>0.1 (0.0;0.1)*</b>	<b>0.02</b>	<b>0.01</b>
18:2 <sup>T9T11</sup>	0.1 (0.1;0.2)	0.1 (0.0;0.1)	0.1 (-0.1;0.1)	0.1 (0.1;0.1)	0.1 (0.0;0.1)	0.0 (0.0;0.1)	0.08	0.79
18:3 (ALA, <i>n</i> -3)	1.7 (0.7;6.6)	<b>1.5 (-4.0;4.7)*</b>	<b>1.1 (-4;3.5)*</b>	1.7 (0.7;5.7)	0.1 (-3.5;5.9)	0.3 (-1.6;7.5)	0.64	0.44
18:3 <sup>C8T10C12</sup>	0.4 (0.2;0.6)	<b>-0.1 (-0.3;0.0)**</b>	-0.1 (-0.2;0.1)	0.3 (0.2;0.5)	0.1 (-0.2;0.3)	0.0 (-0.2;0.1)	<b>0.01</b>	0.65
18:3 <sup>C9T11C13</sup>	0.2 (0.1;0.3)	<b>-0.1 (-0.2;0.0)***</b>	<b>-0.1 (-0.1;0.0)*</b>	0.1 (0.1;0.2)	<b>0.1 (0.0;0.2)*</b>	0.0 (-0.1;0.1)	<b>&lt; 0.001</b>	<b>0.05</b>
18:3 <sup>T8T10C12</sup>	0.1 (0.1;0.2)	0.0 (0.0;0.1)	0.0 (0.0;0.1)	0.0 (0.0;0.1)	0.0 (0.0;0.1)	0.0 (0.0;0.1)	0.89	1
18:3 <sup>T9T11T13</sup>	0.1 (0.0;0.1)	0.0 (-0.1;0.0)	0.0 (-0.1;0.0)	0.1 (0.0;0.1)	0.0 (-0.1;0.0)	0.0 (-0.1;0.0)	0.76	0.56
20:0 (Arachidic acid)	0.8 (0.5;1.0)	<b>-0.2 (-0.3;0.1)*</b>	<b>-0.1 (-0.5;0.1)*</b>	0.6 (0.5;0.9)	<b>0.2 (-0.2;0.5)*</b>	0.0 (-0.2;0.2)	<b>&lt; 0.001</b>	0.09

Accepted manuscript

22:0 (Behenic acid)	0.6 (0.3;0.8)	-0.1 (-0.3;0.1)	-0.1 (-0.4;0.2)	0.6 (0.4;0.8)	<b>0.1 (-0.1;0.5)*</b>	-0.1 (-0.3;0.2)	<b>0.01</b>	0.85
20:4 (ARA, <i>n</i> -6)	0.2 (0.1;0.4)	<b>-0.1 (-0.2;0.0)*</b>	<b>-0.1 (-0.3;0.0)*</b>	0.2 (0.1;0.2)	0.0 (0.0;0.2)	0.0 (0.0;0.1)	<b>&lt; 0.001</b>	<b>&lt; 0.001</b>
20:5 (EPA, <i>n</i> -3)	0.1 (0.1;0.2)	-0.1 (-0.1;0.0)	-0.1 (-0.1;0.0)	0.1 (0.0;0.1)	0.0 (-0.1;0.1)	0.0 (-0.1;0.1)	0.3	0.44
22:5 (DPA, <i>n</i> -3)	0.2 (0.1;0.4)	<b>-0.1 (-0.2;0.0)***</b>	<b>0.1 (-0.3;0.0)(*)</b>	0.2 (0.1;0.3)	0.0 (-0.1;0.2)	0.0 (-0.1;0.1)	<b>&lt; 0.001</b>	0.15
22:6 (DHA, <i>n</i> -3)	0.2 (0.1;0.4)	-0.1 (-0.2;0.0)	-0.1 (-0.2;0.1)	0.2 (0.1;0.2)	0.1 (-0.1;0.2)	0.1 (0.0;0.1)	0.11	<b>0.04</b>

Median and interquartile ranges (25<sup>th</sup>;75<sup>th</sup> percentiles) are shown. Difference between the study groups at baseline and difference between the group's shifts was tested using the Mann-Whitney *U* test. There was no between-group difference at baseline. Within-group difference between baseline and V1/V2 was tested using the Wilcoxon signed-rank test (<sup>(\*)</sup>*P* < 0.08; \**P* < 0.05; \*\**P* < 0.01; \*\*\**P* < 0.001). Abbreviations: ALA, alpha-linoleic acid; ARA, arachidonic acid; EPA, eicosapentaenoic acid; DPA, docosapentaenoic acid; DHA, docosahexaenoic acid. <sup>1</sup>Numbers denote double bond positions whereas C and T denote *cis*- and *trans*-configuration, respectively.

**Table 3. Correlation analyses between the shifts in diet, the proportion of fecal fatty acids (FAs), and intestinal barrier biomarkers.** Shown are the data for the shift between baseline (BL) and month 3 ( $\Delta$  V1-BL) and for the shift between BL and month 12 ( $\Delta$  V2-BL). Correlations were only performed with parameters which changed during the study (see Table 2 and Supplementary Table 1).

1. Variable $\Delta$	2. Variable $\Delta$	Intervention group	
		$\Delta$ V1-BL r (R <sup>2</sup> ), P value	$\Delta$ V2-BL r (R <sup>2</sup> ), P value
16:1 <sup>C9</sup> (Palmitoleic acid) [% total fecal acid]	MedD-Score [Points]	-0.42 (0.18) P = 0.03	-0.53 (0.28) P = 0.005
16:1 <sup>C9</sup> (Palmitoleic acid)	LBP [ $\mu$ g/ml]	0.72 (0.52) P < 0.001	0.39 (0.15) P < 0.05

Statistics: Spearman correlation. Abbreviations: LBP, lipopolysaccharide binding protein; MedD-Score, Mediterranean Diet Score. We only show correlations which were found (i) consistently for the intervention group and the control group and/or (ii) consistently for the two shifts  $\Delta$ V1-BL and  $\Delta$ V2-BL. There were no significant results in the control group for  $\Delta$  V1-BL and  $\Delta$  V2-BL.